



King's Research Portal

DOI: 10.1093/cid/ciz047

Document Version Peer reviewed version

Link to publication record in King's Research Portal

Citation for published version (APA):

Mehra, V., Rhone, E., Widya, S., Zuckerman, M., Potter, V., Raj, K., Kulasekararaj, A. G., McLornan, D. P., de Lavallade, H., Benson-Quarm, N., Lim, C., Ware, S., Sudhanva, M., Malik, O., Nicholas, R., Muraro, P. A., Marsh, J., Mufti, G. J., Silber, E., ... Kazmi, M. (2019). Epstein-Barr Virus and Monoclonal Gammopathy of Clinical Significance in Autologous Stem Cell Transplantation for Multiple Sclerosis. *Clinical Infectious Diseases*, 69(10), 1757-1763. https://doi.org/10.1093/cid/ciz047

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- •Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- •You may not further distribute the material or use it for any profit-making activity or commercial gain •You may freely distribute the URL identifying the publication in the Research Portal

If you believe that this document breaches copyright please contact librarypure@kcl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

Download date: 28. Dec. 2024

Clinical Infectious Diseases

EBV and Monoclonal Gammopathy of Clinical Significance in Autologous Stem Cell Transplantation for Multiple Sclerosis --Manuscript Draft--

	OUR ASSESSED
Manuscript Number:	CID-92283R2
Full Title:	EBV and Monoclonal Gammopathy of Clinical Significance in Autologous Stem Cell Transplantation for Multiple Sclerosis
Short Title:	EBV complications in Auto-HSCT for MS
Article Type:	Major Article
Corresponding Author:	Varun Mehra, MRCP (UK), FRCPath King's College Hospital London, UNITED KINGDOM
Corresponding Author Secondary Information:	
Corresponding Author's Institution:	King's College Hospital
Corresponding Author's Secondary Institution:	
First Author:	Varun Mehra, MRCP (UK), FRCPath
First Author Secondary Information:	
Order of Authors:	Varun Mehra, MRCP (UK), FRCPath
	Elijah Rhone
	Stefani Widya
	Mark Zuckerman
	Victoria Potter
	Kavita Raj
	Austin Kulasekararaj
	Donal McLornan
	Hugues de Lavallade
	Nana Benson-Quarm
	Christina Lim
	Sarah Ware
	Malur Sudhanva
	Omar Malik
	Richard Nicholas
	Paolo A Muraro
	Judith Marsh
	Ghulam J Mufti
	Eli Silber
	Antonio Pagliuca
	Majid A. Kazmi
Order of Authors Secondary Information:	

Manuscript Region of Origin:	UNITED KINGDOM			
Abstract:	Introduction Autologous haematopoietic stem cell transplantation (AHSCT) with anti-thymocyte globulin (ATG) conditioning as treatment of active multiple sclerosis (MS) is rapidly increasing across Europe (EBMT registry data 2017). Clinically significant Epstein Barr virus reactivation (EBV-R) following AHSCT with ATG for severe autoimmune conditions is an under-recognised complication relative to T-cell deplete transplants performed for haematological diseases. This retrospective study reports EBV-R associated significant clinical sequelae in MS patients undergoing AHSCT with rabbit ATG. Methods Retrospective data was analysed for 36 consecutive MS-AHSCT patients at Kings College Hospital, London. All patients routinely underwent weekly EBV DNA PCR monitoring and serum electrophoresis for monoclonal gammopathy (MG or M-protein). EBV-R with rising EBV viral load, M-protein and associated clinical sequelae were captured from clinical records. Results All patients had evidence of rising EBV DNA-emia, including 7 who were lost to long term follow-up, with a number of them developing high EBV viral load & associated lymphoproliferative disorder (LPD). Nearly 72% (n-18/29) developed de-novo MG, some with significant neurological consequences with high M-protein and EBV-R. Six patients required anti-CD20 therapy (rituximab) with complete resolution of EBV related symptoms. Receiver operating characteristics (ROC) estimated a peak EBV viraemia of >500,000 DNA copies/ml correlated with high sensitivity (85.5%) & specificity (82.5%) (AUC-0.87; p-0.004) in predicting EBV-R related significant clinical events. Conclusion Symptomatic EBV reactivation increases risk of neurological sequelae and LPD in MS-AHSCT. We recommend regular monitoring for EBV and serum electrophoresis for MG			
Response to Reviewers:	To Dr Barbara D Alexander M.D. Associate Editor Clinical Infectious Diseases Dated: 30th Dec 2018 Dear Dr Alexander Subject: Response to Reviewers Manuscript Title: EBV and Monoclonal Gammopathy of Clinical Significance in Autologous Stem Cell Transplantation for Multiple Sclerosis. We would like to thank the journal for provisionally accepting our work. Considering the reviewer's comments, we have made revisions to the manuscript with responses outlined for each of the queries raised by the reviewer, as below: The Authors simply must have the manuscript edited for English grammar as many of the mistakes change the meaning of the sentence. Some (but not all) of the issues are as follows: Response: Please accept our apologies for the grammatical errors in the manuscript. We have reviewed and edited these errors where appropriate including the ones highlighted below. Line 138 and line 357. deleted " be mandated". Cohort is too small to warrant "mandate". but your data can lead to recommendation Response: We have edited and replaced the word 'mandate' from the phrase. Line 212-215. Please include the conversion factor for your assay to IU/ml in the methods section i.e 10 EBV DNA copies/ml=10 IU/ml			

Response:

This has been rephrased within methods section; line 202-203

line 282: HAS versus IS?...I think "is"

Response:

Correction made to "is"

Line 298 and 300- not sure systemic sclerosis needs to be capitalized. But if so, needs to be so throughout manuscript

Response

We have edited and removed un-necessary capitalisation for similar errors across the manuscript.

Lines 309-312. This sentence is not understandable based on current punctuation. Please address. ????This is further corroborated by the fact that similar LPD risk has not been observed in other ADs managed with ATG in our center. For example, among patients with Crohn' disease treated with ATG-AHSCT and those with severe aplastic anemia treated with ATG/cyclopsorin, only 52% (x/x) developed EBV-R (unpublished data) and none had LPD, suggesting that the problem may not be ATG specific. Response:

Thank you for the suggestion. We have rephrased this to reflect our experience with other Autoimmune diseases (lines 310-315).

Line 319 delete the words "may still have"

Response:

correction made.

Line 330 seems to "be"?

Response:

correction made.

Line 340 "copies"/ml

Response:

correction made.

Thank you again for your review of the revised manuscript. We hope these revisions are satisfactory and will allow formal acceptance for publication.

Yours sincerely

On behalf of all co-authors:

Dr Varun Mehra, MRCP(UK), FRCPath Department of Haematological Medicine Dr Majid Kazmi; FRCP(UK) FRCPath Chief of Cancer Division & Consultant

Haematologist

Kings College Hospital, London, UK

Kings, Guy's & St Thomas' Hospital, London,

UK

Varun.Mehra@nhs.net; Ph-004478865087013Majidkazmi@nhs.net; PH-004478027617716

King's College Hospital

Editor revised Cover Letter.docx

NHS Foundation Trust



To Professor Robert Schooley, M.D. The Editor-in-Chief Clinical Infectious Diseases

Dated: 20th Dec 2018

Dear Professor Schooley (Editor-in-Chief) and Dr Alexander (Associate Editor)

We are pleased to submit our revised article entitled; "EBV & Monoclonal Gammopathy of Clinical Significance in Autologous Stem Cell Transplantation for Multiple Sclerosis" for consideration of publication in your internationally reputed journal, Clinical Infectious Diseases.

Just to summarise again: Autologous Stem Cell Transplants (AHSCT) with anti-thymocyte globulin ATG) based conditioning is a novel approach to treatment of active multiple sclerosis (MS) and recent data from MIST study collaborators (Burt et al; Clinical Trial Registry: NCT00273364) have shown some exciting preliminary results showing superiority of AHSCT over established disease modifying therapies, confirming results from other UK and international studies in this field. However, as the evidence builds, safety aspects of these procedures needs to be seriously considered.

This study reports rates of Epstein Barr virus (EBV) reactivation and associated clinical sequelae with monoclonal gammopathy (M-protein), in cohort of Multiple Sclerosis patients who underwent ATG conditioned immunosuppressive AHSCT in a single centre. We report a significantly higher proportion of MS patients had detectable EBV DNA post-AHSCT; were more likely to develop clinically significant EBV viraemia of >500,000 DNA copies/ml and develop de-novo M-protein of clinical significance with clinical events ranging from probable lymphoproliferative disorders and disabling neurological complications, unrelated to MS. This report of significant clinical complications related to EBV and M-protein, possibly reflect underlying altered immunopathological state of MS disease and its interactions with reactivation of EBV virus, which if monitored and treated pre-emptively may reduce associated morbidity and improve outcomes.

To help readers, we have also described two interesting clinical vignettes as a supplementary to this report, highlighting significant risk of neurological events following development of M-protein, triggered following EBV reactivations in MS patients.

We can confirm that this manuscript has not been published and is not under consideration for publication elsewhere. All authors have seen, approved and contributed to this work. We have no conflicts of interest to disclose. We believe that this report fits well within the scope of your journal, highlighting important clinical message about EBV complications in ATG conditioned AHSCT for MS and will appeal to journal's readers interested in infectious complications related to immunosuppressive therapies including AHSCTs for autoimmune conditions, with a potential to change clinical practice in this area. We have provided point to point responses to the reviewer's comments.

Thank you for your consideration of this revised manuscript and looking forward to your acceptance.

Yours Sincerely

On behalf of all co-authors: Dr Varun Mehra, MRCP(UK), FRCPath Department of Haematological Medicine Kings College Hospital, London, UK Varun.Mehra@nhs.net; Ph-004478865087013

Dr Majid Kazmi; FRCP(UK) FRCPath

Chief of Cancer Division & Consultant Haematologist Kings, Guy's & St Thomas' Hospital, London, UK Majidkazmi@nhs.net; PH-004478027617716

EBV Monoclonal Gammopathy of Clinical Significance and in 1

- Autologous Stem Cell Transplantation for Multiple Sclerosis. 2
- 3 Varun Mehra*, Elijah Rhone*, Stefani Widya, Mark Zuckerman, Victoria Potter, Kavita Raj,
- 4 Austin Kulasekararaj, Donal McLornan, Hugues de Lavallade, Nana Benson-Quarm, Christina
- 5 Lim, Sarah Ware, Malur Sudhanva, Omar Malik, Richard Nicholas, Paolo A Muraro, Judith
- Marsh, Ghulam J Mufti, Eli Silber, Antonio Pagliuca and Majid A. Kazmi 6

Author Affiliations:

7

8

9

11 12

13 14

15

16

17 18

19 20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36 37

38

39

40

41 42

43

44

- 1. Dr Varun Mehra*; Department of Haematology, King's College Hospital NHS Foundation 10 Trust, Denmark Hill, London, United Kingdom.
 - 2. Dr Elijah Rhone*: Department of Neurology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - 3. Stefani Widya: GKT School of Medical Education, Kings College London University, London
 - 4. Dr Mark Zuckerman: Department of Virology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - 5. Dr Victoria Potter: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - 6. Dr Kavita Raj: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom AND Department of Haematology, Guy's and St. Thomas' NHS Foundation Trust, London, United Kingdom
 - 7. Dr Austin Kulasekararaj: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - 8. Dr Donal McLornan Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom AND Department of Haematology, Guy's and St. Thomas' NHS Foundation Trust, London, United Kingdom
 - 9. Dr Hugues de Lavallade: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - 10. Nana Benson-Quarm: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - 11. Christina Lim: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill. London. United Kingdom.
 - 12. Sarah Ware: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **13. Dr Malur Sudhanva:** Department of Virology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - 14. Dr Omar Malik: Department of Neurology, Imperial College Healthcare, London, United Kingdom
 - 15. Dr Richard Nicholas: Department of Neurology, Imperial College Healthcare, London, United Kinadom
 - **16. Professor Paolo A. Muraro:** Department of Neurology, Imperial College Healthcare, London, United Kingdom AND Department of Neuroimmunology, Imperial College London, London, United Kingdom
 - 17. Professor Judith Marsh: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom

18. Professor Ghulam J. Mufti: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom 19. Dr Eli Silber: Department of Neurology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom 20. Professor Antonio Pagliuca: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom 21. Dr Majid Kazmi: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom AND Department of Haematology, Guy's and St. Thomas' NHS Foundation Trust, London, United Kingdom *These authors contributed equally to this work Corresponding authors: Dr Varun Mehra; Varun.Mehra@nhs.net; +442032995378 Running Title: EBV complications in Auto-HSCT for MS Summary: EBV reactivation is common post-transplant with ATG for multiple sclerosis (MS), with significant lymphoproliferative & neurological sequelae associated with rising Mprotein. Serial monitoring of EBV & M-protein is recommended post-transplant, as is pre-emptive anti-CD20 therapy with EBV DNA >500,000 copies/ml.

Abstract

Introduction

Autologous haematopoietic stem cell transplantation (AHSCT) with anti-thymocyte globulin (ATG) conditioning as treatment of active multiple sclerosis (MS) is rapidly increasing across Europe (EBMT registry data 2017). Clinically significant Epstein Barr virus reactivation (EBV-R) following AHSCT with ATG for severe autoimmune conditions is an under-recognised complication relative to T-cell deplete transplants performed for haematological diseases. This retrospective study reports EBV-R associated significant clinical sequelae in MS patients undergoing AHSCT with rabbit ATG.

Methods

Retrospective data was analysed for 36 consecutive MS-AHSCT patients at Kings College
Hospital, London. All patients routinely underwent weekly EBV DNA PCR monitoring and
serum electrophoresis for monoclonal gammopathy (MG or M-protein). EBV-R with rising
EBV viral load, M-protein and associated clinical sequelae were captured from clinical
records.

Results

All patients had evidence of rising EBV DNA-emia, including 7 who were lost to long term follow-up, with a number of them developing high EBV viral load & associated lymphoproliferative disorder (LPD). Nearly 72% (n-18/29) developed de-novo MG, some with significant neurological consequences with high M-protein and EBV-R. Six patients required anti-CD20 therapy (rituximab) with complete resolution of EBV related symptoms. Receiver operating characteristics (ROC) estimated a peak EBV viraemia of >500,000 DNA

103	copies/ml correlated with high sensitivity (85.5%) & specificity (82.5%) (AUC-0.87; p-0.004
104	in predicting EBV-R related significant clinical events.
105	Conclusion
106	Symptomatic EBV reactivation increases risk of neurological sequelae and LPD in MS
107	AHSCT. We recommend regular monitoring for EBV and serum electrophoresis for MG in
108	MS patients in the first 3 months post AHSCT
109	Key Words:
110 111 112	Multiple Sclerosis; Autologous Hematopoietic Stem Cell Transplantation, Epstein-Barr Virus Infection; Monoclonal Gammopathy; Post-transplant Lymphoproliferative Disorder
113	
114	
115	
116	
117	
118	
119	

INTRODUCTION:

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

Multiple sclerosis (MS) is a chronic autoimmune, inflammatory, demyelinating disease of the central nervous system[1]⁻[2], with a relapsing-remitting (RRMS) presentation in the majority of patients at diagnosis. Recovery from relapses may be complete or partial[3][4]. After a variable period of time, people with RRMS may develop a more progressive disability accumulation with or without superimposed relapses; termed secondary progressive multiple sclerosis (SPMS). A minority experience progressive disability from the onset of disease, termed primary progressive multiple sclerosis (PPMS)[4]. A number of immunomodulatory disease modifying therapies (DMTs) are currently licensed for treatment of RRMS with an aim of reducing number of relapses and accrual of disability, although with variable efficacy[5]. Since 1996, Autologous Hematopoietic Stem Cell Transplantation (AHSCT) has been a novel approach for MS management, using immunoablation followed by immunomodulation mechanisms, with evidence of significant suppression of inflammatory activity and qualitative changes in the reconstituted immune system (immune reset theory)[6-8]. AHSCT appears most effective for MS patients with evidence of inflammatory activity on MRI, younger age, a shorter disease duration, low to moderate disability levels (Expanded Disability Status Scale [EDSS] <6 or up to 6.5 if recent progression) and failure of at least 1 highly active DMT (natalizumab or alemtuzumab) with no significant comorbidities[9-11]. Recently reported preliminary results of randomised MIST study[12] found AHSCT to be superior to standard disease modifying therapy (DMT)

for RRMS with respect to both treatment failure and disability progression.

