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1 PREDICTION OF BIOCONCENTRATION FACTORS IN FISH AND

2 INVERTEBRATES USING MACHINE LEARNING

- 3 Thomas H. Miller^{a*}, Matteo D. Gallidabino^b, James R. MacRae^c, Stewart F. Owen^d,
- 4 Nicolas R. Bury^{ef}, Leon P. Barron^{a*}
- 5
- ⁶ ^aDepartment of Analytical, Environmental & Forensic Sciences, School of Population
- 7 Health & Environmental Sciences, Faculty of Life Sciences and Medicine, King's
- 8 College London, 150 Stamford Street, London, SE1 9NH, UK.
- ⁹ ^bDepartment of Applied Sciences, Northumbria University, Newcastle Upon Tyne, NE1
- 10 8ST, UK.
- ¹¹ ^cMetabolomics Laboratory, The Francis Crick Institute, 1 Midland Road, London, NW1
- 12 1AT, UK.
- ¹³ ^dAstraZeneca, Global Environment, Alderley Park, Macclesfield, Cheshire SK10 4TF,
 ¹⁴ UK.
- ¹⁵ ^eDivision of Diabetes and Nutritional Sciences, Faculty of Life Sciences and Medicine,
- 16 King's College London, Franklin Wilkins Building, 150 Stamford Street, London, SE1
- 17 9NH, UK.
- ¹⁸ ^fFaculty of Science, Health and Technology, University of Suffolk, James Hehir
 ¹⁹ Building, University Avenue, Ipswich, Suffolk, IP3 0FS, UK.
- 20
- 21 *Corresponding authors
- Email: thomas.miller@kcl.ac.uk (Tel: +44 20 7848 4978) or leon.barron@kcl.ac.uk;
 (Tel.: +44 20 7848 3842)
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GRAPHICAL ABSTRACT



30 Abstract

The application of machine learning has recently gained interest from ecotoxicological 31 fields for its ability to model and predict chemical and/or biological processes, such as 32 the prediction of bioconcentration. However, comparison of different models and the 33 prediction of bioconcentration in invertebrates has not been previously evaluated. A 34 comparison of 24 linear and machine learning models is presented herein for the 35 prediction of bioconcentration in fish and important factors that influenced 36 accumulation identified. R² and root mean square error (RMSE) for the test data (n = 37 38 110 cases) ranged from 0.23 – 0.73 and 0.34 – 1.20, respectively. Model performance was critically assessed with neural networks and tree-based learners showing the best 39 performance. An optimised 4-layer multi-layer perceptron (14 descriptors) was 40 selected for further testing. The model was applied for cross-species prediction of 41 bioconcentration in a freshwater invertebrate, Gammarus pulex. The model for G. 42 *pulex* showed good performance with R² of 0.99 and 0.93 for the verification and test 43 data, respectively. Important molecular descriptors determined to influence 44 bioconcentration were molecular mass (MW), octanol-water distribution coefficient 45 (logD), topological polar surface area (TPSA) and number of nitrogen atoms (nN) 46 among others. Modelling of hazard criteria such as PBT, showed potential to replace 47 the need for animal testing. However, the use of machine learning models in the 48 regulatory context has been minimal to date and is critically discussed herein. The 49 movement away from experimental estimations of accumulation to in silico modelling 50 would enable rapid prioritisation of contaminants that may pose a risk to environmental 51 health and the food chain. 52

Keywords modelling, PBT, pharmaceutical, bioconcentration, BCF, machine
learning

55 Introduction

Both terrestrial and aquatic environments experience pollution from a wide range of chemical contaminants. The presence of these contaminants is a cause for concern as they may elicit adverse effects to environmental and public health. Bioaccumulation of chemicals is critically important for understanding the risk of chemicals in the environment. The complexity of confounding factors that affect uptake make simple relationships that can confidently predict the accumulation elusive; but it may not have to be that way.

63 Live animal exposure studies are currently the norm, using many hundreds of fish for each assessment [1]. Across the European Union (EU), various guidelines 64 have been established for industry to minimise the risk posed by their chemical 65 products. For pharmaceuticals in the EU this is regulated by the European Medicines 66 Agency (EMA) and for other chemicals substances the regulations are outlined by the 67 Registration, Evaluation, Authorisation and restriction of CHemicals (REACH) [2, 3]. 68 According to REACH, any manufacturer of a chemical that exceeds quantities of 10 69 tonnes per annum must submit a chemical safety assessment (CSA). For 70 environmental risk assessment, part of the CSA includes 71 persistence, bioaccumulation and toxicity (PBT) assessments. Alternatively, for pharmaceuticals 72 environmental risk assessment (ERA) follows an initial screening (Phase I) where 73 74 physico-chemical properties of the compound are determined (e.g. logP) and the expected exposure is estimated. The Phase I exposure estimation is calculated as the 75 predicted environmental concentration (PEC). If the PEC is $>0.01 \ \mu g \ L^{-1}$ then the 76 pharmaceutical must undergo further testing to assess environmental fate and toxicity. 77 However, it should be noted that substances with a logP >4.5, will trigger a PBT 78 assessment (following REACH guidelines) regardless of the Phase I PEC. 79

For PBT assessments, existing available screening data and prior assessment 80 information are used to determine whether a chemical is bioaccumulative (B) or very 81 bioaccumulative (vB) by estimation of a bioconcentration factor (BCF) or 82 bioaccumulation factor (BAF). Currently, pharmaceuticals are not restricted or 83 replaced as would normally be defined under REACH. Furthermore, whilst PBT 84 assessments are implemented, the persistence and bioaccumulation outcome of 85 86 these assessments are not taken into consideration for authorisation purposes, as no legal provisions specifically cover persistent, bioaccumulative and toxic substances 87 88 for pharmaceuticals [4].

Laboratory testing for PBT brings with it a significant level of planning, quality 89 control and cost [1]. Therefore, in silico methodologies to predict BCF or BAF offers a 90 potential advantage to more intelligently use data to characterise potential exposure 91 and risk. Quantitative Structure Activity Relationships (QSARs) are becoming 92 increasingly popular within ecotoxicological fields as they represent, perhaps, the only 93 realistically feasible scenario to assess the environmental risk of the several thousand 94 chemicals that are available on the market [5]. In addition, such models can be used 95 to ethically reduce or replace animal testing and falls under the replacement, reduction 96 and refinement (3Rs) framework [6]. Further, effective in silico models could also be 97 utilised to help shape future drugs in terms of 'green by design' ambitions [7]. 98

More recently, more complex machine learning-based QSAR models involving artificial neural networks (ANNs), tree-based learners or support vector machines (SVMs) have been used to model BCF in fish [8-11]. However, several variations of machine learning-type models exist and wider applications of such models for bioaccumulation prediction have not yet been evaluated to identify any added benefits. Furthermore, current QSAR models have only been applied to modelling fish

bioaccumulation data and do not incorporate pharmaceutical data. The potential for
 application to other taxa such as invertebrates is also non-existent, mainly due to a
 shortage of available data.