141

142

143

144

145

146

147

148

149

150

140

subsequent opportunistic infections following However, risk rise in immunosuppressive therapies remain a potential concern[13]. MS patients undergoing AHSCT have often been exposed to a number of immunomodulating DMTs; the addition of immunosuppressive rabbit anti-thymocyte globulin (rATG) to their conditioning regimen may confer a higher risk of viral reactivation in these patients. The number of AHSCTs performed for MS is rising significantly in Europe[14] and as more centres perform AHSCT for this indication, it is increasingly important to recognise the unique problems faced by these patients post AHSCT. This retrospective study reports for the first time, EBV-R associated neurological sequelae and lymphoproliferative disorder (LPD) in MS patients undergoing rATG conditioned AHSCT in our centre.

152

153

154

155

156

157

158

159

151

METHODS

Patients and procedures

Data was collected retrospectively on 36 consecutive MS patients undergoing AHSCT between February 2012 and February 2017 at Kings College Hospital, London. Peripheral blood stem cells were collected following standard mobilisation strategy consisting of cyclophosphamide 4g/m² over 2 days and granulocyte colony-stimulating factor for 7 days. Thirty-five (97%) patients were conditioned using standard protocol of cyclophosphamide

(50mg/kg for 4 days) and rATG (2.5mg/kg/day for 3 days) for in-vivo lymphodepletion followed by stem cell infusion. One patient was conditioned with carmustine/etoposide/cytarabine/melphalan regimen along with an equivalent dose of rATG (BEAM-ATG) prior to stem cell infusion. The median CD34 stem cell dose returned was 7.17 x10^6/kg (range 4.0-17.1x10^6/kg).

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

160

161

162

163

164

Prior exposure to EBV was assessed by serological evidence (EBV viral capsid antigen; EBV DNA load monitoring was performed on whole blood samples by VCA IgG). standardised quantitative real-time polymerase chain reaction (RT-PCR) using Rotor-Gene[™] (Qiagen) assay of EBV BZLF1 DNA. This assay was adapted from our published assay using LightCycler (Roche)[15] and since been validated against the recently published WHO standard, with our lab's EBV DNA quantification of 10 copies/ml considered equivalent to 10 IU/ml DNA reported with the WHO reference method[16]. EBV-R was defined as rising EBV DNA load of >10 copies/millilitre (ml) detected on two consecutive tests based on our assay sensitivity. Symptomatic EBV-R was captured by peak blood EBV DNA load, presence of B symptoms (defined by presence of either unexplained weight loss, recurrent fever, night sweats); which was in-turn defined by clinical, radiological and/or histological evidence based on recent ECIL-6 guidelines[17]. In addition, significant 'clinical events' were also defined as new & persistent organ dysfunction (e.g. neurological events) temporally associated with rising EBV viraemia in MS patients. Serum protein electrophoresis was routinely tested around 3 months post HSCT as part of our institutional practice, with immunoglobulin subclasses identified by immunofixation electrophoresis. Patient outcomes were assessed at last follow up as of April 2017.

Statistics

The database of transplants and outcomes was built in Microsoft Excel 2016 and statistical analyses were performed using IBM SPSS Statistics version 24.0. Patient characteristics are presented as medians (with inter-quartile ranges; IQR) for data with non-normal distribution. Comparisons of baseline characteristics used Mann-Whitney U test, Fisher's exact test, or Chi-squared test for trend as appropriate. Receiver Operating characteristics (ROC) curve was obtained correlating LPD and clonal gammopathy associated clinical events with rising EBV viraemia (copies/ml).

RESULTS

Baseline characteristics are presented in **Table 1**. Most MS patients (88.9%) had RRMS phenotype with median of 2 (range 0-3) DMTs prior to AHSCT. Twenty-two MS patients had prior exposure to natalizumab and 7 were treated with Alemtuzumab (6 patients received both). All 36 (100%) MS patients were serologically positive for EBV VCA IgG pretreatment, indicating prior EBV exposure and had detectable EBV DNA post-AHSCT. Seven MS patients were lost to long-term follow up for EBV monitoring. The median time to first EBV DNA detection post-transplant was 30 days (IQR 23-46 days). EBV DNA levels peaked at a median of 32 days post-transplant (IQR 31-53 days). All MS patients had normal baseline lymphocyte counts (median) pre-HSCT with a median time of 46 days (range 14-404 days) to lymphocyte recovery (defined by total lymphocyte count >1.0x106/ml) following AHSCT (**See Figure 1**). A high proportion (86%; n-25/29) of the MS patients in active follow-up recovered lymphocyte counts around D56 with a median

lymphocyte count of 1.56 (10⁶ cells/ml); Four patients remained lymphopenic at last follow up.

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

206

207

All patients were stratified into following 3 groups according to peak rise/burden of EBV DNA-aemia (copies/ml): <100,000 (<100k) copies/ml, 100,001-500,00 (100k-500k) copies/ml and >500,000 (>500k) copies/ml to identify any specific thresholds for clinically significant events related to rising EBV-R (Table 1). The majority of patients (76%) with rising EBV viral load >100k copies/ml were routinely screened by computed tomographic (CT) scans to assess for evidence of LPD, in line with our institutional policy. One third (34.5%) of patients developed peak EBV viraemia of >500k copies/ml. Eight patients (27.6%) developed symptomatic EBV-R; defined as persistent fever, lymphadenopathy and/or B symptoms. Of these 8 patients, only 1 (12.5%) had a peak EBV viraemia <100kDNA copies/ml with the remaining 7 (87.5%) patients having a peak EBV viraemia of >500k copies/ml. Three patients with rising EBV viraemia >500k copies/ml had findings consistent with probable LPD on CT imaging; however, none had definitive histological diagnosis. Three MS patients had worsening neurological symptoms concurrent with rising EBV viraemia >500k copies/ml and clonal gammopathy, as described below.

223

224

225

Interestingly, we also observed frequent de novo monoclonal gammopathy (MG or M-protein) in 18 MS patients (62.4%) following rising EBV viraemia; the majority (n-16) of

whom developed IgG subtype and the remaining 2 developed IgA and IgM M-protein. Concerningly two of these patients developed clinically significant M-Protein burden; one patient with IgG Kappa M-protein of 45.6g/L developed hyper-viscosity and neurological symptoms mimicking MS relapse, requiring plasma exchange. Another patient developed significant lumbosacral radiculopathy with rising EBV-R and high IgM lambda M-protein (IgM 48.5g/L) (see supplementary case vignettes). Figure 2 highlights the association of neurological symptom onset following rising EBV viraemia (log copies), falling lymphocyte counts (x 10⁶/ml) with significant rise in M-protein (gm/lt) levels post AHSCT. A third patient developed painful lower limb paraesthesia following rising EBV viraemia >500k copies/ml, although did not have any M-protein detected. Their symptoms persisted at last follow up despite no evidence of MS related new disease activity.

Six patients were treated with anti-CD20 antibody, rituximab (375mg/m2 weekly up to 4 weeks), due to clinical severity of EBV reactivations and leading to reduction in EBV viral load and concurrent improvement/resolution of EBV related symptoms. ROC curve analysis (**Figure 3**) confirmed EBV viraemia of >500k copies/ml correlated with high sensitivity (85.5%) and specificity (82.5%) (AUC-0.87; 95%CI-0.73-1.0; p-0.004) in predicting significant EBV related clinical events (evidence of LPD and/or neurological symptoms) that may require treatment with rituximab. The sensitivity dropped significantly on lower estimates for events below 500k copies/ml.

The median time to resolution of EBV viraemia post-rituximab was 21 days (IQR 19-124 days) in 5 patients with >500k copies/ml (one patient was treated for late onset persistent symptomatic clonal gammopathy, despite fall in EBV levels). No significant adverse events were noted in the treated group. Nine patients had a persistent low level EBV viraemia detectable at last follow up. All patients who underwent AHSCT were alive as of April 2017.

DISCUSSION:

MS as an autoimmune disorder (AD) is theorised to have generally similar underlying pathophysiological immune dysregulation mechanisms[18–22] relative to other chronic autoimmune conditions. Epstein-Barr virus (EBV) is increasingly implicated in the pathogenesis of MS by virtue of epidemiological and serological evidence, impaired CD8+T cell immune responses to EBV and possible underlying genetic susceptibility for autoimmunity (with EBV encoded protein interactions), as recently described by Harley et al and others[1,23–25].

Generally, EBV-R related LPD has been reported in both allogeneic (allo-HSCT) and solid organ transplants treated with immunosuppressive therapy, often with a significant impact on organ function and overall survival[26–30]. It is observed that reduced intensity allo-HSCT for malignant haematological conditions using alemtuzumab have a relatively lower

overall risk of LPD compared to ATG based treatments, possibly mediated by more effective pan-B & T cell lymphodepletion. In contrast, ATG primarily affects the T-cell repertoire with delayed EBV specific CD8+ T cell recovery[31]. Clinically significant endogenous viral infections including EBV following ATG conditioned AHSCT for severe ADs such as Crohn's disease and systemic sclerosis is increasingly recognised, but the development of lymphoproliferative disorders (LPD) remains rare in these ADs[13,32,33]. Nash et al[32] concerningly reported 2 deaths (1 MS and 1 systemic sclerosis patient) from EBV related LPD in 56 ATG conditioned AHSCT for autoimmune diseases. Additionally, EBV associated haemophagocytosis in ATG-AHSCT for ADs have also been reported[34], with one resulting in death of the patient[35].

Our report of suspected EBV related LPDs (~10%) in MS-AHSCT group is relatively higher than published reports with allo-HSCTs (4.5-7%)[36,37] and our own centre's unpublished T-cell depleted allo-HSCT experience (6.5%); possibly a reflection of underlying immunopathological state of MS itself[38]. This is further corroborated by the fact that similar LPD risk has not been observed in other ADs, e.g. Crohn's disease, treated with ATG -AHSCT in this centre. Another example from our centre's experience of severe aplastic anaemia (n-40) treated with ATG/ciclosporin, only 52% (n-21/40) developed EBV-R (unpublished data) and none had LPD or required any treatment, suggesting that the problem may not be ATG specific.

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

Our study's observation of significant persistent neurological events (with no evidence of new MS disease activity) associated with clonal gammopathy suggest a potentially new clinical syndrome, described for the first time in ATG conditioned AHSCTs in MS and possibly induced by clonal B cell dysregulation following EBV-R. It could be hypothesised that any remaining EBV infected latent B cells, surviving despite high doses of cyclophosphamide (given with mobilisation and conditioning in MS ASHCT)[8] and compounded by depletion of CD8+ T cells by ATG, may serve as potential source for EBV escape while interacting abnormally within the host immune micro-environment[39] and leading to rise in M-protein, LPD and neuro-inflammatory insults in some of the MS patients post AHSCT. Reports of lower incidence of EBV-R and LPD in another commonly used protocol for MS-AHSCT using BEAM-ATG (Ricardo Saccardi, Carregi University Hospital, Florence; personal communication) may reflect the greater myeloablative effect of BEAM chemotherapy which could further deplete the residual B cell pool and thus lower potential for EBV proliferation. It is plausible that dose of rATG is critical, given we have not seen similar reports from other centres where less rATG doses were given for MS-AHSCT (range between 5.0-6.5 mg/Kg; personal communication) but there seems to be some variability in prospective serial EBV monitoring in these patients.

The clinical threshold for EBV viral load as a significant risk factor for post-transplant LPD is widely debated. This study reports peak EBV viral load >500k copies/ml post AHSCT is significantly associated with probable LPD and neurological events in MS patients with high sensitivity (85.5%) and specificity (82.5%) (p-0.004) (Fig 3, ROC curve). Our ROC curve estimates are potentially limited by the relatively small number of events analysed but this has consistently been useful in our MS-AHSCT experience for predicting clinical events with high EBV load. Our EBV PCR assay has been validated against the recently defined standard WHO reference method (i.e. 10 copies/ml=10 IU/ml EBV DNA) [16] and thus this EBV threshold for pre-emptive treatment with Rituximab, can potentially be applied in relevant clinical context in other centres using similar validated essays. Rituximab treatment delivered good overall response in our symptomatic patients, with resolution of EBV related clinical symptoms and no subsequent viral or bacterial infections at last follow up. The role of prophylactic or pre-transplant rituximab in MS-AHSCT is also a potential area of interest in reducing risk of EBV-LPD in stem cell transplants as observed by Burns et al[36] and future randomised studies are required to investigate its potential benefit.

320

321

322

323

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

Our study limitations include its retrospective nature and that no suspected LPD patients had histological confirmation, mainly related to patient refusal or technical difficulties. Seven MS patients were lost to follow up for EBV monitoring following discharge, which limits the

EBV related clinical events with previous DMT exposure in MS patients. In conclusion, symptomatic EBV reactivation increases risk of neurological sequelae and LPD in MS-AHSCT. Regular monitoring for rising EBV viraemia, as recommended by Snowden et al [10] and Muraro et al [11], and serum electrophoresis for M-protein should be considered in the first 3 months post-AHSCT for MS. We recommend persistent high EBV viraemia > 500k DNA copies/ml as potential trigger for consideration of pre-emptive anti-CD20 therapy and potentially reduce associated morbidity.

findings of this study. Additionally, our numbers were too small to identify any association of

341	Confl	ict of interest: The authors declare no competing financial interests as below:
342	1.	Varun Mehra- no competing financial interests
343	2.	Elijah Rhone- no competing financial interests
344	3.	Stefani Widya- no competing financial interests
345	4.	Mark Zuckerman- no competing financial interests
346	5.	Victoria Potter- no competing financial interests
347	6.	Kavita Raj- no competing financial interests
348	7.	Austin Kulasekararaj- no competing financial interests
349	8.	Donal McLornan- no competing financial interests
350	9.	Hugues de Lavallade- no competing financial interests
351	10.	Nana Benson-Quarm- no competing financial interests
352	11.	Christina Lim- no competing financial interests
353	12.	Sarah Ware- no competing financial interests
354	13.	Malur Sudhanva- no competing financial interests
355	14.	Omar Malik- no competing financial interests
356	15.	Richard Nicholas- no competing financial interests
357	16.	Paolo A Muraro- no competing financial interests
358	17.	Judith Marsh- no competing financial interests
359	18.	Ghulam J Mufti- no competing financial interests
360	19.	Eli Silber- no competing financial interests
361	20.	Antonio Pagliuca- no competing financial interests
362	21.	Majid A. Kazmi- no competing financial interests
363		
364		
365		
366		
367		

Acknowledgements: To our patients and their families and carers in supporting this study.

340

REFERENCES

2	7	Λ
	1	u
_	•	~

- 1. Pender MP, Burrows SR. Epstein–Barr virus and multiple sclerosis: potential opportunities for
- immunotherapy. Clinical & Translational Immunology **2014**; 3:e27. Available at:
- 373 http://doi.wiley.com/10.1038/cti.2014.25.
- 2. Reich DS, Lucchinetti CF, Calabresi PA. Multiple Sclerosis. New England Journal of Medicine 2018;
- 375 378:169–180. Available at: http://www.nejm.org/doi/10.1056/NEJMra1401483.
- 376 3. Compston A, Coles A. Multiple sclerosis. The Lancet 2008; 372:1502–1517. Available at:
- 377 http://linkinghub.elsevier.com/retrieve/pii/S0140673608616207.
- Lublin FD, Reingold SC, Cohen JA, et al. Defining the clinical course of multiple sclerosis: The 2013
- 379 revisions. Neurology **2014**; 83:278–286. Available at:
- 380 http://www.neurology.org/cgi/doi/10.1212/WNL.000000000000560.
- 5. Comi G, Radaelli M, Soelberg Sørensen P. Evolving concepts in the treatment of relapsing multiple
- 382 sclerosis. The Lancet **2017**; 389:1347–1356. Available at:
- https://linkinghub.elsevier.com/retrieve/pii/S0140673616323881.
- Muraro PA, Douek DC, Packer A, et al. Thymic output generates a new and diverse TCR repertoire
- after autologous stem cell transplantation in multiple sclerosis patients. The Journal of Experimental
- 386 Medicine **2005**; 201:805–816. Available at: http://www.jem.org/lookup/doi/10.1084/jem.20041679.
- Muraro PA, Robins H, Malhotra S, et al. T cell repertoire following autologous stem cell transplantation
- for multiple sclerosis. J Clin Invest **2014**; 124:1168–1172. Available at:
- 389 http://www.ncbi.nlm.nih.gov/pubmed/24531550.
- 390 8. Cull G, Hall D, Fabis-Pedrini M, et al. Lymphocyte reconstitution following autologous stem cell
- 391 transplantation for progressive MS. Multiple Sclerosis Journal Experimental, Translational and
- 392 Clinical **2017**; 3:205521731770016. Available at: https://doi.org/10.1177/2055217317700167.
- 393 9. Sormani MP, Muraro PA, Schiavetti I, et al. Autologous hematopoietic stem cell transplantation in
- 394 multiple sclerosis. Neurology **2017**; 88:2115–2122. Available at:
- 395 http://www.neurology.org/lookup/doi/10.1212/WNL.000000000003987.
- 396 10. Snowden JA, Saccardi R, Allez M, et al. Haematopoietic SCT in severe autoimmune diseases:
- 397 updated guidelines of the European Group for Blood and Marrow Transplantation. Bone Marrow
- Transplantation **2012**; 47:770–790. Available at: http://www.nature.com/articles/bmt2011185.
- 399 11. Muraro PA, Pasquini M, Atkins HL, et al. Long-term Outcomes After Autologous Hematopoietic Stem
- 400 Cell Transplantation for Multiple Sclerosis. JAMA Neurology **2017**; 74:459. Available at:
- 401 http://archneur.jamanetwork.com/article.aspx?doi=10.1001/jamaneurol.2016.5867.
- 402 12. Burt RK, Balabanov R, Snowden JA, Sharrack B, Oliveira MC, Burman J. Non-myeloablative
- hematopoietic stem cell transplantation (HSCT) is superior to disease modifying drug (DMD) treatment
- in highly active Relapsing Remitting Multiple Sclerosis (RRMS): interim results of the Multiple Sclerosis

- 405 International Stem cell Transp. Neurology **2018**; 90. Available at:
- http://n.neurology.org/content/90/15_Supplement/S36.004.abstract.
- 407 13. Daikeler T, Tichelli A, Passweg J. Complications of autologous hematopoietic stem cell transplantation
- for patients with autoimmune diseases. Pediatric Research **2012**; 71:439–444. Available at:
- 409 http://www.nature.com/doifinder/10.1038/pr.2011.57.
- 410 14. Snowden JA, Badoglio M, Labopin M, et al. Evolution, trends, outcomes, and economics of
- hematopoietic stem cell transplantation in severe autoimmune diseases. Blood Advances **2017**;
- 412 1:2742–2755. Available at:
- http://www.bloodadvances.org/lookup/doi/10.1182/bloodadvances.2017010041.
- 414 15. Patel S, Zuckerman M, Smith M. Real-time quantitative PCR of Epstein–Barr virus BZLF1 DNA using
- the LightCycler. Journal of Virological Methods **2003**; 109:227–233. Available at:
- 416 http://linkinghub.elsevier.com/retrieve/pii/S0166093403000764.
- 417 16. Semenova T, Lupo J, Alain S, et al. Multicenter Evaluation of Whole-Blood Epstein-Barr Viral Load
- Standardization Using the WHO International Standard. Journal of Clinical Microbiology **2016**; 54:1746
- 419 LP-1750. Available at: http://jcm.asm.org/content/54/7/1746.abstract.
- 420 17. Styczynski J, van der Velden W, Fox CP, et al. Management of Epstein-Barr Virus infections and post-
- 421 transplant lymphoproliferative disorders in patients after allogeneic hematopoietic stem cell
- transplantation: Sixth European Conference on Infections in Leukemia (ECIL-6) guidelines.
- 423 Haematologica **2016**; 101:803–811. Available at:
- http://www.haematologica.org/cgi/doi/10.3324/haematol.2016.144428.
- 425 18. Dejaco C, Duftner C, Grubeck-Loebenstein B, Schirmer M. Imbalance of regulatory T cells in human
- 426 autoimmune diseases. Immunology **2006**; 117:289–300. Available at:
- 427 http://doi.wiley.com/10.1111/j.1365-2567.2005.02317.x.
- 428 19. Arellano G, Acuña E, Reyes LI, et al. Th1 and Th17 Cells and Associated Cytokines Discriminate
- among Clinically Isolated Syndrome and Multiple Sclerosis Phenotypes. Frontiers in immunology **2017**;
- 430 8:753. Available at: http://journal.frontiersin.org/article/10.3389/fimmu.2017.00753/full.
- 431 20. Jha S, Srivastava SY, Brickey WJ, et al. The Inflammasome Sensor, NLRP3, Regulates CNS
- Inflammation and Demyelination via Caspase-1 and Interleukin-18. Journal of Neuroscience **2010**;
- 433 30:15811–15820. Available at: http://www.jneurosci.org/cgi/doi/10.1523/JNEUROSCI.4088-10.2010.
- 434 21. Beynon V, Quintana FJ, Weiner HL. Activated Human CD4+CD45RO+ Memory T-Cells Indirectly
- Inhibit NLRP3 Inflammasome Activation through Downregulation of P2X7R Signalling. PLoS ONE
- 436 **2012**; 7:e39576. Available at: https://dx.plos.org/10.1371/journal.pone.0039576.
- 437 22. Fernández-Menéndez S, Fernández-Morán M, Fernández-Vega I, Pérez-Álvarez A, Villafani-Echazú J.
- Epstein–Barr virus and multiple sclerosis. From evidence to therapeutic strategies. Journal of
- 439 Neurological Sciences **2016**; 361:213–219. Available at:
- 440 https://www.clinicalkey.com.ezsecureaccess.balamand.edu.lb/service/content/pdf/watermarked/1-s2.0-
- 441 S0022510X16300132.pdf?locale=en_US.

- 23. Pender MP. The Essential Role of Epstein-Barr Virus in the Pathogenesis of Multiple Sclerosis. The
- 443 Neuroscientist **2011**; 17:351–367. Available at:
- 444 http://journals.sagepub.com/doi/10.1177/1073858410381531.
- 445 24. Ascherio A, Munger KL. Environmental risk factors for multiple sclerosis. Part I: The role of infection.
- 446 Annals of Neurology **2007**; 61:288–299. Available at: http://doi.wiley.com/10.1002/ana.21117.
- 447 25. Harley JB, Chen X, Pujato M, et al. Transcription factors operate across disease loci, with EBNA2
- implicated in autoimmunity. Nature Genetics **2018**; 50:699–707. Available at:
- 449 http://www.nature.com/articles/s41588-018-0102-3.
- Loren AW, Porter DL, Stadtmauer EA, Tsai DE. Post-transplant lymphoproliferative disorder: a review.
- Bone Marrow Transplantation **2003**; 31:145–155. Available at:
- 452 http://www.nature.com/articles/1703806.
- 453 27. Dotti G, Fiocchi R, Motta T, et al. Lymphomas occurring late after solid-organ transplantation: influence
- of treatment on the clinical outcome. Transplantation **2002**; 74:1095–102. Available at:
- http://www.ncbi.nlm.nih.gov/pubmed/12438953.
- 456 28. Nagle SJ, Reshef R, Tsai DE. Posttransplant Lymphoproliferative Disorder in Solid Organ and
- Hematopoietic Stem Cell Transplantation. Clinics in Chest Medicine **2017**; 38:771–783. Available at:
- https://linkinghub.elsevier.com/retrieve/pii/S0272523117300874.
- 459 29. Meijer E, Dekker AW, Weersink AJL, Rozenberg-Arska M, Verdonck LF. Prevention and treatment of
- 460 epstein-barr virus-associated lymphoproliferative disorders in recipients of bone marrow and solid
- organ transplants. British Journal of Haematology **2002**; 119:596–607. Available at:
- 462 http://dx.doi.org/10.1046/j.1365-2141.2002.03887.x.
- 463 30. Dierickx D, Habermann TM. Post-Transplantation Lymphoproliferative Disorders in Adults. New
- 464 England Journal of Medicine **2018**; 378:549–562. Available at:
- 465 http://dx.doi.org/10.1056/NEJMra1702693.
- 466 31. Meij P. Impaired recovery of Epstein-Barr virus (EBV)--specific CD8+ T lymphocytes after partially T-
- depleted allogeneic stem cell transplantation may identify patients at very high risk for progressive
- EBV reactivation and lymphoproliferative disease. Blood **2003**; 101:4290–4297. Available at:
- http://www.bloodjournal.org/cgi/doi/10.1182/blood-2002-10-3001.
- 470 32. Nash R a, Dansey R, Storek J, et al. Epstein-Barr virus-associated posttransplantation
- 471 lymphoproliferative disorder after high-dose immunosuppressive therapy and autologous CD34-
- selected hematopoietic stem cell transplantation for severe autoimmune diseases. Biology of blood
- and marrow transplantation: journal of the American Society for Blood and Marrow Transplantation
- **2003**; 9:583–91. Available at: http://www.ncbi.nlm.nih.gov/pubmed/14506660.
- 475 33. Daikeler T, Tzankov A, Hoenger G, et al. Minimal T-cell requirements for triggering
- 476 haemophagocytosis associated with Epstein–Barr virus-driven B-cell proliferation: a clinical case
- 477 study. Annals of the Rheumatic Diseases **2011**; 70:1338 LP-1339. Available at:
- http://ard.bmj.com/content/70/7/1338.abstract.

479 34. Daikeler T, Tzankov A, Hoenger G, et al. Minimal T-cell requirements for triggering 480 haemophagocytosis associated with Epstein-Barr virus-driven B-cell proliferation: a clinical case study. 481 Annals of the Rheumatic Diseases 2011; 70:1338–1339. Available at: 482 http://ard.bmj.com/cgi/doi/10.1136/ard.2010.139246. 483 35. Brinkman DMC, de Kleer IM, ten Cate R, et al. Autologous stem cell transplantation in children with 484 severe progressive systemic or polyarticular juvenile idiopathic arthritis: Long-term followup of a 485 prospective clinical trial. Arthritis & Rheumatism 2007; 56:2410-2421. Available at: 486 http://doi.wiley.com/10.1002/art.22656. 487 36. Burns DM, Rana S, Martin E, et al. Greatly reduced risk of EBV reactivation in rituximab-experienced 488 recipients of alemtuzumab-conditioned allogeneic HSCT. Bone Marrow Transplantation 2016; 51:825-489 832. Available at: http://www.nature.com/articles/bmt201619. 490 37. van Esser JWJ. Epstein-Barr virus (EBV) reactivation is a frequent event after allogeneic stem cell 491 transplantation (SCT) and quantitatively predicts EBV-lymphoproliferative disease following T-cell-492 depleted SCT. Blood 2001; 98:972-978. Available at: 493 http://www.bloodjournal.org/cgi/doi/10.1182/blood.V98.4.972. 494 38. Tørring C, Andreasen C, Gehr N, Bjerg L, Petersen T, Höllsberg P. Higher incidence of Epstein-Barr 495 virus-induced lymphocyte transformation in multiple sclerosis. Acta Neurologica Scandinavica 2014; 496 130:90-96. Available at: http://doi.wiley.com/10.1111/ane.12249. 497 39. Martinez OM, Krams SM. The Immune Response to Epstein Barr Virus and Implications for 498 Posttransplant Lymphoproliferative Disorder. Transplantation 2017; 101:2009–2016. Available at: 499 http://insights.ovid.com/crossref?an=00007890-201709000-00018. 500 501 502 503 504 505 506 507 508 509 510 511 512 513 514 515 516 517 518

Table 1: Baseline patient characteristics and EBV related clinical events according to peak EBV DNA-aemia burden.

Baseline characteristics (n-36)		Patient Groups according to peak EBV DNA in copies/ml (n-29)	0 - 100,000	100,001	>500,000
Median age at time of AHSCT in years (range)	43.5 (36– 47)	No of patients (%)	16 (55.2)	3 (10.3)	10 (34.5
Gender Male Female	19 (52.8%) 17 (47.2%)	M-Protein (n)	11	0	7
Disease Type (n; %) Relapsing Remitting MS Secondary Progressive MS Primary Progressive MS	22 (61.1%) 10 (27.8%) 4 (11.1%)	Median EBV DNA log value at peak (IQR)	4.8 (3.5-4.8)	5.5 (N/A)	6.25 (6.1-6.9
Median number of previous DMT (range)	2 (0 – 6)	Median number of prior DMTs	2	3	2
Previous use of high efficacy DMT (n) Natalizumab Alemtuzumab Both	22 8 6	Symptomatic EBV (n)	1	0	7
Median EDSS (range)	6.0 (2.5 – 8.0)	LPD diagnosis (CT/Biopsy) (n)	0	0	3 by CT alor
Median follow up post AHSCT in days (range)	436 (188 – 785)	Neuro/autoimmune complications (n)	0	0	3
No. of patients with prior EBV exposure (n; %)	36 (100%)	Treated with Rituximab (n)	0	0	6
No. of patients with detectable EBV post AHSCT (n; %)	29 (80.5%)	Confirmed EBV resolution at last follow up (n)	7	2	7
No. of patients lost to long term follow up (n; %)	7 (19.5%)	Detectable EBV DNA at last follow up (n)	9	1	3
Median Time to EBV detection post AHSCT in days (IQR)	30 (23-46)	Median time for EBV resolution (IQR in days)	67 days (44-155)	47 days (N/A)	63 days (45 - 170
Median Time to peak EBV DNA levels in days (IQR)	32 (31-53)	Median Time to peak EBV DNA levels in days (IQR)	40 days (25-85)	30 days (N/A)	39 days (32-43)

524 Abbreviations:

520

- 525 AHSCT- Autologous Haematopoietic Stem cell transplant; CT- computed tomography;
- 526 DMT- Disease modifying therapy; EBV- Epstein Barr virus; EDSS- Kurtzke Expanded
- 527 Disability Status Scale; IQR- Interquartile range; LPD- Lymphoproliferative disease; M-
- 528 Protein: Monoclonal paraprotein or gammopathy; MS- Multiple Sclerosis

Figure Legends

Figure 1: Trend of Lymphocyte count recovery following ATG in MS patients.