The aim of this work was to develop and critically evaluate several machine 108 learning-based modelling tools for prediction of bioconcentration factor (BCF) in both 109 a fish (Cyprinus carpio) and an invertebrate species (Gammarus pulex) for the first 110 time. An open access fish BCF dataset was used in the first instance to build and 111 compare 24 different models for 352 different compounds. Subsequently, the best 112 113 model was applied to both a set of fish and invertebrate BCF data to assess its potential for cross-species prediction. The invertebrate dataset also contained mainly 114 pharmaceuticals. In parallel, independent models were developed ab initio on a 115 smaller set of invertebrate BCF data alone to assess the degree of commonality with 116 the model developed on fish BCF data. Finally, the importance of molecular 117 descriptors to understand the potential for a chemical to accumulate in biota was 118 assessed. The use of such rapid and flexible modelling approaches is now critical to 119 support the 3Rs, aid greener design and to help meet the demand for PBT 120 assessments of potentially large numbers of compounds, which could be expanded to 121 new and emerging environmental contaminants across different species. 122

123

124 Materials and Methods

125 Dataset generation and pre-processing

Bioconcentration factors were collated from the European Chemical Industry Council Long-range Research Initiative (Cefic LRI) project EC07 in collaboration with European Academy for Standardisation e.V (EURAS) which established the BCF gold standard database across multiple fish species and is freely available at

http://ambit.sourceforge.net/euras/. BCFs were down-selected to reduce variability 130 between different species and experimental conditions within the database. The BCF 131 data used herein were specific to C. carpio and were included by the Chemicals 132 Inspection and Testing Institute [12]. Out of all BCF data, this sub-selection resulted 133 in the largest dataset with a single fish species (n=352) for modelling purposes. The 134 reported BCFs represented whole-body values only and included pigments, 135 136 pesticides, fungicides, herbicides, insecticides, polyaromatic hydrocarbons (PAHs) and polychlorinated biphenyls (PCBs), organochlorines, nitroaromatics, alkylphenols, 137 138 aromatic hydrocarbons, organosulfurs and organotins. Approximately 36 % of the dataset contained ionisable compounds (estimated from ACD labs, Percepta 139 software). The invertebrate BCF dataset (n=34) was collated from literature reported 140 data [13-17] for the benthic freshwater organism, G. pulex. This species was selected 141 as there was a relatively large amount of BCF data available when compared with 142 other invertebrate species. For these, BCF data were only available for 143 pharmaceuticals and pesticides and, again, represented whole-body values. 144

Simplified molecular input line entry system (SMILES) strings were generated 145 for each compound using Chemspider (Royal Society of Chemistry, UK). Molecular 146 descriptors were generated from SMILES strings using Parameter Client (Virtual 147 Computational Chemistry Laboratory, Munich, Germany), and ACD Labs Percepta 148 149 (Advanced Chemistry Development Laboratories, ON, Canada). Approximately 450 descriptors were initially generated covering constitutional, topological, geometrical 150 and physico-chemical properties. The fish and invertebrate datasets were pre-151 processed to remove any zero variance descriptors or descriptors that were 152 erroneous. All BCF data used for modelling was log transformed for improved 153 predictive accuracy. 154

155

156 Feature selection

Descriptors were down-selected using three different feature selection 157 algorithms, the first of which was a genetic algorithm (GA). The GA parameters were 158 set to population = 500, generations = 250, mutation rate = 0.1 and cross-over rate = 159 1. The remaining two selection methods were part of stepwise regression which 160 included a forward selection algorithm (FA) and backwards selection algorithm (BA). 161 The feature selection algorithms used a generalised regression neural networks 162 163 (GRNN) to monitor the error associated with the selected descriptors, where descriptor sets were optimised when the error showed no improvement. The use of GRNN for 164 descriptor selection is very fast and requires minimal processing power. The 165 performance of each feature selection algorithm was characterised by then testing 166 several thousand neural networks and evaluating the predictive performance of the 167 models based on the error of the predictions. The best feature selection method was 168 the GA, which resulted in the down-selection of descriptors to a total of 14 that included 169 6 topological descriptors; radial centric information index (ICR), Narumi harmonic 170 topological function (Hnar), ramification index (Ram), superpendentic index (SPI), 171 spanning tree number (STN), topological polar surface area (TPSA), 4 constitutional 172 descriptors; number of hydrogens (nH), number of carbons (nC), number of nitrogens 173 174 (nN), molecular weight (MW), 3 electrotopological descriptors; maximal electrotopological negative variation (MAXDN), maximal electrotopological positive 175 variation (MAXDP), mean atomic Sanderson electronegativity (Me) and 1 physico-176 chemical property; the octanol-water distribution coefficient (logD) (See SI, Table S3). 177 178

179 Modelling approaches

Two different software packages were used to assess the applicability of 180 several in silico models in predicting bioconcentration. Trajan 6.0 (Trajan Software 181 Ltd., Lincolnshire, UK) was used to build and evaluate artificial neural networks. In 182 addition, this software was also used for the feature selection and the same 183 descriptors were used in both modelling software packages. Models developed and 184 optimised in Trajan included generalised regression neural networks (GRNN), radial 185 basis function networks (RBF) and 3-/4-layer multilayer perceptrons (MLP). Training 186 of the MLPs used two training algorithms referred to as back propagation (BP) and 187 188 conjugate gradient descent (CGD), models were trained for 100 iterations. The optimised model was a four-layer MLP. The first and fourth layers were the inputs 189 (molecular descriptors) and outputs (logBCF), respectively. The second and third 190 layers (hidden layers) contained 14 and 10 nodes, respectively. Regularisation was 191 performed with the use of early stopping to prevent over-training of the dataset. 192 Parameter tuning was performed by changing the number of hidden layers and nodes 193 and assessing the model performance on the verification and test subsets. The 194 subsets of cases presented to the neural networks were split so that 242 compounds 195 (70%) were used for training, 55 compounds (15%) for verification and 55 compounds 196 (15 %) for testing the networks. Normalisation of the input features showed no 197 improvement in performance of the networks and training was performed without 198 199 centred or scaled descriptors.

In the second software package, modelling was performed using the R statistical computing language (freely available from https://www.r-project.org). Here, predictive models from different kinds of learner categories including both linear and non-linear models were trained and tested. These included, ordinary leastsquares regression (OLM, package: *stats*), partial least-squares (PLS, package: *pls*),

205 ridge regression (RR, package: elasticnet), elastic net (EN, package: elasticnet), quantile regression with LASSO penalty (QRL, package: rgPen) multivariate adaptive 206 regression splines (MARS & B-MARS, package: earth), k-nearest neighbours 207 regression (KNN, package: caret), extreme learning machines (ELM, package: 208 elmNN), support vector machines with radial basis function (SVM-R, package: 209 kernlab) and polynomial (SVM-P, package: kernlab) kernels, random forest exploiting 210 classification and regression trees (RF-CART, package: randomForest) and 211 conditional inference trees (RF-CIT, package: party) algorithms as base learners, 212 boosted trees (BT, package: gbm) and Cubist regression (CR, package: Cubist). MLPs 213 (3-5 layers) with 1 hidden layer (ANN-1HL, package: *nnet*), averaged 1 hidden layer 214 (ANN-a1HL, package: nnet), 2 hidden layers (ANN-2HL, package: RSNNS) and 3 215 216 hidden layers (ANN-3HL, package: RSNNS) were also tested. For this modelling approach, the same molecular descriptors and logBCF were used again as input and 217 output variables. The dataset was split into two subsets, training data (70 %) and test 218 data (30 %). Normalisation of the data was required for the modelling application and 219 the dataset was both centred and scaled. Parameter tuning was performed by 220 resampling of the training subset following a 10-fold cross-validation scheme repeated 221 five times and implemented through the *caret* package. Performance of each model 222 was assessed from the root-mean square error (RMSE) and the correlation coefficient 223 224 (R²). The best model for each regression method was then selected, retrained on the entire training dataset and used to predict cases in the test dataset. Final datasets 225 used for modelling the optimised models are given in the SI (Table S1 & S2). The 226 227 finalised models were all tested according to OECD guidelines [18] for QSAR model validation. 228