Legend: This figure shows trends of lymphocyte count from baseline to recovery post AHSCT for MS patients. Majority of patients had normal baseline lymphocyte counts pre-HSCT and became lymphopenic post ATG with counts recovering towards d+28 and >85% of patients recovered counts by d+56, with some overshooting from their baseline, possibly reflective of EBV related lymphoproliferation in some of these patients.

AHSCT: Autologous Haematopoietic Stem Cell Transplantation; ATG: Anti-Thymocyte Globulin; d+: Days post AHSCT; MS: Multiple Sclerosis.

Figure 2: Paraprotein, EBV & Lymphocyte trends in two MS patients with significant neurological sequelae post AHSCT.

Legend: This figure demonstrates trends of EBV copies (log), paraprotein levels (g/lt) and Lymphocyte levels (counts x10^6/ml) in two MS patients with significant neurological symptoms following EBV reactivation. Both patients had significant EBV viraemia (log>5.2 or >500,000 copy number) and developed significant paraproteinaemia, which was only noted after persistent unexplained neurological symptoms. The trend reversed following anti-CD20 (rituximab) therapy, with limited recovery in neurological symptoms.

D: Days post AHSCT; EBV-R: Epstein Barr Virus reactivation; MS: Multiple sclerosis; AHSCT: Autologous Haematopoietic Stem Cell Transplants.

Figure 3: ROC curve estimates for peak EBV viraemia levels and significant clinical events in MS post AHSCT.

Legend: ROC curve demonstrating significant correlation between high EBV levels and clinical events (LPD & neurological events) in MS-AHSCT patients, with highest sensitivity and specificity noted with peak EBV viraemia of >500,000 copies/ml (p-0.0004).

EBV: Epstein Barr Virus; LPD- Lymphoproliferative disorder; MS-AHSCT: Multiple Sclerosis patients treated with autologous haematopoietic stem cell transplants; ROC: receiver operating characteristics

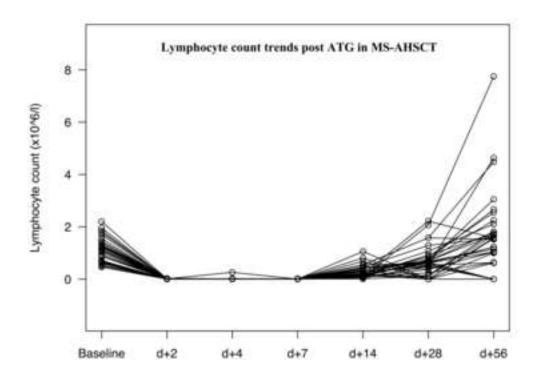
- 22 -

Table 1: Baseline patient characteristics and EBV related clinical events according to peak EBV DNA-aemia burden.

Baseline characteristics (n-36)		Patient Groups according to peak EBV DNA in copies/ml (n-29)	0 - 100,000	100,001 - 500,000	>500,000
Median age at time of AHSCT in years (range)	43.5 (36– 47)	No of patients (%)	16 (55.2)	3 (10.3)	10 (34.5)
Gender Male Female	19 (52.8%) 17 (47.2%)	M-Protein (n)	11	0	7
Disease Type (n; %) Relapsing Remitting MS Secondary Progressive MS Primary Progressive MS	22 (61.1%) 10 (27.8%) 4 (11.1%)	Median EBV DNA log value at peak (IQR)	4.8 (3.5-4.8)	5.5 (N/A)	6.25 (6.1-6.9)
Median number of previous DMT (range)	2 (0 – 6)	Median number of prior DMTs	2	3	2
Previous use of high efficacy DMT (n) Natalizumab Alemtuzumab Both	22 8 6	Symptomatic EBV (n)	1	0	7
Median EDSS (range)	6.0 (2.5 – 8.0)	LPD diagnosis (CT/Biopsy) (n)	0	0	3 by CT alone
Median follow up post AHSCT in days (range)	436 (188 – 785)	Neuro/autoimmune complications (n)	0	0	3
No. of patients with prior EBV exposure (n; %)	36 (100%)	Treated with Rituximab (n)	0	0	6
No. of patients with detectable EBV post AHSCT (n; %)	29 (80.5%)	Confirmed EBV resolution at last follow up (n)	7	2	7
No. of patients lost to long term follow up (n; %)	7 (19.5%)	Detectable EBV DNA at last follow up (n)	9	1	3
Median Time to EBV detection post AHSCT in days (IQR)	30 (23-46)	Median time for EBV resolution (IQR in days)	67 days (44-155)	47 days (N/A)	63 days (45 - 170)
Median Time to peak EBV DNA levels in days (IQR)	32 (31-53)	Median Time to peak EBV DNA levels in days (IQR)	40 days (25-85)	30 days (N/A)	39 days (32-43)

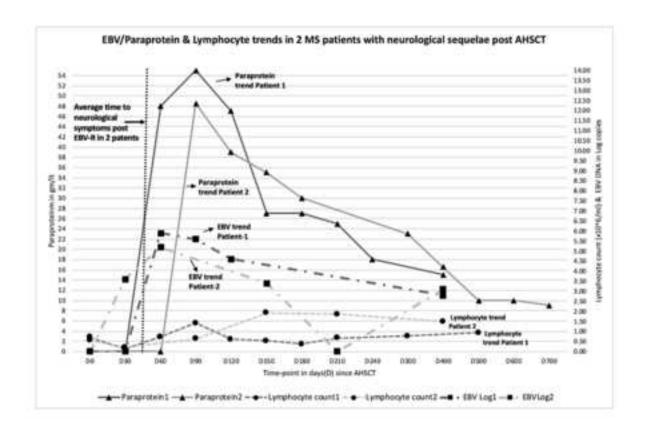
AHSCT- Autologous Haematopoietic Stem cell transplant; CT- computed tomography; DMT- Disease modifying therapy; EBV- Epstein Barr virus; EDSS- Kurtzke Expanded Disability Status Scale; IQR- Interquartile range; LPD- Lymphoproliferative disease; M-Protein: Monoclonal paraprotein or gammopathy; MS- Multiple Sclerosis

Figure 1: Trend of Lymphocyte count recovery following ATG conditioned AHSCT in MS patients.



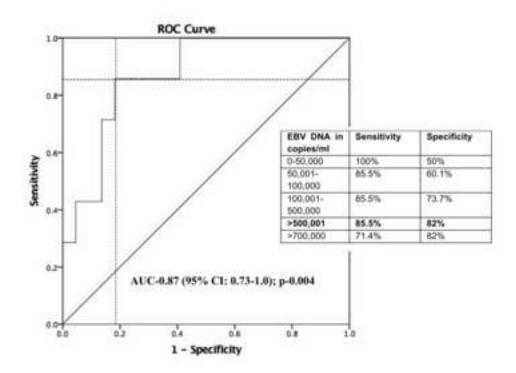
AHSCT: Autologous Haematopoietic Stem Cell Transplantation; ATG: Anti-Thymocyte Globulin; d+: Days post AHSCT; MS: Multiple Sclerosis.

Figure 2. Paraprotein, EBV & Lymphocyte trends in two MS patients with significant neurological sequelae post AHSCT



D: Days post AHSCT; EBV-R: Epstein Barr Virus reactivation; MS: Multiple sclerosis; AHSCT: Autologous Haematopoietic Stem Cell Transplants.

Figure 3: ROC curve estimates for peak EBV viraemia levels and significant clinical events in MS post AHSCT.



EBV: Epstein Barr Virus; LPD- Lymphoproliferative disorder; MS-AHSCT: Multiple Sclerosis patients treated with autologous haematopoietic stem cell transplants; ROC: receiver operating characteristics Supplemental -Case Vignettes

EBV and Monoclonal Gammopathy of Clinical Significance in Autologous

Stem Cell Transplantation for Multiple Sclerosis.

Running Title: EBV complications in Auto-HSCT for MS

SUPPLEMENTARY:

Case vignettes of 2 MS patients describing EBV-related significant paraproteinaemia

and neurological sequelae.

Patient 1

43-year-old female with Relapsing Remitting Multiple Sclerosis (RRMS), previously treated

with natalizumab and three courses of alemtuzumab but continued to have breakthrough

disease. She had a relatively mild baseline disability with an Expanded Disability Severity

Scale (EDSS) of 2.5. She had an uncomplicated inpatient stay for the Autologous

Haematopoeitic Stem cell transplant (AHSCT) procedure and was discharged on day 15 post-

transplant. A blood test on day 26 demonstrated Epstein-Barr Virus (EBV) reactivation

(155,845 copies/ml). A repeat test on day 34 showed an increase in copy number to 638,634

copies/ml. She was asymptomatic, and the plan was to monitor this closely. On day 37 post-

transplant she developed a significant deterioration in strength in the right lower limb and on

day 42 she developed pyrexia and was admitted to a local hospital. She was found to have

CMV reactivation which was treated with IV ganciclovir as well as ongoing EBV reactivation

and she remained an inpatient for 4 weeks. She did not receive rituximab at the local centre

but on repeat testing at day 145 the copy number was vastly reduced at 2,355 copies/ml. A

high IgM paraproteinaemia was first detected at day 92 post-transplant (48.58g/L). This had

not been routinely monitored previously. This paraproteinaemia was initially felt to be

asymptomatic and was monitored closely, slowly improving over time. A CT scan was

performed which demonstrated a single 1.7cm right hilar lymph node requiring observation.

A bone marrow aspirate showed a small excess of plasma cells (5-9%) on aspirate with no other significant findings.

The EBV reactivation initially settled at 6 months post-transplant. At one-year post transplant she had a persistent IgM paraprotein (23g/L) and her right leg weakness had continued to progress with her EDSS now at 5.0. There was also a mild recurrence of EBV (DNA at 1,335 copies/ml). It was considered that as the onset of the right leg weakness had coincided with the high level of EBV reactivation and paraproteinaemia that these factors may have driven a peripheral neuropathy. She was treated with rituximab 375mg/m² weekly for 4 weeks at 19 months post-transplant following which EBV DNA again became undetectable and the paraprotein reduced to 9g/L. Despite this, there was no improvement in strength of the right leg. Nerve conduction studies subsequently confirmed an L5-S1 radiculopathy but without a generalised polyneuropathy neuropathy. She has had no new or active demyelinating lesions on MRI head and spine post-transplant that would account for these symptoms and the slowly progressive nature of the weakness does not suggest an MS relapse. The cause of the weakness is likely an atypical IgM paraprotein associated radiculo-neuropathy was strongly suspected.

Patient 2

42-year-old male with Secondary Progressive Multiple Sclerosis (SPMS), previously treated with interferon and copaxone which were discontinued due to side effects and ongoing relapses, respectively. He was then treated with natalizumab for 2 years but continued to progress and was offered HSCT. He had a moderate level of baseline disability with an EDSS of 5.5 (walking at least 100m unaided). The transplant procedure was complicated by neutropenic sepsis which was treated successfully, and he was discharged on day 13 post-transplant with no new neurological symptoms. He was readmitted on day 17 post-transplant with pyrexia and rigors. Blood cultures grew *Stenotrophomonas maltophilia* and he was treated for line sepsis with appropriate antibiotics and fully recovered. An EBV viraemia of

58,324 copies/ml was detected for the first time on this admission. On day 22 he had developed new urinary urgency, diplopia and significant deterioration in mobility. This was felt to represent either a pseudorelapse driven by infection or a true relapse and an MRI was performed which demonstrated no new demyelinating lesions and no other significant pathology. A repeat EBV DNA assessment on day 28 demonstrated a significant rise in EBV viraemia to >10 million copies/ml (log change).

His neurological symptoms persisted and on day 34 he began spiking temperatures again; antibiotics were restarted but blood and urine cultures came back negative, but his EBV viraemia had risen to over 39 million copies/ml. He continued to experience intermittent pyrexia, which possibly was attributed to his EBV viraemia. No evidence of lymphadenopathy was noted during this period. Due to significant neurological decline, he was consequently commenced on rituximab 375mg/m² weekly for 4 weeks on day 38 post-transplant. Testing on day 51 demonstrated a reduction in EBV viraemia to DNA of 2.2 million copies/ml and on day 52, a significant IgG kappa paraproteinaemia (45.6 g/L) was identified. This had not been routinely monitored previously. It was considered that this degree of paraproteinemia and resulting hyperviscosity may have been a driver of his neurological symptoms. These values continued to improve over time with further doses of rituximab and the EBV vireamia was <100,000 copies/ml and the IgG kappa paraprotein down to 8.63 g/L by Day 87. However, due to persistence of these markers as well as his ongoing neurological symptoms, he was given a single plasma exchange on day 80 that was of minimal symptomatic benefit.

He had ongoing rehabilitation, including a short admission in a specialist neuro-rehabilitation ward. neurorehabilitation unit. At one year review he still required bilateral support to walk, putting his EDSS at 6.5. A repeat MRI at 12 months post-transplant was again stable with no new demyelinating lesions. This patient demonstrated significant deterioration in his condition post-transplant and although there may be an element of disease progression, we suspect this was in large part driven by EBV viraemia and associated paraproteinaemia/hyperviscosity.

The EBV viraemia was undetectable at the last follow up, although there was ongoing paraproteinaemia with an IgG kappa of 15 g/L.