229

230 Results and Discussion

231

232 Down-selection of input features for modelling BCFs in fish

The down-selection of the input features was assessed using three different 233 feature-selection algorithms. Stepwise methods that included forwards or backwards 234 selection (FA/BA) reduced the number of descriptors from 180 down to 72, whilst the 235 GA reduced the number of descriptors to 66. The GA showed better correlation 236 between selected descriptors with logBCF compared to stepwise algorithms (Figure 237 238 S1). For both BA and FA, the selection process converged to the same local minima indicating that there was no difference in using either algorithm. The improved 239 performance of the GA is due to selection of descriptors from multiple points in the 240 descriptor space, as opposed to FA or BA that start selection from a single point. Thus, 241 approaching global minima is more likely to arise when using the GA over stepwise 242 selection methods. 243

From the 66 descriptors selected by the GA, the top 22 descriptors plus an 244 additional two user curated descriptors were selected for further modelling (See SI, 245 Table S3). These additional descriptors were logD and number of hydrogen acceptor 246 groups (nHAcc) and were chosen for their previously demonstrated influence on 247 accumulation in biota [19, 20]. All descriptors were then tested across several 248 thousand MLPs (three and four-layer) where the Trajan software sub-selected the best 249 from the group of 24 descriptors based on model performance (MLPs yielded the best 250 performance over other model types in terms of R² and RMSE). The descriptors were 251 down-selected to a total of 14 that showed relatively good performance across MLPs 252 tested and were subsequently used in both modelling approaches discussed herein 253 (Table S3). Given the scale of BCF data used for training (n=242), the 5:1 Topliss 254

threshold set out by the OECD guidelines [18] for the ratio of numbers of cases todescriptors was acceptable at 17:1.

257

258 Comparison of model performances for prediction of fish BCFs

The results of both modelling approaches are shown in Table 1. For models 259 trained in R, the highest RMSE values were observed for OLM (1.203), followed by 260 PLS (1.164) and then QRL (1.112). The relatively poor performance of such linear 261 models may be expected as modelling such a biologically complex process is not likely 262 263 to follow linear relationships using simple molecular descriptors. Even with wellstudied descriptors, such as logP, there is a non-linear trend with accumulation over 264 a specific threshold (generally, logP >6) [21]. However, when used as a sole 265 descriptor, logP may exclude processes that are also important for accumulation. For 266 example, elimination and metabolism rates may impact net accumulation as well as 267 more specific physiology such as carrier mediated transport and protein binding [22] 268 will also influence accumulation, especially for emerging contaminant classes such as 269 pharmaceuticals. By comparison, better performance was achieved using higher 270 complexity models. The lowest RMSEs were observed for RF-CART (0.771), followed 271 by BT (0.789) and RF-CIT (0.821), i.e. three tree-based machine learners. Next, ANNs 272 and SVMs performed very similarly to tree learners, e.g. SVM-R (0.841), ANN-a1HL 273 274 (0.859) and ANN-3HL (0.880).

275 Models tested in Trajan showed particularly good performance, in comparison 276 to those built in R. The lowest RMSE value was observed for a 4-layer MLP (0.524), 277 followed by 3-layer MLP (0.538), RBF (0.689), GRNN (0.893) and Linear (1.052). In 278 absolute terms, definitive conclusions cannot be drawn from direct comparison of 279 modelling approaches (i.e., Trajan vs. R), as tuning and training methods between

modelling software packages are slightly different. However, overall results converged
to support the higher reliability of non-linear approaches for modelling logBCF from
molecular descriptors.

Model complexity does not necessarily mean better predictive performance by 283 default, as several non-linear machine learners did not perform well at all. These 284 included ELM and SVM-P, where the RMSE values observed on the test set were >1. 285 286 Although ELM is a feedforward neural network, the weights associated with the neurons in the network are not updated and thus the initialisation of the network is a 287 288 random selection of weights that may not model the output reliably. The EN outperformed QRL and RR models, where the EN is a combination of the penalties 289 (L1 and L2 regularisation) used by both models that usually leads to better predictive 290 performance. The RR model RMSE for the test set data was also lower than the RMSE 291 for the QRL model. This can be observed when comparing RR and QRL methods, as 292 the penalty associated with LASSO can lead to the omission of highly correlated 293 covariables and thus lead to lower model robustness. 294

Limitations of predictive performance may also stem from the raw data. For 295 example, the dataset used herein did not report individual experimental pH, but instead 296 reported a range from 6.0 to 8.5. Therefore, descriptors such as logD that require pH 297 data may become limited and especially where molecular pK_a lies within this 2.5 pH 298 299 unit range. LogD has been shown in several works to influence uptake and accumulation [23-25]. As a compromise, we calculated logD at pH 7, but this may have 300 been different to the exact experimental pH and may have added to predictive 301 inaccuracy across the whole analyte set. Lastly, it is also likely that BCF/BAF 302 prediction will be influenced by variance in biotic factors such as ventilation rates, age, 303

304 genetic factors and metabolism and lay beyond our ability to determine in more detail305 [26, 27].

MLP models trained in Trajan offered the best performance. Consequently, this 306 model was chosen for further investigation in line with the OECD validation guidelines 307 to assess validity of QSAR modelling. The mean absolute error (MAE) corresponded 308 to 0.38 logBCF units for the verification subset (internal validation set) and 0.53 309 310 logBCF for the test subset (external validation set), as shown in Table 1. The RMSE for verification and test subsets were 0.524 and 0.644, respectively. The predictive 311 312 performance of this model was better or comparable to all models in the literature that have attempted to model accumulation processes. Dearden and Shinnawei [28] used 313 a linear QSAR approach to predict BCFs for 135 chemicals with an R^2 of 0.637 and 314 RMSE of 0.661 logBCF units. Another QSAR model by Sahu and Singh [29] used 315 multiple linear regression to predict BCFs for 131 organic compounds with a RMSE of 316 0.556 log units. However, this model was not validated against a test subset and 317 therefore generalised applicability of the model performance is arguably limited. 318

In alternative approaches to linear QSAR models, other machine learning 319 approaches have also been reported [8-10]. A MLP predicted BCFs for 9 test 320 compounds with an average absolute error of 0.33 ±0.22 log units [8]. Whilst the errors 321 322 were low, too few compounds were tested to provide a reliable assessment of its generalisability. In another approach, Zhao et al., [10] used SVM, RBF and MLR 323 models individually. Better performance was observed when two RBF models (using 324 different descriptors) were combined into a 'hybrid' model to predict logBCF. The 325 developed model showed an R^2 of 0.6917 for an external test set with a reported 326 RMSE of 0.69 logBCF units for 119 compounds showing similar performance to the 327 fish-based MLP presented here, using a single MLP. The hybrid model also showed 328

a limitation in the training set, where several cases were not modelled correctly
between the ranges of logBCF 4 to 5 and was observed by a plateau in the regression
analysis.