- 1 EBV and Monoclonal Gammopathy of Clinical Significance in Autologous
- **2 Stem Cell Transplantation for Multiple Sclerosis.**
- 3 Running Title: EBV complications in Auto-HSCT for MS
- 4 Varun Mehra*, Elijah Rhone*, Stefani Widya, Mark Zuckerman, Victoria Potter, Kavita Raj,
- 5 Austin Kulasekararaj, Donal McLornan, Hugues de Lavallade, Nana Benson-Quarm, Christina
- 6 Lim, Sarah Ware, Malur Sudhanva, Omar Malik, Richard Nicholas, Paolo A Muraro, Judith
- 7 Marsh, Ghulam J Mufti, Eli Silber, Antonio Pagliuca and Majid A. Kazmi
- ^{*}These authors contributed equally to this work as 1st Authors.
- 11 Key Points:

8

10

16

- 12 EBV reactivation is common post-transplant with ATG for multiple sclerosis (MS),
- with significant lymphoproliferative & neurological sequelae associated with rising
- M-protein. Serial monitoring of EBV & M-protein is recommended post-transplant, as
- is pre-emptive anti-CD20 therapy with EBV DNA >500,000 copies/ml.
- 17 Corresponding authors:
- 1. Dr Varun Mehra; Varun.Mehra@nhs.net; +442032995378
- 19 Alternate Corresponding Author:
- 20 2. Dr Majid Kazmi; majidkazmi@nhs.net; +442071882757
- 22 **Key Words:**
- 23 Multiple Sclerosis; Autologous Hematopoietic Stem Cell Transplantation, Epstein-
- 24 Barr Virus Infection; Monoclonal Gammopathy; Post-transplant Lymphoproliferative
- 25 **Disorder**

Author Affiliations:

- **1. Dr Varun Mehra***; Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
- 2. Dr Elijah Rhone*: Department of Neurology, King's College Hospital NHS Foundation Trust,
 Denmark Hill, London, United Kingdom
 - 3. Stefani Widya: GKT School of Medical Education, Kings College London University, London
 - **4. Dr Mark Zuckerman:** Department of Virology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **5. Dr Victoria Potter:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **6. Dr Kavita Raj:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom **AND** Department of Haematology, Guy's and St. Thomas' NHS Foundation Trust, London, United Kingdom
 - **7. Dr Austin Kulasekararaj:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **8. Dr Donal McLornan**: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom **AND** Department of Haematology, Guy's and St. Thomas' NHS Foundation Trust, London, United Kingdom
 - **9. Dr Hugues de Lavallade**: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **10. Nana Benson-Quarm:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **11. Christina Lim:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **12. Sarah Ware:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **13. Dr Malur Sudhanva:** Department of Virology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - **14. Dr Omar Malik:** Department of Neurology, Imperial College Healthcare, London, United Kingdom
 - **15. Dr Richard Nicholas:** Department of Neurology, Imperial College Healthcare, London, United Kingdom
 - **16. Professor Paolo A. Muraro:** Department of Neurology, Imperial College Healthcare, London, United Kingdom **AND** Department of Neuroimmunology, Imperial College London, London, United Kingdom
 - **17. Professor Judith Marsh:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - **18. Professor Ghulam J. Mufti:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - **19. Dr Eli Silber:** Department of Neurology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - **20. Professor Antonio Pagliuca:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - **21. Dr Majid Kazmi**: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom **AND** Department of Haematology, Guy's and St. Thomas' NHS Foundation Trust, London, United Kingdom

78	Confl	ict of interest: The authors declare no competing financial interests as below:
79	1.	Varun Mehra- no competing financial interests
80	2.	Elijah Rhone- no competing financial interests
81	3.	Stefani Widya- no competing financial interests
82	4.	Mark Zuckerman- no competing financial interests
83	5.	Victoria Potter- no competing financial interests
84	6.	Kavita Raj- no competing financial interests
85	7.	Austin Kulasekararaj- no competing financial interests
86	8.	Donal McLornan- no competing financial interests
87	9.	Hugues de Lavallade- no competing financial interests
88	10.	Nana Benson-Quarm- no competing financial interests
89	11.	Christina Lim- no competing financial interests
90	12.	Sarah Ware- no competing financial interests
91	13.	Malur Sudhanva- no competing financial interests
92	14.	Omar Malik- no competing financial interests
93	15.	Richard Nicholas- no competing financial interests
94	16.	Paolo A Muraro- no competing financial interests
95	17.	Judith Marsh- no competing financial interests
96	18.	Ghulam J Mufti- no competing financial interests
97	19.	Eli Silber- no competing financial interests
98	20.	Antonio Pagliuca- no competing financial interests
99	21.	Majid A. Kazmi- no competing financial interests
100		
101		
102		
103		
104		
105		
106		
107		
108		

Abstract

Introduction

Autologous haematopoietic stem cell transplantation (AHSCT) with anti-thymocyte globulin (ATG) conditioning as treatment of active multiple sclerosis (MS) is rapidly increasing across Europe (EBMT registry data 2017). Clinically significant Epstein Barr virus reactivation (EBV-R) following AHSCT with ATG for severe autoimmune conditions is an under-recognised complication relative to T-cell deplete transplants performed for haematological diseases. This retrospective study reports EBV-R associated significant clinical sequelae in MS patients undergoing AHSCT with rabbit ATG.

Methods

Retrospective data was analysed for 36 consecutive MS-AHSCT patients at Kings College Hospital, London. All patients routinely underwent weekly EBV DNA PCR monitoring and serum electrophoresis for monoclonal gammopathy (MG or M-protein). EBV-R with rising EBV viral load, M-protein and associated clinical sequelae were captured from clinical records.

Results

All patients had evidence of rising EBV DNA-emia, including 7 who were lost to long term follow-up, with a number of them developing high EBV viral load & associated lymphoproliferative disorder (LPD). Nearly 72% (n-18/29) developed de-novo MG, some with significant neurological consequences with high M-protein and EBV-R. Six patients required anti-CD20 therapy (rituximab) with complete resolution of EBV related symptoms. Receiver operating characteristics (ROC) estimated a peak EBV viraemia of >500,000 DNA copies/ml

133	correlated with high sensitivity (85.5%) & specificity (82.5%) (AUC-0.87; p-0.004) in
134	predicting EBV-R related significant clinical events.
135	Conclusion
136	Symptomatic EBV reactivation increases risk of neurological sequelae and LPD in MS-
137	AHSCT. We recommend regular monitoring for EBV and serum electrophoresis for MG be
138	mandated in MS patients in the first 3 months post AHSCT
139	
140	
141	
142	
143	
144	
145	
146	
147	
148	
149	
150	

INTRODUCTION:

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

Multiple sclerosis (MS) is a chronic autoimmune, inflammatory, demyelinating disease of the central nervous system[1]-[2], with a relapsing-remitting (RRMS) presentation in the majority of patients at diagnosis. Recovery from relapses may be complete or partial[3]-[4]. After a variable period of time, people with RRMS may develop a more progressive disability accumulation with or without superimposed relapses; termed secondary progressive multiple sclerosis (SPMS). A minority experience progressive disability from the onset of disease, termed primary progressive multiple sclerosis (PPMS)[4]. A number of immunomodulatory disease modifying therapies (DMTs) are currently licensed for treatment of RRMS with an aim of reducing number of relapses and accrual of disability, although with variable efficacy[5]. Since 1996, Autologous Hematopoietic Stem Cell Transplantation (AHSCT) has been a novel approach for MS management, using immunoablation followed by immunomodulation mechanisms, with evidence of significant suppression of inflammatory activity and qualitative changes in the reconstituted immune system (immune reset theory)[6-8]. AHSCT appears most effective for MS patients with evidence of inflammatory activity on MRI, younger age, a shorter disease duration, low to moderate disability levels (Expanded Disability Status Scale [EDSS] <6 or up to 6.5 if recent progression) and failure of at least 1 highly active DMT (natalizumab or alemtuzumab) with no significant comorbidities[9-11]. Recently reported preliminary results of randomised MIST study[12] found AHSCT to be superior to standard disease modifying therapy (DMT) for RRMS with respect to both

treatment failure and disability progression.

172

173

174

175

176

177

178

179

180

181

171

subsequent opportunistic infections following However, risk rise in immunosuppressive therapies remain a potential concern[13]. MS patients undergoing AHSCT have often been exposed to a number of immunomodulating DMTs; the addition of immunosuppressive rabbit anti-thymocyte globulin (rATG) to their conditioning regimen may confer a higher risk of viral reactivation in these patients. The number of AHSCTs performed for MS is rising significantly in Europe[14] and as more centres perform AHSCT for this indication, it is increasingly important to recognise the unique problems faced by these patients post AHSCT. This retrospective study reports for the first time, EBV-R associated neurological sequelae and lymphoproliferative disorder (LPD) in MS patients undergoing rATG conditioned AHSCT in our centre.

183

184

185

186

187

188

189

190

182

METHODS

Patients and procedures

Data was collected retrospectively on 36 consecutive MS patients undergoing AHSCT between February 2012 and February 2017 at Kings College Hospital, London. Peripheral blood stem cells were collected following standard mobilisation strategy consisting of cyclophosphamide 4g/m² over 2 days and granulocyte colony-stimulating factor for 7 days. Thirty-five (97%) patients were conditioned using standard protocol of cyclophosphamide

(50mg/kg for 4 days) and rATG (2.5mg/kg/day for 3 days) for in-vivo lymphodepletion, followed by stem cell infusion. One patient was conditioned with carmustine/etoposide/cytarabine/melphalan regimen along with an equivalent dose of rATG (BEAM-ATG) prior to stem cell infusion. The median CD34 stem cell dose returned was 7.17 x10^6/kg (range 4.0-17.1x10^6/kg).

196

197

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

191

192

193

194

195

Prior exposure to EBV was assessed by serological evidence (EBV viral capsid antigen; VCA IgG). EBV DNA load monitoring was performed on whole blood samples by standardised quantitative real-time polymerase chain reaction (RT-PCR) using Rotor-GeneTM (Qiagen) assay of EBV BZLF1 DNA. This assay was, adapted from our published assay using LightCycler (Roche)[15] and since been validated against the recently published WHO standard, with our lab's EBV DNA quantification of 10 copies/ml considered equivalent to 10 IU/ml DNA reported with the WHO reference method[16]. -EBV-R was defined as rising EBV DNA load of >10 copies/millilitre (ml) detected on two consecutive tests based on our assay sensitivity. Symptomatic EBV-R was captured by peak blood EBV DNA load, presence of B symptoms (defined by presence of either unexplained weight loss, recurrent fever, night sweats); which was in-turn defined by clinical, radiological and/or histological evidence based on recent ECIL-6 guidelines[17]. In addition, significant 'clinical events' were also defined as new & persistent organ dysfunction (e.g. neurological events) temporally associated with rising EBV viraemia in MS patients. Serum protein electrophoresis was routinely tested around 3 months post HSCT, as part of our institutional practice, with immunoglobulin subclasses identified by immunofixation electrophoresis. Patient outcomes were assessed at last follow up as of April 2017.

Statistics

The database of transplants and outcomes was built in Microsoft Excel 2016 and statistical analyses were performed using IBM SPSS Statistics version 24.0. Patient characteristics are presented as medians (with inter-quartile ranges; IQR) for data with non-normal distribution. Comparisons of baseline characteristics used Mann-Whitney U test, Fisher's exact test, or Chi-squared test for trend as appropriate. Receiver Operating characteristics (ROC) curve was obtained correlating LPD and clonal gammopathy associated clinical events with rising EBV viraemia (copies/ml).

RESULTS

Baseline characteristics are presented in **Table 1**. Most MS patients (88.9%) had RRMS phenotype with median of 2 (range 0-3) DMTs prior to AHSCT. Twenty-two MS patients had prior exposure to natalizumab and 7 were treated with Alemtuzumab (6 patients received both). All 36 (100%) MS patients were serologically positive for EBV VCA IgG pre-treatment, indicating prior EBV exposure and had detectable EBV DNA post-AHSCT. Seven MS patients were lost to long-term follow up for EBV monitoring. The median time to first EBV DNA detection post-transplant was 30 days (IQR 23-46 days). EBV DNA levels peaked at a median of 32 days post-transplant (IQR 31-53 days). All MS patients had normal baseline lymphocyte counts (median) pre-HSCT with a median time of 46 days (range 14-404 days) to lymphocyte recovery (defined by total lymphocyte count >1.0x106/ml) following AHSCT (**See Figure 1**). A high proportion (86%; n-25/29) of the MS patients in active follow-up

recovered lymphocyte counts around D56 with a median lymphocyte count of 1.56 (10⁶ cells/ml); Four patients remained lymphopenic at last follow up.

238

239

240

241

242

243

244

245

246

247

248

249

250

251

237

236

All patients were stratified into following 3 groups according to peak rise/burden of EBV DNAaemia (copies/ml): <100,000 (<100k) copies/ml, 100,001-500,00 (100k-500k) copies/ml and >500,000 (>500k) copies/ml to identify any specific thresholds for clinically significant events related to rising EBV-R (Table 1). The majority of patients (76%) with rising EBV viral load >100k copies/ml were routinely screened by computed tomographic (CT) scans to assess for evidence of LPD, in line with our institutional policy. One third (34.5%) of patients developed peak EBV viraemia of >500k copies/ml. Eight patients (27.6%) developed symptomatic EBV-R; defined as persistent fever, lymphadenopathy and/or B symptoms. Of these 8 patients, only 1 (12.5%) had a peak EBV viraemia <100kDNA copies/ml with the remaining 7 (87.5%) patients having a peak EBV viraemia of >500k copies/ml. Three patients with rising EBV viraemia >500k copies/ml had findings consistent with probable LPD on CT imaging; however, none had definitive histological diagnosis. Three MS patients had worsening neurological symptoms concurrent with rising EBV viraemia >500k copies/ml and clonal gammopathy, as described below.

253

254

255

252

Interestingly, we also observed frequent de novo monoclonal gammopathy (MG or M-protein) in 18 MS patients (62.4%) following rising EBV viraemia; the majority (n-16) of whom

developed IgG subtype and the remaining 2 developed IgA and IgM M-protein. Concerningly two of these patients developed clinically significant M-Protein burden; one patient with IgG Kappa M-protein of 45.6g/L developed hyper-viscosity and neurological symptoms mimicking MS relapse, requiring plasma exchange. Another patient developed significant lumbosacral radiculopathy with rising EBV-R and high IgM lambda M-protein (IgM 48.5g/L) (see supplementary case vignettes). Figure 2 highlights the association of neurological symptom onset following rising EBV viraemia (log copies), falling lymphocyte counts (x 106/ml) with significant rise in M-protein (gm/lt) levels post AHSCT. A third patient developed painful lower limb paraesthesia following rising EBV viraemia >500k copies/ml, although did not have any M-protein detected. Their symptoms persisted at last follow up despite no evidence of MS related new disease activity.

Six patients were treated with anti-CD20 antibody, rituximab (375mg/m2 weekly up_to 4 weeks), due to clinical severity of EBV reactivations and leading to 4 reduction in EBV viral load and concurrent improvement/resolution of EBV related symptoms. ROC curve analysis (Figure 3) confirmed EBV viraemia of >500k copies/ml correlated with high sensitivity (85.5%) and specificity (82.5%) (AUC-0.87; 95%Cl-0.73-1.0; p-0.004) in predicting significant EBV related clinical events (evidence of LPD and/or neurological symptoms) that may require treatment with rituximab. The sensitivity dropped significantly on lower estimates for events below 500k copies/ml.

The median time to resolution of EBV viraemia post-rituximab was 21 days (IQR 19-124 days) in 5 patients with >500,000-k copies/ml (one patient was treated for late onset persistent symptomatic clonal gammopathy, despite fall in EBV levels). No significant adverse events were noted in the treated group. Nine patients had a persistent low level EBV viraemia detectable at last follow up. All patients who underwent AHSCT were alive as of April 2017.

DISCUSSION:

MS as an autoimmune disorder (AD) has is theorised to have generally similar underlying pathophysiological immune dysregulation mechanisms[18–22] relative to other chronic autoimmune conditions. Epstein-Barr virus (EBV) is increasingly implicated in the pathogenesis of MS by virtue of epidemiological and serological evidence, impaired CD8+ T cell immune responses to EBV and possible underlying genetic susceptibility for autoimmunity (with EBV encoded protein interactions), as recently described by Harley et al and others[1,23–25].

Generally, EBV-R related LPD has been reported in both allogeneic (allo-HSCT) and solid organ transplants treated with immunosuppressive therapy, often with a significant impact on organ function and overall survival[26–30]. It is observed that reduced intensity allo-HSCT for malignant haematological conditions using alemtuzumab have a relatively lower overall

risk of LPD compared to ATG based treatments, possibly mediated by more effective pan-B & T cell lymphodepletion. In contrast, ATG primarily affects the T-cell repertoire with delayed EBV specific CD8+ T cell recovery[31]. Clinically significant endogenous viral infections including EBV following ATG conditioned AHSCT for severe ADs such as Crohn's disease and seystemic sectors is increasingly recognised, but the development of lymphoproliferative disorders (LPD) remains rare in these ADs[13,32,33]. Nash et al[32] concerningly reported 2 deaths (1 MS and 1 systemic sclerosis patient) from EBV related LPD in 56 ATG conditioned AHSCT for autoimmune diseases. Additionally, EBV associated haemophagocytosis in ATG-AHSCT for ADs have also been reported[34], with one resulting in death of the patient[35].

Our report of suspected EBV related LPDs (~10%) in MS-AHSCT group is relatively higher than published reports with allo-HSCTs (4.5-7%)[36,37] and our own centre's unpublished T-cell depleted allo-HSCT experience (6.5%); possibly a reflection of underlying immunopathological state of MS itself[38]. This is further corroborated by the fact that similar LPD risk has not been observed in other ADds, e.g. Crohn's disease, treated with ATG -AHSCT in this centre. Another example from oueur centre's experience of in-severe aplastic anaemiaa (n-40), a type of AD causing severe bone marrow failure(n-40) treated & treated with ATG/ciclosporin, n; only 52% (n-21/40) patients developed EBV-R (unpublished data) and nhone had LPD or required any treatment, supporting the notion that it may not just be

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

316

Our study's observation of significant persistent neurological events (with no evidence of new MS disease activity) associated with clonal gammopathy, suggest a potentially new clinical syndrome, described for the first time in ATG conditioned AHSCTs in MS_and, possibly induced by clonal B cell dysregulation following EBV-R. It could be hypothesised that any remaining EBV infected latent B cells, which may still have survivedsurviving despite high doses of cyclophosphamide (given with mobilisation and conditioning in MS ASHCT)[8] and compounded by depletion of CD8+ T cells by ATG, may serve as potential source for EBV escape while interacting abnormally within the host immune micro-environment[39] and leading to rise in M-protein, LPD and neuro-inflammatory insults in some of the MS patients post AHSCT. Reports of lower incidence of EBV-R and LPD in another commonly used protocol for MS-AHSCT using BEAM-ATG (Ricardo Saccardi, Carregi University Hospital, Florence; personal communication) may reflect the greater myeloablative effect of BEAM chemotherapy which could further deplete the residual B cell pool and thus lower potential for EBV proliferation. It is plausible that dose of rATG is critical, given we have not seen similar reports from other centres where less rATG doses were given for MS-AHSCT (range between 5.0-6.5 mg/Kg; personal communication), but there seems to be some variability in prospective serial EBV monitoring in these patients.

The clinical threshold for EBV viral load as a significant risk factor for post-transplant LPD is widely debated. This study reports peak EBV viral load >500k copies/ml post AHSCT is significantly associated with probable LPD and neurological events in MS patients with high sensitivity (85.5%) and specificity (82.5%) (p-0.004) (Fig 3, ROC curve). Our ROC curve estimates are potentially limited by the relatively small number of events analysed but this has consistently been useful in our MS-AHSCT experience for predicting clinical events with high EBV load. Our EBV PCR assay has been validated against the recently defined standard WHO reference method (i.e. 10 copiesy/ml=10 IU/ml EBV DNA) [16] and thus this EBV threshold for pre-emptive treatment with Rituximab, can potentially be applied in relevant clinical context in other centres using similar validated essays. Rituximab treatment delivered good overall response in our symptomatic patients, with resolution of EBV related clinical symptoms and no subsequent viral or bacterial infections at last follow up. The role of prophylactic or pre-transplant rituximab in MS-AHSCT is also a potential area of interest in reducing risk of EBV-LPD in stem cell transplants as observed by Burns et al[36] and future randomised studies are required to investigate its potential benefit.

351

352

353

354

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

Our study limitations include its retrospective nature and that no suspected LPD patients had histological confirmation, mainly related to patient refusal or technical difficulties. Seven MS patients were lost to follow up for EBV monitoring following discharge, which limits the

findings of this study. Additionally, our numbers were too small to identify any association of EBV related clinical events with previous DMT exposure in MS patients. In conclusion, symptomatic EBV reactivation increases risk of neurological sequelae and LPD in MS-AHSCT. Regular monitoring for rising EBV viraemia, as recommended by Snowden et al [10] and Muraro et al [11], and serum electrophoresis for M-protein should be considered mandated in the first 3 months post-AHSCT for MS_ and Wwe recommend persistent high EBV viraemia > 500k DNA copies/ml as potential trigger for consideration of pre-emptive anti-CD20 therapy and potentially reduce associated morbidity. **Acknowledgements:** To our patients and their families and carers in supporting this study.

376 **REFERENCES**

2	7	7
3	/	/

- 1. Pender MP, Burrows SR. Epstein–Barr virus and multiple sclerosis: potential opportunities for
- immunotherapy. Clinical & Translational Immunology **2014**; 3:e27. Available at:
- 380 http://doi.wiley.com/10.1038/cti.2014.25.
- Reich DS, Lucchinetti CF, Calabresi PA. Multiple Sclerosis. New England Journal of Medicine 2018;
- 382 378:169–180. Available at: http://www.nejm.org/doi/10.1056/NEJMra1401483.
- 383 3. Compston A, Coles A. Multiple sclerosis. The Lancet **2008**; 372:1502–1517. Available at:
- http://linkinghub.elsevier.com/retrieve/pii/S0140673608616207.
- Lublin FD, Reingold SC, Cohen JA, et al. Defining the clinical course of multiple sclerosis: The 2013
- 386 revisions. Neurology **2014**; 83:278–286. Available at:
- 387 http://www.neurology.org/cgi/doi/10.1212/WNL.0000000000000560.
- 5. Comi G, Radaelli M, Soelberg Sørensen P. Evolving concepts in the treatment of relapsing multiple
- 389 sclerosis. The Lancet **2017**; 389:1347–1356. Available at:
- 390 https://linkinghub.elsevier.com/retrieve/pii/S0140673616323881.
- 391 6. Muraro PA, Douek DC, Packer A, et al. Thymic output generates a new and diverse TCR repertoire
- 392 after autologous stem cell transplantation in multiple sclerosis patients. The Journal of Experimental
- 393 Medicine **2005**; 201:805–816. Available at: http://www.jem.org/lookup/doi/10.1084/jem.20041679.
- 394 7. Muraro PA, Robins H, Malhotra S, et al. T cell repertoire following autologous stem cell transplantation
- for multiple sclerosis. J Clin Invest **2014**; 124:1168–1172. Available at:
- 396 http://www.ncbi.nlm.nih.gov/pubmed/24531550.
- 397 8. Cull G, Hall D, Fabis-Pedrini M, et al. Lymphocyte reconstitution following autologous stem cell
- 398 transplantation for progressive MS. Multiple Sclerosis Journal Experimental, Translational and
- 399 Clinical **2017**; 3:205521731770016. Available at: https://doi.org/10.1177/2055217317700167.
- 400 9. Sormani MP, Muraro PA, Schiavetti I, et al. Autologous hematopoietic stem cell transplantation in
- 401 multiple sclerosis. Neurology **2017**; 88:2115–2122. Available at:
- 402 http://www.neurology.org/lookup/doi/10.1212/WNL.000000000003987.
- 403 10. Snowden JA, Saccardi R, Allez M, et al. Haematopoietic SCT in severe autoimmune diseases:
- 404 updated guidelines of the European Group for Blood and Marrow Transplantation. Bone Marrow
- Transplantation **2012**; 47:770–790. Available at: http://www.nature.com/articles/bmt2011185.
- 406 11. Muraro PA, Pasquini M, Atkins HL, et al. Long-term Outcomes After Autologous Hematopoietic Stem
- 407 Cell Transplantation for Multiple Sclerosis. JAMA Neurology **2017**; 74:459. Available at:
- 408 http://archneur.jamanetwork.com/article.aspx?doi=10.1001/jamaneurol.2016.5867.
- 409 12. Burt RK, Balabanov R, Snowden JA, Sharrack B, Oliveira MC, Burman J. Non-myeloablative
- 410 hematopoietic stem cell transplantation (HSCT) is superior to disease modifying drug (DMD) treatment
- 411 in highly active Relapsing Remitting Multiple Sclerosis (RRMS): interim results of the Multiple Sclerosis

- International Stem cell Transp. Neurology **2018**; 90. Available at:
- http://n.neurology.org/content/90/15_Supplement/S36.004.abstract.
- 13. Daikeler T, Tichelli A, Passweg J. Complications of autologous hematopoietic stem cell transplantation
- for patients with autoimmune diseases. Pediatric Research **2012**; 71:439–444. Available at:
- 416 http://www.nature.com/doifinder/10.1038/pr.2011.57.
- 417 14. Snowden JA, Badoglio M, Labopin M, et al. Evolution, trends, outcomes, and economics of
- hematopoietic stem cell transplantation in severe autoimmune diseases. Blood Advances **2017**;
- 419 1:2742–2755. Available at:
- http://www.bloodadvances.org/lookup/doi/10.1182/bloodadvances.2017010041.
- 421 15. Patel S, Zuckerman M, Smith M. Real-time quantitative PCR of Epstein–Barr virus BZLF1 DNA using
- 422 the LightCycler. Journal of Virological Methods **2003**; 109:227–233. Available at:
- 423 http://linkinghub.elsevier.com/retrieve/pii/S0166093403000764.
- 424 16. Semenova T, Lupo J, Alain S, et al. Multicenter Evaluation of Whole-Blood Epstein-Barr Viral Load
- Standardization Using the WHO International Standard. Journal of Clinical Microbiology **2016**; 54:1746
- 426 LP-1750. Available at: http://jcm.asm.org/content/54/7/1746.abstract.
- 427 17. Styczynski J, van der Velden W, Fox CP, et al. Management of Epstein-Barr Virus infections and post-
- 428 transplant lymphoproliferative disorders in patients after allogeneic hematopoietic stem cell
- transplantation: Sixth European Conference on Infections in Leukemia (ECIL-6) guidelines.
- 430 Haematologica **2016**; 101:803–811. Available at:
- http://www.haematologica.org/cgi/doi/10.3324/haematol.2016.144428.
- 432 18. Dejaco C, Duftner C, Grubeck-Loebenstein B, Schirmer M. Imbalance of regulatory T cells in human
- 433 autoimmune diseases. Immunology **2006**; 117:289–300. Available at:
- 434 http://doi.wiley.com/10.1111/j.1365-2567.2005.02317.x.
- 435 19. Arellano G, Acuña E, Reyes LI, et al. Th1 and Th17 Cells and Associated Cytokines Discriminate
- among Clinically Isolated Syndrome and Multiple Sclerosis Phenotypes. Frontiers in immunology **2017**;
- 437 8:753. Available at: http://journal.frontiersin.org/article/10.3389/fimmu.2017.00753/full.
- 438 20. Jha S, Srivastava SY, Brickey WJ, et al. The Inflammasome Sensor, NLRP3, Regulates CNS
- Inflammation and Demyelination via Caspase-1 and Interleukin-18. Journal of Neuroscience **2010**;
- 440 30:15811–15820. Available at: http://www.jneurosci.org/cgi/doi/10.1523/JNEUROSCI.4088-10.2010.
- 441 21. Beynon V, Quintana FJ, Weiner HL. Activated Human CD4+CD45RO+ Memory T-Cells Indirectly
- Inhibit NLRP3 Inflammasome Activation through Downregulation of P2X7R Signalling. PLoS ONE
- 443 **2012**; 7:e39576. Available at: https://dx.plos.org/10.1371/journal.pone.0039576.
- 444 22. Fernández-Menéndez S, Fernández-Morán M, Fernández-Vega I, Pérez-Álvarez A, Villafani-Echazú J.
- 445 Epstein–Barr virus and multiple sclerosis. From evidence to therapeutic strategies. Journal of
- 446 Neurological Sciences **2016**; 361:213–219. Available at:
- 447 https://www.clinicalkey.com.ezsecureaccess.balamand.edu.lb/service/content/pdf/watermarked/1-s2.0-
- 448 S0022510X16300132.pdf?locale=en_US.

- 449 23. Pender MP. The Essential Role of Epstein-Barr Virus in the Pathogenesis of Multiple Sclerosis. The
- 450 Neuroscientist **2011**; 17:351–367. Available at:
- 451 http://journals.sagepub.com/doi/10.1177/1073858410381531.
- 452 24. Ascherio A, Munger KL. Environmental risk factors for multiple sclerosis. Part I: The role of infection.
- 453 Annals of Neurology **2007**; 61:288–299. Available at: http://doi.wiley.com/10.1002/ana.21117.
- 454 25. Harley JB, Chen X, Pujato M, et al. Transcription factors operate across disease loci, with EBNA2
- implicated in autoimmunity. Nature Genetics **2018**; 50:699–707. Available at:
- 456 http://www.nature.com/articles/s41588-018-0102-3.
- Loren AW, Porter DL, Stadtmauer EA, Tsai DE. Post-transplant lymphoproliferative disorder: a review.
- 458 Bone Marrow Transplantation **2003**; 31:145–155. Available at:
- 459 http://www.nature.com/articles/1703806.
- 460 27. Dotti G, Fiocchi R, Motta T, et al. Lymphomas occurring late after solid-organ transplantation: influence
- of treatment on the clinical outcome. Transplantation **2002**; 74:1095–102. Available at:
- http://www.ncbi.nlm.nih.gov/pubmed/12438953.
- 463 28. Nagle SJ, Reshef R, Tsai DE. Posttransplant Lymphoproliferative Disorder in Solid Organ and
- Hematopoietic Stem Cell Transplantation. Clinics in Chest Medicine **2017**; 38:771–783. Available at:
- https://linkinghub.elsevier.com/retrieve/pii/S0272523117300874.
- 466 29. Meijer E, Dekker AW, Weersink AJL, Rozenberg-Arska M, Verdonck LF. Prevention and treatment of
- 467 epstein–barr virus-associated lymphoproliferative disorders in recipients of bone marrow and solid
- organ transplants. British Journal of Haematology **2002**; 119:596–607. Available at:
- 469 http://dx.doi.org/10.1046/j.1365-2141.2002.03887.x.
- 470 30. Dierickx D, Habermann TM. Post-Transplantation Lymphoproliferative Disorders in Adults. New
- 471 England Journal of Medicine **2018**; 378:549–562. Available at:
- 472 http://dx.doi.org/10.1056/NEJMra1702693.
- 473 31. Meij P. Impaired recovery of Epstein-Barr virus (EBV)--specific CD8+ T lymphocytes after partially T-
- depleted allogeneic stem cell transplantation may identify patients at very high risk for progressive
- EBV reactivation and lymphoproliferative disease. Blood **2003**; 101:4290–4297. Available at:
- 476 http://www.bloodjournal.org/cgi/doi/10.1182/blood-2002-10-3001.
- 477 32. Nash R a, Dansey R, Storek J, et al. Epstein-Barr virus-associated posttransplantation
- lymphoproliferative disorder after high-dose immunosuppressive therapy and autologous CD34-
- selected hematopoietic stem cell transplantation for severe autoimmune diseases. Biology of blood
- 480 and marrow transplantation : journal of the American Society for Blood and Marrow Transplantation
- **2003**; 9:583–91. Available at: http://www.ncbi.nlm.nih.gov/pubmed/14506660.
- 482 33. Daikeler T, Tzankov A, Hoenger G, et al. Minimal T-cell requirements for triggering
- 483 haemophagocytosis associated with Epstein–Barr virus-driven B-cell proliferation: a clinical case
- 484 study. Annals of the Rheumatic Diseases **2011**; 70:1338 LP-1339. Available at:
- http://ard.bmj.com/content/70/7/1338.abstract.