332

333 A remark on outliers and the applicability domain

Training and testing of all models led to the observation of several common 334 335 outliers. The reason for poor prediction for such cases may stem from under representation in the dataset used for modelling. The spread of input and output data 336 337 between training and validation subsets showed that there was no significant difference between the spread or skew of the data (Figure S2). However, using PCA 338 analysis and distances between the descriptor spaces there were several cases that 339 did not cluster well with the remaining data (Figure 1a). For example, logBCF for 340 perfluorotributylamine was predicted poorly across the majority of trained models. The 341 use of PCA and descriptor data spacing in this way enabled characterisation of the 342 applicability domain (AD) for a given model. A threshold may then be used to 343 determine cases that fall outside the domain and are likely to have higher predictive 344 error (Figure 1b) [30, 31]. 345

According to the OECD QSAR model validation guidance [18], consideration of 346 models for regulatory purposes must be associated with a defined domain of 347 applicability under Principle 3. However, one key consideration in the use of distance-348 based ADs is that input descriptors are not used equally by the model [32]. Therefore, 349 such ADs may not accurately identify those cases having a greater predictive error in 350 every case. This was observed for outliers in the PCA analysis, but where logBCF was 351 predicted relatively well and vice versa. For example, di-2-naphthyldisulfide was not 352 an outlier in the AD but was poorly predicted across all models. On the other hand, 353

pigment yellow-12 was an AD outlier, but logBCF was predicted well by the majorityof models.

Poor predictive accuracy for molecularly similar compounds could be also 356 caused by other factors such as poor quality raw data or too few representative training 357 cases for the model to learn from. It has been shown previously that experimental BCF 358 data can vary from 0.42 to 0.75 log units [9, 33, 34]. Nevertheless, even with the 359 360 limitations associated with defining an AD, it is useful and important to identify any cases that might not be reliably predicted so that rapid prioritisation of compounds can 361 362 begin. Only for these cases, may it then be appropriate to revert to experimental testing. 363

364

365 Machine learning in a regulatory context

Several of the developed machine learning tools in Table 1 showed potential 366 for the replacement and reduction in animal use. However, it is important to recognise 367 the complexities of machine learning approaches from the outset, especially where 368 they are intended for use in regulation. Under Principle 2 of the OECD guidelines, 369 models used in this way must be based on "unambiguous algorithms". In particular, it 370 is highlighted that two significant limitations exist regarding artificial neural networks, 371 for example. These are: (a) the necessity for large (BCF) datasets to develop suitable 372 373 models (which do not exist for some classes of compounds, like pharmaceuticals) and also (b) that these types of machine learning tools are more ambiguous than other 374 types of model, especially those that are linear in nature. For the latter, the guidance 375 is vague concerning appropriateness of ANNs for use under this specific principle but 376 infers that it is an acceptable limitation. Furthermore, the definition of an unambiguous 377 algorithm is in fact ambiguous and should be further refined to prevent confusion to 378

the reader. This principle could be applied in different ways to different models and may cover the generation of molecular descriptors, the feature selection algorithms used, the learning process (for machine learners where the ambiguity lies) and the final model [35]. The majority of the literature seems to have focused on linear models perhaps as a result, mainly to aid in mechanistic understanding and to allow expert interpretation of individual chemicals to provide extra assurance in predicted data (linked to Principle 5).

Principle 5 of the OECD guidelines relates to mechanistic interpretability of 386 387 QSAR models (if possible). This can be considered a limitation for machine learning algorithms if the aim is to achieve an interpretable model, such as would normally be 388 expected of linear models such as OLS or PLS regression. The OECD guidelines also 389 390 remain vague regarding mechanistic interpretation of machine learners. However, whist linear relationships may not be apparent, descriptor sensitivity analyses can 391 indicate the importance of individual descriptors and thus enables interpretation of 392 factors that influence the modelled process. Bioconcentration processes are not 393 simple and extensive datasets are extremely impractical to curate experimentally. 394 Therefore, complex non-linear models may provide a more rapid solution to regulatory 395 decision-making meantime. Therefore, we suggest that guidelines for QSAR model 396 validation need to be expanded to better define the scope of applicability of all the 397 398 different types of machine learning tools and their fitness for purpose in a regulatory context. 399

For PBT testing, the same regulations are triggered when a threshold for bioaccumulation is reached, regardless of the extent to which the threshold is exceeded. Thus, if the value is classified within the correct category of nonbioaccumulative (nB), bioaccumulative (B) or very bioaccumulative (vB), the model will

be useful in the context of PBT assessments. Variability in measurement can arise
from kinetic modelling approaches [17], biological/physiological variability (age, health,
lipid content etc.) [27, 36-39] and experimental conditions (pH, temperature, etc.) [23,
407 40]. As such, reported BCFs have been shown to differ by 1-2 orders of magnitude
even within the same species [27].

The 4-layer MLP here showed a correct classification rate of 90 % across the 409 410 verification and test subsets. The 10 % misclassification of cases was split to 6 % of cases predicted as false negatives and 4 % of cases predicted as false positives (See 411 SI, Figure S3). This is consistent with the hybrid model developed by Zhao et al. which 412 has shown classification accuracies ranging from 91 % to 98 % [9, 10]. It is possible 413 that using QSARs for classification instead of regression analysis may improve the 414 accuracy and without the need for the application of a bias. This would be particularly 415 suitable for bioaccumulation assessments where only a threshold value determines 416 the level of regulation enforced. 417

Some studies have reported the application of models for classification of 418 bioaccumulation thresholds, with accuracies ranging from 84.5 - 91.1 % (depending 419 on model type) [41] and 91.7 % [11]. The authors that used tree-based learners also 420 used these models for quantitative prediction achieving RMSE of 0.554 and R² of 421 0.836 on the test set data [11]. The models tested across the literature have tended to 422 achieve similar performance for both classification and prediction. The agreement in 423 performance between different works and the comprehensive model evaluation here, 424 support that *in silico* methods should be adopted for chemicals where environmental 425 uptake data are limited to enable flexible, cheap and rapid PBT assessment for 426 compound prioritisation. Furthermore, it suggests that the use of chemical descriptors 427 may only be able to achieve a certain level of predictive or classification performance 428

for modelling approaches where other variables become important as mentionedabove.