486 34. Daikeler T, Tzankov A, Hoenger G, et al. Minimal T-cell requirements for triggering 487 haemophagocytosis associated with Epstein-Barr virus-driven B-cell proliferation: a clinical case study. 488 Annals of the Rheumatic Diseases 2011; 70:1338–1339. Available at: 489 http://ard.bmj.com/cgi/doi/10.1136/ard.2010.139246. 490 35. Brinkman DMC, de Kleer IM, ten Cate R, et al. Autologous stem cell transplantation in children with 491 severe progressive systemic or polyarticular juvenile idiopathic arthritis: Long-term followup of a 492 prospective clinical trial. Arthritis & Rheumatism 2007; 56:2410-2421. Available at: 493 http://doi.wiley.com/10.1002/art.22656. 494 36. Burns DM, Rana S, Martin E, et al. Greatly reduced risk of EBV reactivation in rituximab-experienced 495 recipients of alemtuzumab-conditioned allogeneic HSCT. Bone Marrow Transplantation 2016; 51:825-496 832. Available at: http://www.nature.com/articles/bmt201619. 497 37. van Esser JWJ. Epstein-Barr virus (EBV) reactivation is a frequent event after allogeneic stem cell 498 transplantation (SCT) and quantitatively predicts EBV-lymphoproliferative disease following T-cell-499 depleted SCT. Blood 2001; 98:972-978. Available at: 500 http://www.bloodjournal.org/cgi/doi/10.1182/blood.V98.4.972. 501 38. Tørring C, Andreasen C, Gehr N, Bjerg L, Petersen T, Höllsberg P. Higher incidence of Epstein-Barr 502 virus-induced lymphocyte transformation in multiple sclerosis. Acta Neurologica Scandinavica 2014; 503 130:90-96. Available at: http://doi.wiley.com/10.1111/ane.12249. 504 39. Martinez OM, Krams SM. The Immune Response to Epstein Barr Virus and Implications for 505 Posttransplant Lymphoproliferative Disorder. Transplantation 2017; 101:2009–2016. Available at: 506 http://insights.ovid.com/crossref?an=00007890-201709000-00018. 507 508 509 510 511 512 513 514 515 516 517 518 519 520 521 522 523 524 525 526

Baseline characteristics (Patient Groups according to peak EBV DNA in copies/ml (n-29)	0 - 100,000	100,001 - 500,000	>500,000	
Median age at time of AHSCT in years (range)	43.5 (36– 47)	No of patients (%)	16 (55.2)	3 (10.3)	10 (34.5)
Gender Male Female	19 (52.8%) 17 (47.2%)	M-Protein (n)	11	0	7
Disease Type (n; %) Relapsing Remitting MS Secondary Progressive MS Primary Progressive MS	22 (61.1%) 10 (27.8%) 4 (11.1%)	Median EBV DNA log value at peak (IQR)	4.8 (3.5-4.8)	5.5 (N/A)	6.25 (6.1-6.9)
Median number of previous DMT (range)	2 (0 – 6)	Median number of prior DMTs	2	3	2
Previous use of high efficacy DMT (n) Natalizumab Alemtuzumab Both	22 8 6	Symptomatic EBV (n)	1	0	7
Median EDSS (range)	6.0 (2.5 – 8.0)	LPD diagnosis (CT/Biopsy) (n)	0	0	3 by CT alone
Median follow up post AHSCT in days (range)	436 (188 – 785)	Neuro/autoimmune complications (n)	0	0	3
No. of patients with prior EBV exposure (n; %)	36 (100%)	Treated with Rituximab (n)	0	0	6
No. of patients with detectable EBV post AHSCT (n; %)	29 (80.5%)	Confirmed EBV resolution at last follow up (n)	7	2	7
No. of patients lost to long term follow up (n; %)	7 (19.5%)	Detectable EBV DNA at last follow up (n)	9	1	3
Median Time to EBV detection post AHSCT in days (IQR)	30 (23-46)	Median time for EBV resolution (IQR in days)	67 days (44-155)	47 days (N/A)	63 days (45 - 170)
Median Time to peak EBV DNA levels in days (IQR)	32 (31-53)	Median Time to peak EBV DNA levels in days (IQR)	40 days (25-85)	30 days (N/A)	39 days (32-43)

531 Abbreviations:

- 532 AHSCT- Autologous Haematopoietic Stem cell transplant; CT- computed tomography; DMT-
- 533 Disease modifying therapy; EBV- Epstein Barr virus; EDSS- Kurtzke Expanded Disability
- 534 Status Scale; IQR- Interquartile range; LPD- Lymphoproliferative disease; M-Protein:
- 535 Monoclonal paraprotein or gammopathy; MS- Multiple Sclerosis

Figure Legends

538 Figure 1: Trend of Lymphocyte count recovery following ATG in MS patients.

Legend: This figure shows trends of lymphocyte count from baseline to recovery post AHSCT for MS patients. Majority of patients had normal baseline lymphocyte counts pre-HSCT and became lymphopenic post ATG with counts recovering towards d+28 and >85% of patients recovered counts by d+56, with some overshooting from their baseline, possibly reflective of EBV related lymphoproliferation in some of these patients.

AHSCT: Autologous Haematopoietic Stem Cell Transplantation; ATG: Anti-Thymocyte Globulin; d+: Days post AHSCT; MS: Multiple Sclerosis.

Figure 2: Paraprotein, EBV & Lymphocyte trends in two MS patients with significant neurological sequelae post AHSCT.

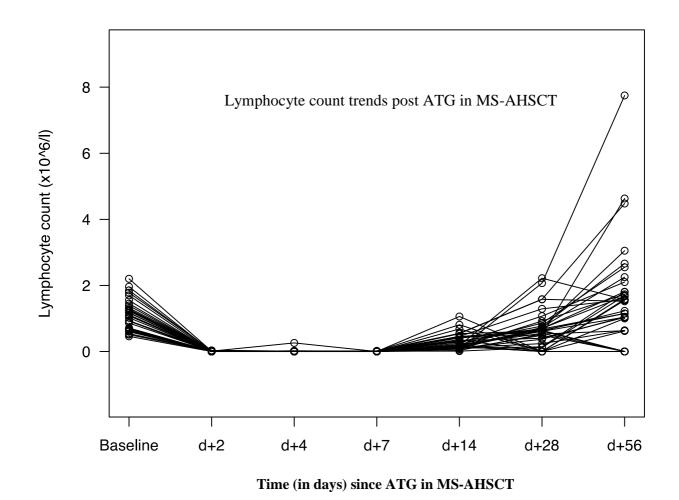
Legend: This figure demonstrates trends of EBV copies (log), paraprotein levels (g/lt) and Lymphocyte levels (counts x10^6/ml) in two MS patients with significant neurological symptoms following EBV reactivation. Both patients had significant EBV viraemia (log>5.2 or >500,000 copy number) and developed significant paraproteinaemia, which was only noted after persistent unexplained neurological symptoms. The trend reversed following anti-CD20 (rituximab) therapy, with limited recovery in neurological symptoms.

D: Days post AHSCT; EBV-R: Epstein Barr Virus reactivation; MS: Multiple sclerosis; AHSCT: Autologous Haematopoietic Stem Cell Transplants.

Figure 3: ROC curve estimates for peak EBV viraemia levels and significant clinical events in MS post AHSCT.

Legend: ROC curve demonstrating significant correlation between high EBV levels and clinical events (LPD & neurological events) in MS-AHSCT patients, with highest sensitivity and specificity noted with peak EBV viraemia of >500,000 copies/ml (p-0.0004).

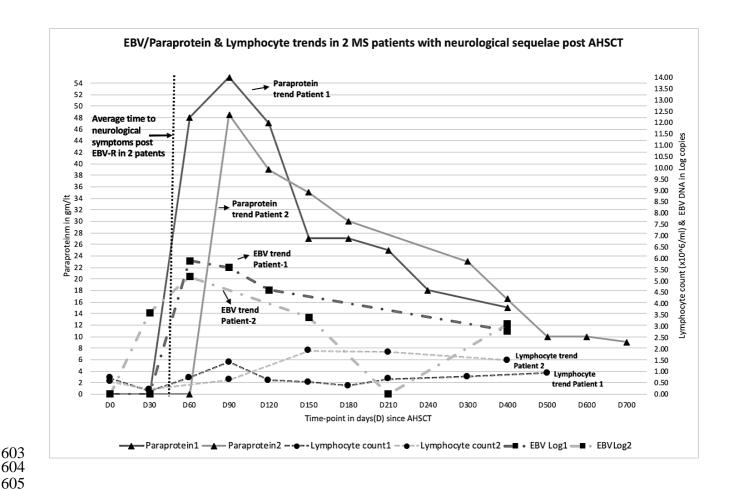
EBV: Epstein Barr Virus; LPD- Lymphoproliferative disorder; MS-AHSCT: Multiple Sclerosis patients treated with autologous haematopoietic stem cell transplants; ROC: receiver operating characteristics



Abbreviations:

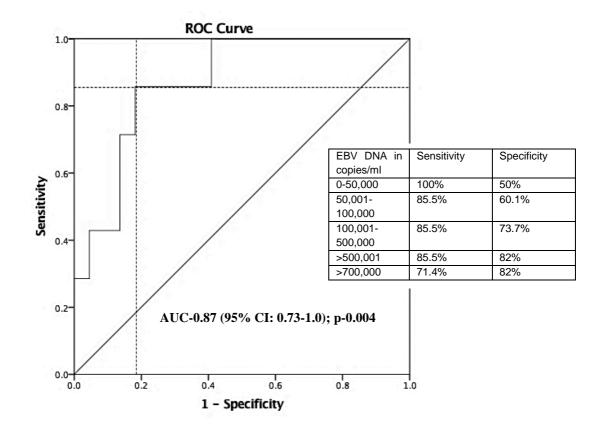
AHSCT: Autologous Haematopoietic Stem Cell Transplantation; ATG: Anti-Thymocyte Globulin; d+: Days post AHSCT; MS: Multiple Sclerosis.

Figure 2. Paraprotein, EBV & Lymphocyte trends in two MS patients with significant neurological sequelae post AHSCT



Abbreviations:

D: Days post AHSCT; EBV-R: Epstein Barr Virus reactivation; MS: Multiple sclerosis; AHSCT: Autologous Haematopoietic Stem Cell Transplants.



Abbreviations:

EBV: Epstein Barr Virus; LPD- Lymphoproliferative disorder; MS-AHSCT: Multiple Sclerosis patients treated with autologous haematopoietic stem cell transplants; ROC: receiver operating characteristics

- **EBV and Monoclonal Gammopathy of Clinical Significance in Autologous** 1
- **Stem Cell Transplantation for Multiple Sclerosis.** 2
- 3 **Running Title**: EBV complications in Auto-HSCT for MS
- 4 Varun Mehra*, Elijah Rhone*, Stefani Widya, Mark Zuckerman, Victoria Potter, Kavita Raj,
- 5 Austin Kulasekararaj, Donal McLornan, Hugues de Lavallade, Nana Benson-Quarm, Christina
- 6 Lim, Sarah Ware, Malur Sudhanva, Omar Malik, Richard Nicholas, Paolo A Muraro, Judith
- 7 Marsh, Ghulam J Mufti, Eli Silber, Antonio Pagliuca and Majid A. Kazmi
- 9 *These authors contributed equally to this work as 1st Authors.
- 11 Key Points:

10

16

- 12 EBV reactivation is common post-transplant with ATG for multiple sclerosis (MS),
- 13 with significant lymphoproliferative & neurological sequelae associated with rising
- 14 M-protein. Serial monitoring of EBV & M-protein is recommended post-transplant, as
- 15 is pre-emptive anti-CD20 therapy with EBV DNA >500,000 copies/ml.
- Corresponding authors: 17
- 1. Dr Varun Mehra; Varun.Mehra@nhs.net: +442032995378 18
- Alternate Corresponding Author: 19
- 20 2. Dr Majid Kazmi; majidkazmi@nhs.net; +442071882757
- 22 **Key Words:**
- 23 Multiple Sclerosis; Autologous Hematopoietic Stem Cell Transplantation, Epstein-
- 24 Barr Virus Infection; Monoclonal Gammopathy; Post-transplant Lymphoproliferative
- Disorder 25

Author Affiliations:

- **1. Dr Varun Mehra***; Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
- 2. Dr Elijah Rhone*: Department of Neurology, King's College Hospital NHS Foundation Trust,
 Denmark Hill, London, United Kingdom
 - 3. Stefani Widya: GKT School of Medical Education, Kings College London University, London
 - **4. Dr Mark Zuckerman:** Department of Virology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **5. Dr Victoria Potter:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **6. Dr Kavita Raj:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom **AND** Department of Haematology, Guy's and St. Thomas' NHS Foundation Trust, London, United Kingdom
 - **7. Dr Austin Kulasekararaj:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **8. Dr Donal McLornan**: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom **AND** Department of Haematology, Guy's and St. Thomas' NHS Foundation Trust, London, United Kingdom
 - **9. Dr Hugues de Lavallade**: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **10. Nana Benson-Quarm:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **11. Christina Lim:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **12. Sarah Ware:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom.
 - **13. Dr Malur Sudhanva:** Department of Virology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - **14. Dr Omar Malik:** Department of Neurology, Imperial College Healthcare, London, United Kingdom
 - **15. Dr Richard Nicholas:** Department of Neurology, Imperial College Healthcare, London, United Kingdom
 - **16. Professor Paolo A. Muraro:** Department of Neurology, Imperial College Healthcare, London, United Kingdom **AND** Department of Neuroimmunology, Imperial College London, London, United Kingdom
 - **17. Professor Judith Marsh:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - **18. Professor Ghulam J. Mufti:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - **19. Dr Eli Silber:** Department of Neurology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - **20. Professor Antonio Pagliuca:** Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom
 - **21. Dr Majid Kazmi**: Department of Haematology, King's College Hospital NHS Foundation Trust, Denmark Hill, London, United Kingdom **AND** Department of Haematology, Guy's and St. Thomas' NHS Foundation Trust, London, United Kingdom

78	Confl	ict of interest: The authors declare no competing financial interests as below:
79	1.	Varun Mehra- no competing financial interests
80	2.	Elijah Rhone- no competing financial interests
81	3.	Stefani Widya- no competing financial interests
82	4.	Mark Zuckerman- no competing financial interests
83	5.	Victoria Potter- no competing financial interests
84	6.	Kavita Raj- no competing financial interests
85	7.	Austin Kulasekararaj- no competing financial interests
86	8.	Donal McLornan- no competing financial interests
87	9.	Hugues de Lavallade- no competing financial interests
88	10.	Nana Benson-Quarm- no competing financial interests
89	11.	Christina Lim- no competing financial interests
90	12.	Sarah Ware- no competing financial interests
91	13.	Malur Sudhanva- no competing financial interests
92	14.	Omar Malik- no competing financial interests
93	15.	Richard Nicholas- no competing financial interests
94	16.	Paolo A Muraro- no competing financial interests
95	17.	Judith Marsh- no competing financial interests
96	18.	Ghulam J Mufti- no competing financial interests
97	19.	Eli Silber- no competing financial interests
98	20.	Antonio Pagliuca- no competing financial interests
99	21.	Majid A. Kazmi- no competing financial interests
100		
101		
102		
103		
104		
105		
106		
107		
108		

Abstract

Introduction

Autologous haematopoietic stem cell transplantation (AHSCT) with anti-thymocyte globulin (ATG) conditioning as treatment of active multiple sclerosis (MS) is rapidly increasing across Europe (EBMT registry data 2017). Clinically significant Epstein Barr virus reactivation (EBV-R) following AHSCT with ATG for severe autoimmune conditions is an under-recognised complication relative to T-cell deplete transplants performed for haematological diseases. This retrospective study reports EBV-R associated significant clinical sequelae in MS patients undergoing AHSCT with rabbit ATG.