431

432 Can the developed model be used for cross phylum prediction?

There is little understanding of whether accumulation will be similar across the 433 invertebrate phylum. The dominant site of uptake for waterborne micropollutants in 434 435 fish is across the gills and therefore accumulation across taxa may be significantly different for differing modes of respiration. Other factors such as size, enzyme 436 437 speciation and lipid content may also influence the accumulation potential [27]. The optimised model for fish was applied to the prediction of logBCF in a freshwater 438 invertebrate, Gammarus pulex (Figure 3a). The accumulation data in G. pulex 439 predominantly covered pharmaceuticals and pesticides. The fish-based MLP showed 440 relatively low predictive performance for the invertebrate accumulation factors. The 441 correlation between observed and predicted BCF was R^2 0.3295 with a MAE of 0.80 442 ±0.65 log units, which indicated that the model generalisations between species were 443 limited. The largest predictive error was for the compound imipramine that was 444 overestimated by 2.7 logBCF units. This compound in a previous study had 445 considerable variation in the estimated BCF (212 - 4533) depending on the method 446 of estimation used [17]. 447

A significant difference in BCFs between trophic levels has been shown with higher trophic levels displaying increased BCFs [42]. This trend would suggest that the BCF predictions of the invertebrates might be overestimated but the opposite was observed (62 % of cases were underestimated). In addition to the biological complexity between species, another confounding factor to affect the predictive accuracy and generalisability is the compound class. The fish model included no pharmaceutical

454 compounds whereas the invertebrate BCF data contained 18 cases (~53%). 455 Inspection of the molecular similarity between the datasets indicated that the 456 invertebrate and fish datasets were dissimilar (Figure S4). Thus, the bioconcentration 457 potential may not follow the same relationships with neutral hydrophobic organic 458 contaminants.

The fish-based model was subsequently reinitialised and trained on the 459 invertebrate dataset only (using the same descriptors) (Figure 3b). The invertebrate 460 model showed good correlation with R^2 of 0.9605 with 0.972 for the training set, 0.9932 461 462 for the verification set and 0.9323 for the test set. The model demonstrated good accuracy across the verification and test subset with a MAE of 0.07 ±0.08 logBCF 463 units for the verification set and 0.29 ±0.27 logBCF units for the test set. The 464 successful retraining of the model to invertebrate data suggests that case 465 representation (i.e. compound class) is likely to limit models that are applied across 466 taxa. An alternative approach to overcome this could involve development of a model 467 with two or more outputs to represent different species, but commonality in BCF cases 468 would be required for both species. Whilst the predictive accuracy of the retrained 469 model was very good, it is also limited by the small number of cases used. 470 Generalisability is also likely to be limited given the ratio of cases to descriptors 471 (Topliss ratio of ~2.5:1) Nevertheless, and as new BCF data emerges, this approach 472 473 holds excellent potential by using the same molecular descriptors for BCF predictions in two very different species. In addition, to using the fish-based model to predict 474 invertebrate BCFs we also used the invertebrate-based model to predict fish BCFs of 475 pharmaceuticals reported in the literature (Figure S5). The invertebrate model was 476 able to predict BCFs within the reported range for 45 % of the compounds selected (n 477 = 11). The remaining compounds, with the exception of sertraline and gemfibrozil, 478

were predicted relatively well even though they were not within the reported ranges. 479 Sertraline is an interesting case as although it has not shown very high 480 bioconcentration in fish (BCFs: <1 – 626) [43-47] there have been reported BCF values 481 of up to 32,022 in invertebrates (namely, Lasmigona costata [48] and 990 in Planorbid 482 sp. [49]). As the model used here was trained on BCFs from an invertebrate species, 483 it may not correlate well with fish BCF data, suggesting that cross-phylum predictive 484 modelling may be limited by both case representation and biological variation. 485 However, as the models here used the same descriptors this enables flexibility in 486 487 retraining optimised models and inevitably as more BCF data is generated for the same compounds in different species, this technology could be used to map 488 accumulation across taxa more effectively. It is critically important to understand 489 uptake (internal concentration) across taxa as the conservation of pharmaceutical 490 targets extends widely [50]. 491

492

493 Model sensitivity to descriptors: interpreting accumulation through chemistry

Whilst machine learning models are more difficult to interpret due to the non-494 linear functionality, collinearity and/or curvilinearity; the importance of the 14 495 descriptors described here still offered some mechanistic understanding of the 496 processes involved (Figure 4). For the fish-based model, the most important descriptor 497 498 was TPSA with an error ratio of 2.08. Higher error ratios correspond to increased predictive error for all compounds upon removal of this descriptor from the dataset. 499 Previous investigations have demonstrated that descriptors related to polarisability, 500 hydrophobicity and hydrogen bonding of the molecule is important to modelling BCFs 501 [10, 28, 51]. TPSA is defined as the surface area occupied by nitrogen and oxygen 502 atoms including connected hydrogen atoms [52]. Polar surface area has also been 503

shown to influence drug absorption in humans, where increasing polar surface area decreases the drug fraction absorbed [19, 53]. The relationship between bioconcentration and TPSA may be dependent on several factors such as permeation through the lipid bilayer, binding of polar functional groups to epithelial membranes and the size of hydration shell around a molecule [54].

Permeation through cellular membranes was further supported by the 509 importance of MW to the model. The size of a molecule also affects permeation and 510 diffusion through membranes (Lipinski's rule of five [55]). It has previously been 511 512 demonstrated that dye pigments did not show bioaccumulation in fish due to their large molecular size [56]. In another study, it was suggested that there is a threshold 513 diameter value of 1.5 nm which governed bioconcentration in addition to 514 hydrophobicity [57]. Strempel et al., [11] also found that molecular weight, molecular 515 diameter, TPSA and logD were important for classification and prediction of 516 bioaccumulation. 517

Topological descriptors such as STN, Hnar, Ram, SPI and ICR were also found 518 to be important. These indices are useful especially for differentiating constitutional 519 isomers (except enantiomers) [58]. Error ratios for STN, Hnar, ICR, SPI and Ram 520 spanned from 1.31 – 1.72. These indices are related to molecular branching/shape 521 and the importance of these descriptors relate to molecular size which can influence 522 bioconcentration [59, 60]. MAXDN and MAXDP relate to the partial charges on atoms 523 relative to their topological position within the molecule and therefore relate to the 524 nucleophilicity and electrophilicity of a molecule [61]. Aside from polarity-related 525 accumulation across cellular membranes, it is also possible that these are associated 526 with metabolic activity (from nucleophilic or electrophilic attack). The importance of 527

528 other electrotopological descriptors (along with molecular flexibility) has been 529 previously shown for modelling bioconcentration [62].

Interpretation of the relative importance of descriptors is affected by collinearity 530 or multicollinearity (See SI, Table S4 & S5). The collinearity of the descriptors showed 531 that molecular weight was collinear with SPI (R=0.794) and Ram (R=0.696). The 532 descriptor Ram was also collinear with SPI (R=0.787) and STN was collinear with 533 534 HNar (R=0.748). The relation between these topological descriptors and molecular weight is that they all describe molecular size (shape, volume, weight) to some extent. 535 536 Therefore, the rank importance of these particular descriptors should be approached with some caution. Whilst the error ratio is higher for certain descriptors that are 537 collinear, their removal from the network model may not correctly determine the ratio 538 value due to redundant information. Nevertheless, the descriptor sensitivity can still be 539 useful for directing mechanistic and experimental studies. This was shown recently in 540 a neural network application to passive sampling [63] which was later followed by a 541 mechanistic study [64], that supported the interpretation of the model. 542

The invertebrate-based MLP used the same descriptors as the fish-based 543 model, but the network was reinitialised and retrained. The retraining of the network 544 also showed that the importance of the descriptors changed from the fish-based 545 model. The most important descriptor was HNar (error ratio = 5.75) followed by nN 546 (error ratio = 5.09) and logD (error ratio = 4.71). The increased importance of the 547 number of nitrogen atoms likely reflected the number of pharmaceutical compounds 548 in the dataset. In addition, logD increased in rank to the top three descriptors in the 549 invertebrate model. The increased sensitivity of the model to logD also relates to 550 training of the model with ionisable pharmaceuticals and is in agreement with other 551 studies showing logD to be important in accumulative processes [11, 64]. Whilst 552

hydrophobicity may be a principal factor of bioconcentration, it is possible that carriermediated transport may also play an important role. Both models here demonstrated
that other variables also strongly influence BCF prediction. Thus, QSAR models that
rely solely on logP or logD in our opinion are limited in their application.

It is important to consider that descriptors not used in this work may also have a potential for BCF modelling. For example, the major mechanism of transport across epithelia tissue is passive diffusion and so it is also possible that diffusion coefficients could potentially be an important descriptor for consideration among others, however these descriptors are difficult to acquire and therefore reduce the practicability of a model based on these.

563 **Conclusions**

The work presented herein has shown that in silico modelling approaches are 564 a powerful approach to predict bioconcentration of environmental contaminants, 565 enabling rapid prioritisation of compounds during ERA. The approach could be used 566 to better understand bioaccumulation, and the molecular descriptors that drive it; 567 moving the science beyond simple hydrophobicity models that poorly account for the 568 complexity of pharmaceuticals. Cross-species prediction of accumulation warrants 569 further investigation as the results indicate both case representation and biological 570 variability might limit prediction of accumulation between different taxonomic groups. 571 Nevertheless, the use of machine learning has been increasing within the field and is 572 necessary to improve our understanding of biological processes that affect 573 environmental health. The interpretation of descriptors here is critical as it 574 demonstrates that, in addition to rapid prediction of bioconcentration factors, in silico 575 models are useful for mechanistic understanding which in turn can be used to direct 576 further work. This is particularly true for pharmaceutical uptake in biota, where the 577

mechanisms that govern uptake, elimination and accumulation processes are still not 578 fully understood. Excellent potential exists for rapid screening using machine learning 579 technology in future ERA, without the need for costly and ethically challenging animal 580 experiments. Finally, the OECD QSAR validation guidelines for machine learners are 581 inexplicit and we suggest these guidelines should be expanded with more focus on 582 this type of modelling approach. This will begin to address the applicability and 583 584 usefulness of these models for regulatory schemes such as REACH where PBT assessments are required for several thousand chemicals. 585

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599 **References**

 Rovida, C. and T. Hartung, *Re-evaluation of animal numbers and costs for in vivo tests to* accomplish REACH legislation requirements for chemicals-a report by the Transatlantic Think Tank for Toxicology (t4). ALTEX-Alternatives to animal experimentation, 2009. 26(3): p. 187-208.
 Commission, E., Regulation (EC) No 1907/2006 of the European Parliament and of the Council of 18 December 2006 concerning the Registration, Evaluation, Authorisation and

606		Restriction of Chemicals (REACH), establishing a European Chemicals Agency, amending
607		Directive 1999/45/EC and repealing Council Regulation (EEC) No 793/93 and Commission
608		Regulation (EC) No 1488/94 as well as Council Directive 76/769/EEC and Commission
609		Directives 91/155/EEC, 93/67/EEC, 93/105/EC and 2000/21/EC. 2006: Official Journal of the
610		European Union. p. 1 - 849.
611	3.	Agency, E.M., Guideline on the environmental risk assessment of medicinal products for
612		human use. 2006, European Medicines Agency
613	4.	Agency, E.M., Reflection paper on the authorisation of veterinary medicinal products
614 615		containing (potential) persistent, bioaccumulative and toxic (PBT) or very persistent and very bioaccumulative (vPvB) substances. 2016.
616	5.	Gissi, A., et al., Integration of QSAR models for bioconcentration suitable for REACH. Science
617		of The Total Environment, 2013. 456–457 : p. 325-332.
618	6.	de Wolf, W., et al., Animal use replacement, reduction, and refinement: Development of an
619		integrated testing strategy for bioconcentration of chemicals in fish. Integrated
620		Environmental Assessment and Management, 2007. 3 (1): p. 3-17.
621	7.	Lockwood, S. and N. Saïdi, Background document for public consultation on pharmaceuticals
622		in the environment. 2017.
623	8.	Fatemi, M.H., M. Jalali-Heravi, and E. Konuze, Prediction of bioconcentration factor using
624		genetic algorithm and artificial neural network. Analytica Chimica Acta, 2003. 486(1): p. 101-
625		108.
626	9.	Lombardo, A., et al., Assessment and validation of the CAESAR predictive model for
627		<i>bioconcentration factor (BCF) in fish.</i> Chemistry Central Journal, 2010. 4 (1): p. 1.
628	10.	Zhao, C., et al., A new hybrid system of QSAR models for predicting bioconcentration factors
629		<i>(BCF)</i> . Chemosphere, 2008. 73 (11): p. 1701-1707.
630	11.	Strempel, S., et al., Uusing conditional inference tress and random forests to predict the
631		bioaccumulation potential of organic chemicals. Environmental Toxicology and Chemistry,
632		2013. 32 (5): p. 1187-1195.
633	12.	Institute, C.I.a.T., Biodegradation and Bioaccumulation data of existing chemicals based on
634		the CSCL Japan. 1992, Japan: Chemical Industry Ecology-Toxicology & Information Center.
635	13.	Ashauer, R., A. Boxall, and C. Brown, Uptake and Elimination of Chlorpyrifos and
636		Pentachlorophenol into the Freshwater Amphipod Gammarus pulex. Archives of
637		Environmental Contamination and Toxicology, 2006. 51 (4): p. 542-548.
638	14.	Ashauer, R., et al., Bioaccumulation kinetics of organic xenobiotic pollutants in the
639		freshwater invertebrate Gammarus pulex modeled with prediction intervals. Environmental
640		Toxicology and Chemistry, 2010. 29 (7): p. 1625-1636.
641	15.	Meredith-Williams, M., et al., Uptake and depuration of pharmaceuticals in aquatic
642		invertebrates. Environmental Pollution, 2012. 165(Supplement C): p. 250-258.
643	16.	Miller, T.H., et al., Uptake, biotransformation and elimination of selected pharmaceuticals in
644		a freshwater invertebrate measured using liquid chromatography tandem mass
645		spectrometry. Chemosphere, 2017. 183(Supplement C): p. 389-400.
646	17.	Miller, T.H., et al., Assessing the reliability of uptake and elimination kinetics modelling
647		approaches for estimating bioconcentration factors in the freshwater invertebrate,
648		Gammarus pulex. Science of The Total Environment, 2016. 547(Supplement C): p. 396-404.
649	18.	OECD, Guidance Document on the Validation of (Q)SAR Models. 2007.
650	19.	Palm, K., et al., Polar Molecular Surface Properties Predict the Intestinal Absorption of Drugs
651		in Humans. Pharmaceutical Research, 1997. 14(5): p. 568-571.
652	20.	Kah, M. and C.D. Brown, LogD: Lipophilicity for ionisable compounds. Chemosphere, 2008.
653		72 (10): p. 1401-1408.
654	21.	Devillers, J., et al., <i>Fish Bioconcentration Modelling With LogP</i> . Toxicology Methods, 1998.

8(1): p. 1-10.

656	22.	Dobson, P.D. and D.B. Kell, Carrier-mediated cellular uptake of pharmaceutical drugs: an
657		exception or the rule? Nature Reviews Drug Discovery, 2008. 7: p. 205.
658	23.	Nakamura, Y., et al., The effects of pH on fluoxetine in Japanese medaka (Oryzias latipes):
659		Acute toxicity in fish larvae and bioaccumulation in juvenile fish. Chemosphere, 2008. 70 (5):
660		p. 865-873.
661	24.	Rendal, C., K.O. Kusk, and S. Trapp, Optimal choice of pH for toxicity and bioaccumulation
662		studies of ionizing organic chemicals. Environmental Toxicology and Chemistry, 2011. 30 (11):
663		p. 2395-2406.
664	25.	Karlsson, M.V., et al., Novel Approach for Characterizing pH-Dependent Uptake of Ionizable
665		Chemicals in Aquatic Organisms. Environmental Science & Technology, 2017. 51(12): p.
666		6965-6971.
667	26.	Mackay, D. and A. Fraser, Bioaccumulation of persistent organic chemicals: mechanisms and
668		models. Environmental Pollution, 2000. 110 (3): p. 375-391.
669	27.	Rubach, M.N., et al., Toxicokinetic variation in 15 freshwater arthropod species exposed to
670		the insecticide chlorpyrifos. Environmental Toxicology and Chemistry, 2010. 29(10): p. 2225-
671		2234.
672	28.	Dearden, J.C. and N.M. Shinnawei. Improved prediction of fish bioconcentration factor of
673	-	Hydrophobic Chemicals, SAR and OSAR in Environmental Research, 2004, 15 (5-6); p. 449-
674		455.
675	29	Sahu, V.K. and R.K. Singh, Prediction of the Bioconcentration Factor of Organic Compounds in
676		<i>Eish.</i> CLEAN – Soil. Air. Water. 2009. 37 (11): p. 850-857.
677	30	Aalizadeh, R., et al., <i>Ougntitative Structure–Retention Relationship Models To Support</i>
678		Nontaraet High-Resolution Mass Spectrometric Screening of Emerging Contaminants in
679		Environmental Samples, Journal of Chemical Information and Modeling, 2016, 56(7): n
680		138/_1308
681	31	Meaver S and M.P. Gleeson. The importance of the domain of annlicability in OSAR
682	51.	modeling Journal of Molecular Graphics and Modelling 2008 26 (8): p. 1315-1326
682	27	Notzeva TL et al. Current status of methods for defining the annlicability domain of
607	52.	(augntitative) structure activity relationships ATLA 2005 23 : p. 155 172
004 60E	22	(qualitative) structure-activity relationships. ATLA, 2003. 33 . p. 133-173.
005	55.	chemicale SAD and OSAD in Environmental Desearch 2005 16 (6) in E21 EE4
080	24	Chemiculs. SAR and QSAR in Environmental Research, 2005. 16(0): p. 531-554.
687	34.	Arnot, J.A. and F.A.P.C. Gobas, A review of bioconcentration factor (BCF) and
688		bioaccumulation Jactor (BAF) assessments for organic chemicals in aquatic organisms.
689	25	Environmental Reviews, 2006. 14(4): p. 257-297.
690	35.	Gramatica, P., Principles of QSAR models validation: Internal and external. QSAR &
691		Combinatorial Science, 2007. 26 (5): p. 694-701.
692	36.	Verhaar, H.J.M., J. de Jongh, and J.L.M. Hermens, <i>Modeling the Bioconcentration of Organic</i>
693		<i>Compounds by Fish: A Novel Approach.</i> Environmental Science & Technology, 1999. 33 (22):
694		p. 4069-4072.
695	37.	Hendriks, A.J., et al., The power of size. 1. Rate constants and equilibrium ratios for
696		accumulation of organic substances related to octanol-water partition ratio and species
697		weight. Environmental Toxicology and Chemistry, 2001. 20(7): p. 1399-1420.
698	38.	Buchwalter, D.B., J.J. Jenkins, and L.R. Curtis, Respiratory strategy is a major determinant of
699		[3H]water and [14C]chlorpyrifos uptake in aquatic insects. Canadian Journal of Fisheries and
700		Aquatic Sciences, 2002. 59(8): p. 1315-1322.
701	39.	Rubach, M.N., D.J. Baird, and P.J. Van den Brink, A new method for ranking mode-specific
702		sensitivity of freshwater arthropods to insecticides and its relationship to biological traits.
703		Environmental Toxicology and Chemistry, 2010. 29(2): p. 476-487.
704	40.	Karara, A.H. and W.L. Hayton, A pharmacokinetic analysis of the effect of temperature on the
705		accumulation of di-2-ethylhexyl phthalate (DEHP) in sheepshead minnow. Aquatic
706		Toxicology, 1989. 15 (1): p. 27-36.

707 41. Sun, X., et al., Classification of bioaccumulative and non-bioaccumulative chemicals using 708 statistical learning approaches. Molecular Diversity, 2008. 12(3): p. 157. 709 42. LeBlanc, G.A., Trophic-Level Differences in the Bioconcentration of Chemicals: Implications in 710 Assessing Environmental Biomagnification. Environmental Science & Technology, 1995. 711 **29**(1): p. 154-160. 712 43. Grabicova, K., et al., Tissue-specific bioconcentration of antidepressants in fish exposed to 713 effluent from a municipal sewage treatment plant. Science of The Total Environment, 2014. 714 488-489: p. 46-50. 715 44. Lajeunesse, A., et al., Distribution of antidepressants and their metabolites in brook trout 716 exposed to municipal wastewaters before and after ozone treatment – Evidence of biological 717 effects. Chemosphere, 2011. 83(4): p. 564-571. 718 45. Tanoue, R., et al., Simultaneous determination of polar pharmaceuticals and personal care 719 products in biological organs and tissues. Journal of Chromatography A, 2014. 1355: p. 193-720 205. 721 46. Togunde, O.P., et al., Determination of Pharmaceutical Residues in Fish Bile by Solid-Phase 722 Microextraction Couple with Liquid Chromatography-Tandem Mass Spectrometry 723 (LC/MS/MS). Environmental Science & Technology, 2012. 46(10): p. 5302-5309. 724 47. Xie, Z., et al., Occurrence, bioaccumulation, and trophic magnification of pharmaceutically 725 active compounds in Taihu Lake, China. Chemosphere, 2015. 138: p. 140-147. 726 de Solla, S.R., et al., Bioaccumulation of pharmaceuticals and personal care products in the 48. 727 unionid mussel Lasmigona costata in a river receiving wastewater effluent. Chemosphere, 728 2016. **146**: p. 486-496. 729 49. Du, B., et al., Pharmaceutical bioaccumulation by periphyton and snails in an effluent-730 dependent stream during an extreme drought. Chemosphere, 2015. 119: p. 927-934. 731 50. Verbruggen, B., et al., ECOdrug: a database connecting drugs and conservation of their 732 targets across species. Nucleic Acids Research, 2018. 46(D1): p. D930-D936. 733 51. Gramatica, P. and E. Papa, QSAR Modeling of Bioconcentration Factor by theoretical 734 molecular descriptors. QSAR & Combinatorial Science, 2003. 22(3): p. 374-385. 735 52. Pajouhesh, H. and G.R. Lenz, Medicinal Chemical Properties of Successful Central Nervous 736 System Drugs. NeuroRX, 2005. 2(4): p. 541-553. 737 53. Kelder, J., et al., Polar Molecular Surface as a Dominating Determinant for Oral Absorption 738 and Brain Penetration of Drugs. Pharmaceutical Research, 1999. 16(10): p. 1514-1519. 739 54. Skyner, R., et al., A review of methods for the calculation of solution free energies and the 740 modelling of systems in solution. Physical Chemistry Chemical Physics, 2015. 17(9): p. 6174-741 6191. 742 55. Tice, C.M., Selecting the right compounds for screening: does Lipinski's Rule of 5 for 743 pharmaceuticals apply to agrochemicals? Pest Management Science, 2001. 57(1): p. 3-16. 744 56. Anliker, R., P. Moser, and D. Poppinger, Advances in Environmental Hazard and Risk 745 Assessment 1987 Bioaccumulation of dyestuffs and organic pigments in fish. Relationships to 746 hydrophobicity and steric factors. Chemosphere, 1988. 17(8): p. 1631-1644. 747 57. Dimitrov, S.D., et al., Predicting bioconcentration factors of highly hydrophobic chemicals. 748 Effects of molecular size, in Pure and Applied Chemistry. 2002. p. 1823. 749 58. Randić, M., et al., A rational selection of graph-theoretical indices in the QSAR. International 750 Journal of Quantum Chemistry, 1988. 34(S15): p. 267-285. 751 59. Anliker, R., P. Moser, and D. Poppinger, Bioaccumulation of dyestuffs and organic pigments 752 in fish. Relationships to hydrophobicity and steric factors. Chemosphere, 1988. 17(8): p. 753 1631-1644. 754 60. Opperhulzen, A., et al., Relationship between bioconcentration in fish and steric factors of 755 hydrophobic chemicals. Chemosphere, 1985. 14(11): p. 1871-1896.

- Gramatica, P., M. Corradi, and V. Consonni, *Modelling and prediction of soil sorption coefficients of non-ionic organic pesticides by molecular descriptors.* Chemosphere, 2000. **41**(5): p. 763-777.
- Wang, Y., et al., *Estimation of bioconcentration factors using molecular electro-topological state and flexibility*. SAR and QSAR in Environmental Research, 2008. **19**(3-4): p. 375-395.
- 63. Miller, T.H., et al., *The First Attempt at Non-Linear in Silico Prediction of Sampling Rates for*762 *Polar Organic Chemical Integrative Samplers (POCIS)*. Environmental Science & Technology,
 763 2016. 50(15): p. 7973-7981.
- Morin, N.A.O., et al., *Kinetic accumulation processes and models for 43 micropollutants in "pharmaceutical" POCIS.* Science of The Total Environment, 2018. 615(Supplement C): p. 197-207.

		RMSE			R ²			MAE		
	Model	Training	Verification	Test	Training	Verification	Test	Training	Verification	Test
Trajan	Linear	0.785	1.052	0.832	0.532	0.390	0.521	0.619	0.835	0.608
	GRNN	0.830	0.893	0.873	0.673	0.400	0.569	0.664	0.893	0.718
	RBF	0.723	0.689	0.584	0.651	0.635	0.725	0.565	1.600	0.450
	3-MLP	0.689	0.538	0.337	0.675	0.770	0.659	0.548	1.608	0.553
	4-MLP	0.403	0.524	0.644	0.887	0.819	0.702	0.313	0.380	0.530
	Model	Training	Cross-Validation	Test	Training	Cross-Validation	Test	Training	Cross-Validation	Test
R	OLM	0.719	0.771	1.203	0.621	0.570	0.234	0.560	NA	0.778
	PLS	0.722	0.769	1.164	0.618	0.571	0.254	0.564	NA	0.765
	RR	0.725	0.766	1.083	0.614	0.576	0.304	0.568	NA	0.753
	EN	0.729	0.760	1.054	0.612	0.582	0.314	0.577	NA	0.754
	QRL	0.733	0.757	1.112	0.607	0.585	0.284	0.562	NA	0.770
	KNN	0.517	0.683	0.902	0.807	0.665	0.468	0.404	NA	0.648
	ELM	0.673	0.756	1.014	0.668	0.593	0.346	0.529	NA	0.768
	ANN-1HL	0.596	0.751	0.877	0.739	0.597	0.505	0.462	NA	0.620
	ANN-a1HL	0.395	0.672	0.859	0.888	0.678	0.518	0.319	NA	0.612
	ANN-2HL	0.232	0.834	1.022	0.962	0.560	0.370	0.174	NA	0.680
	ANN-3HL	0.454	0.795	0.880	0.860	0.582	0.520	0.345	NA	0.624
	MARS	0.539	0.730	1.014	0.787	0.632	0.390	0.425	NA	0.696
	B-MARS	0.500	0.681	0.899	0.819	0.673	0.479	0.395	NA	0.633
	SVM-R	0.383	0.644	0.841	0.893	0.704	0.537	0.261	NA	0.590
	SVM-P	0.699	0.747	1.029	0.643	0.594	0.340	0.539	NA	0.729
	RF-CART	0.292	0.675	0.771	0.956	0.688	0.633	0.231	NA	0.589
	RF-CIT	0.605	0.739	0.821	0.762	0.630	0.586	0.485	NA	0.652
	BT	0.249	0.660	0.789	0.957	0.687	0.593	0.187	NA	0.587
	CR	0.353	0.678	0.973	0.910	0.673	0.431	0.282	NA	0.628

Table 1: Comparison of model performance for the prediction of BCF in *Cyprinus carpio*. MAE is the mean absolute error and NA indicates the metric was not applicable.



Figure 1: (a) Principal component analysis used for visualisation of the case similarity based on the 14 modelled descriptors (i.e. applicability domain). (b) Distances between cases in the PCA space with a threshold applied (0.975 quantile of χ^2 distribution) designated by the red line (c) the distribution of cases based on distance in the PCA space.



Figure 2: (a) linear regression of the predicted logBCF values versus the observed logBCF values in fish using the 4-MLP developed in approach 1, training data (crosses, n = 242), verification data (circles, n = 55) and test data (triangles, n = 55). (b) Raw residuals of the predicted logBCF data in fish for the verification and test data only.



Figure 3: (a) Comparison of the predicted logBCF data versus the observed logBCF in invertebrates using the fish-based 4-layer MLP. (b) Regression of a separately developed and optimised model trained with the invertebrate BCF data (*Gammarus pulex*), training set (crosses, n = 24), verification set (circles, n = 5) and test set (triangles, n = 5)



Figure 4: Descriptors sensitivity analysis performed by removing a descriptor from the model and assessing the affected performance. Increased error ratios indicate more important descriptors. (a) descriptor sensitivity for the fish-based model and (b) for the invertebrate-based model.