Methods

Retrospective data was analysed for 36 consecutive MS-AHSCT patients at Kings College Hospital, London. All patients routinely underwent weekly EBV DNA PCR monitoring and serum electrophoresis for monoclonal gammopathy (MG or M-protein). EBV-R with rising EBV viral load, M-protein and associated clinical sequelae were captured from clinical records.

Results

All patients had evidence of rising EBV DNA-emia, including 7 who were lost to long term follow-up, with a number of them developing high EBV viral load & associated lymphoproliferative disorder (LPD). Nearly 72% (n-18/29) developed de-novo MG, some with significant neurological consequences with high M-protein and EBV-R. Six patients required anti-CD20 therapy (rituximab) with complete resolution of EBV related symptoms. Receiver operating characteristics (ROC) estimated a peak EBV viraemia of >500,000 DNA copies/ml

133	correlated with high sensitivity (85.5%) & specificity (82.5%) (AUC-0.87; p-0.004) in
134	predicting EBV-R related significant clinical events.
135	Conclusion
136	Symptomatic EBV reactivation increases risk of neurological sequelae and LPD in MS-
137	AHSCT. We recommend regular monitoring for EBV and serum electrophoresis for MG in
138	MS patients in the first 3 months post AHSCT
139	
140	
141	
142	
143	
144	
145	
146	
147	
148	
149	
150	

INTRODUCTION:

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

Multiple sclerosis (MS) is a chronic autoimmune, inflammatory, demyelinating disease of the central nervous system[1]-[2], with a relapsing-remitting (RRMS) presentation in the majority of patients at diagnosis. Recovery from relapses may be complete or partial[3]-[4]. After a variable period of time, people with RRMS may develop a more progressive disability accumulation with or without superimposed relapses; termed secondary progressive multiple sclerosis (SPMS). A minority experience progressive disability from the onset of disease, termed primary progressive multiple sclerosis (PPMS)[4]. A number of immunomodulatory disease modifying therapies (DMTs) are currently licensed for treatment of RRMS with an aim of reducing number of relapses and accrual of disability, although with variable efficacy[5]. Since 1996, Autologous Hematopoietic Stem Cell Transplantation (AHSCT) has been a novel approach for MS management, using immunoablation followed by immunomodulation mechanisms, with evidence of significant suppression of inflammatory activity and qualitative changes in the reconstituted immune system (immune reset theory)[6-8]. AHSCT appears most effective for MS patients with evidence of inflammatory activity on MRI, younger age, a shorter disease duration, low to moderate disability levels (Expanded Disability Status Scale [EDSS] <6 or up to 6.5 if recent progression) and failure of at least 1 highly active DMT (natalizumab or alemtuzumab) with no significant comorbidities[9-11]. Recently reported preliminary results of randomised MIST study[12] found AHSCT to be superior to standard disease modifying therapy (DMT) for RRMS with respect to both

treatment failure and disability progression.

172

173

174

175

176

177

178

179

180

181

171

subsequent opportunistic infections following However, risk rise in immunosuppressive therapies remain a potential concern[13]. MS patients undergoing AHSCT have often been exposed to a number of immunomodulating DMTs; the addition of immunosuppressive rabbit anti-thymocyte globulin (rATG) to their conditioning regimen may confer a higher risk of viral reactivation in these patients. The number of AHSCTs performed for MS is rising significantly in Europe[14] and as more centres perform AHSCT for this indication, it is increasingly important to recognise the unique problems faced by these patients post AHSCT. This retrospective study reports for the first time, EBV-R associated neurological sequelae and lymphoproliferative disorder (LPD) in MS patients undergoing rATG conditioned AHSCT in our centre.

183

184

185

186

187

188

189

190

182

METHODS

Patients and procedures

Data was collected retrospectively on 36 consecutive MS patients undergoing AHSCT between February 2012 and February 2017 at Kings College Hospital, London. Peripheral blood stem cells were collected following standard mobilisation strategy consisting of cyclophosphamide 4g/m² over 2 days and granulocyte colony-stimulating factor for 7 days. Thirty-five (97%) patients were conditioned using standard protocol of cyclophosphamide

(50mg/kg for 4 days) and rATG (2.5mg/kg/day for 3 days) for in-vivo lymphodepletion followed by stem cell infusion. One patient was conditioned with carmustine/etoposide/cytarabine/melphalan regimen along with an equivalent dose of rATG (BEAM-ATG) prior to stem cell infusion. The median CD34 stem cell dose returned was 7.17 x10^6/kg (range 4.0-17.1x10^6/kg).

196

197

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

191

192

193

194

195

Prior exposure to EBV was assessed by serological evidence (EBV viral capsid antigen; VCA IgG). EBV DNA load monitoring was performed on whole blood samples by standardised quantitative real-time polymerase chain reaction (RT-PCR) using Rotor-GeneTM (Qiagen) assay of EBV BZLF1 DNA. This assay was adapted from our published assay using LightCycler (Roche)[15] and since been validated against the recently published WHO standard, with our lab's EBV DNA quantification of 10 copies/ml considered equivalent to 10 IU/ml DNA reported with the WHO reference method[16]. EBV-R was defined as rising EBV DNA load of >10 copies/millilitre (ml) detected on two consecutive tests based on our assay sensitivity. Symptomatic EBV-R was captured by peak blood EBV DNA load, presence of B symptoms (defined by presence of either unexplained weight loss, recurrent fever, night sweats); which was in-turn defined by clinical, radiological and/or histological evidence based on recent ECIL-6 guidelines[17]. In addition, significant 'clinical events' were also defined as new & persistent organ dysfunction (e.g. neurological events) temporally associated with rising EBV viraemia in MS patients. Serum protein electrophoresis was routinely tested around 3 months post HSCT as part of our institutional practice, with immunoglobulin subclasses identified by immunofixation electrophoresis. Patient outcomes were assessed at last follow up as of April 2017.

Statistics

The database of transplants and outcomes was built in Microsoft Excel 2016 and statistical analyses were performed using IBM SPSS Statistics version 24.0. Patient characteristics are presented as medians (with inter-quartile ranges; IQR) for data with non-normal distribution. Comparisons of baseline characteristics used Mann-Whitney U test, Fisher's exact test, or Chi-squared test for trend as appropriate. Receiver Operating characteristics (ROC) curve was obtained correlating LPD and clonal gammopathy associated clinical events with rising EBV viraemia (copies/ml).

RESULTS

Baseline characteristics are presented in **Table 1**. Most MS patients (88.9%) had RRMS phenotype with median of 2 (range 0-3) DMTs prior to AHSCT. Twenty-two MS patients had prior exposure to natalizumab and 7 were treated with Alemtuzumab (6 patients received both). All 36 (100%) MS patients were serologically positive for EBV VCA IgG pre-treatment, indicating prior EBV exposure and had detectable EBV DNA post-AHSCT. Seven MS patients were lost to long-term follow up for EBV monitoring. The median time to first EBV DNA detection post-transplant was 30 days (IQR 23-46 days). EBV DNA levels peaked at a median of 32 days post-transplant (IQR 31-53 days). All MS patients had normal baseline lymphocyte counts (median) pre-HSCT with a median time of 46 days (range 14-404 days) to lymphocyte recovery (defined by total lymphocyte count >1.0x106/ml) following AHSCT (**See Figure 1**). A high proportion (86%; n-25/29) of the MS patients in active follow-up

recovered lymphocyte counts around D56 with a median lymphocyte count of 1.56 (10⁶ cells/ml); Four patients remained lymphopenic at last follow up.

238

239

240

241

242

243

244

245

246

247

248

249

250

251

237

236

All patients were stratified into following 3 groups according to peak rise/burden of EBV DNAaemia (copies/ml): <100,000 (<100k) copies/ml, 100,001-500,00 (100k-500k) copies/ml and >500,000 (>500k) copies/ml to identify any specific thresholds for clinically significant events related to rising EBV-R (Table 1). The majority of patients (76%) with rising EBV viral load >100k copies/ml were routinely screened by computed tomographic (CT) scans to assess for evidence of LPD, in line with our institutional policy. One third (34.5%) of patients developed peak EBV viraemia of >500k copies/ml. Eight patients (27.6%) developed symptomatic EBV-R; defined as persistent fever, lymphadenopathy and/or B symptoms. Of these 8 patients, only 1 (12.5%) had a peak EBV viraemia <100kDNA copies/ml with the remaining 7 (87.5%) patients having a peak EBV viraemia of >500k copies/ml. Three patients with rising EBV viraemia >500k copies/ml had findings consistent with probable LPD on CT imaging; however, none had definitive histological diagnosis. Three MS patients had worsening neurological symptoms concurrent with rising EBV viraemia >500k copies/ml and clonal gammopathy, as described below.

253

254

255

252

Interestingly, we also observed frequent de novo monoclonal gammopathy (MG or M-protein) in 18 MS patients (62.4%) following rising EBV viraemia; the majority (n-16) of whom

developed IgG subtype and the remaining 2 developed IgA and IgM M-protein. Concerningly two of these patients developed clinically significant M-Protein burden; one patient with IgG Kappa M-protein of 45.6g/L developed hyper-viscosity and neurological symptoms mimicking MS relapse, requiring plasma exchange. Another patient developed significant lumbosacral radiculopathy with rising EBV-R and high IgM lambda M-protein (IgM 48.5g/L) (see supplementary case vignettes). Figure 2 highlights the association of neurological symptom onset following rising EBV viraemia (log copies), falling lymphocyte counts (x 10⁶/ml) with significant rise in M-protein (gm/lt) levels post AHSCT. A third patient developed painful lower limb paraesthesia following rising EBV viraemia >500k copies/ml, although did not have any M-protein detected. Their symptoms persisted at last follow up despite no evidence of MS related new disease activity.

Six patients were treated with anti-CD20 antibody, rituximab (375mg/m2 weekly up to 4 weeks), due to clinical severity of EBV reactivations and leading to reduction in EBV viral load and concurrent improvement/resolution of EBV related symptoms. ROC curve analysis (**Figure 3**) confirmed EBV viraemia of >500k copies/ml correlated with high sensitivity (85.5%) and specificity (82.5%) (AUC-0.87; 95%CI-0.73-1.0; p-0.004) in predicting significant EBV related clinical events (evidence of LPD and/or neurological symptoms) that may require treatment with rituximab. The sensitivity dropped significantly on lower estimates for events below 500k copies/ml.

The median time to resolution of EBV viraemia post-rituximab was 21 days (IQR 19-124 days) in 5 patients with >500k copies/ml (one patient was treated for late onset persistent symptomatic clonal gammopathy, despite fall in EBV levels). No significant adverse events were noted in the treated group. Nine patients had a persistent low level EBV viraemia detectable at last follow up. All patients who underwent AHSCT were alive as of April 2017.

DISCUSSION:

MS as an autoimmune disorder (AD) is theorised to have generally similar underlying pathophysiological immune dysregulation mechanisms[18–22] relative to other chronic autoimmune conditions. Epstein-Barr virus (EBV) is increasingly implicated in the pathogenesis of MS by virtue of epidemiological and serological evidence, impaired CD8+ T cell immune responses to EBV and possible underlying genetic susceptibility for autoimmunity (with EBV encoded protein interactions), as recently described by Harley et al and others[1,23–25].

Generally, EBV-R related LPD has been reported in both allogeneic (allo-HSCT) and solid organ transplants treated with immunosuppressive therapy, often with a significant impact on organ function and overall survival[26–30]. It is observed that reduced intensity allo-HSCT for malignant haematological conditions using alemtuzumab have a relatively lower overall

risk of LPD compared to ATG based treatments, possibly mediated by more effective pan-B & T cell lymphodepletion. In contrast, ATG primarily affects the T-cell repertoire with delayed EBV specific CD8+ T cell recovery[31]. Clinically significant endogenous viral infections including EBV following ATG conditioned AHSCT for severe ADs such as Crohn's disease and systemic sclerosis is increasingly recognised, but the development of lymphoproliferative disorders (LPD) remains rare in these ADs[13,32,33]. Nash et al[32] concerningly reported 2 deaths (1 MS and 1 systemic sclerosis patient) from EBV related LPD in 56 ATG AHSCT autoimmune conditioned for diseases. Additionally, EBV associated haemophagocytosis in ATG-AHSCT for ADs have also been reported[34], with one resulting in death of the patient[35].

306

307

308

309

310

311

312

313

314

315

296

297

298

299

300

301

302

303

304

305

Our report of suspected EBV related LPDs (~10%) in MS-AHSCT group is relatively higher than published reports with allo-HSCTs (4.5-7%)[36,37] and our own centre's unpublished T-cell depleted allo-HSCT experience (6.5%); possibly a reflection of underlying immunopathological state of MS itself[38]. This is further corroborated by the fact that similar LPD risk has not been observed in other ADs, e.g. Crohn's disease, treated with ATG -AHSCT in this centre. Another example from our centre's experience of severe aplastic anaemia (n-40) treated with ATG/ciclosporin, only 52% (n-21/40) developed EBV-R (unpublished data) and none had LPD or required any treatment, suggesting that the problem may not be ATG specific.

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

Our study's observation of significant persistent neurological events (with no evidence of new MS disease activity) associated with clonal gammopathy suggest a potentially new clinical syndrome, described for the first time in ATG conditioned AHSCTs in MS and possibly induced by clonal B cell dysregulation following EBV-R. It could be hypothesised that any remaining EBV infected latent B cells, surviving despite high doses of cyclophosphamide (given with mobilisation and conditioning in MS ASHCT)[8] and compounded by depletion of CD8+ T cells by ATG, may serve as potential source for EBV escape while interacting abnormally within the host immune micro-environment[39] and leading to rise in M-protein, LPD and neuro-inflammatory insults in some of the MS patients post AHSCT. Reports of lower incidence of EBV-R and LPD in another commonly used protocol for MS-AHSCT using BEAM-ATG (Ricardo Saccardi, Carregi University Hospital, Florence; personal communication) may reflect the greater myeloablative effect of BEAM chemotherapy which could further deplete the residual B cell pool and thus lower potential for EBV proliferation. It is plausible that dose of rATG is critical, given we have not seen similar reports from other centres where less rATG doses were given for MS-AHSCT (range between 5.0-6.5 mg/Kg; personal communication) but there seems to be some variability in prospective serial EBV monitoring in these patients.

The clinical threshold for EBV viral load as a significant risk factor for post-transplant LPD is widely debated. This study reports peak EBV viral load >500k copies/ml post AHSCT is significantly associated with probable LPD and neurological events in MS patients with high sensitivity (85.5%) and specificity (82.5%) (p-0.004) (Fig 3, ROC curve). Our ROC curve estimates are potentially limited by the relatively small number of events analysed but this has consistently been useful in our MS-AHSCT experience for predicting clinical events with high EBV load. Our EBV PCR assay has been validated against the recently defined standard WHO reference method (i.e. 10 copies/ml=10 IU/ml EBV DNA) [16] and thus this EBV threshold for pre-emptive treatment with Rituximab, can potentially be applied in relevant clinical context in other centres using similar validated essays. Rituximab treatment delivered good overall response in our symptomatic patients, with resolution of EBV related clinical symptoms and no subsequent viral or bacterial infections at last follow up. The role of prophylactic or pre-transplant rituximab in MS-AHSCT is also a potential area of interest in reducing risk of EBV-LPD in stem cell transplants as observed by Burns et al[36] and future randomised studies are required to investigate its potential benefit.

350

351

352

353

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

Our study limitations include its retrospective nature and that no suspected LPD patients had histological confirmation, mainly related to patient refusal or technical difficulties. Seven MS patients were lost to follow up for EBV monitoring following discharge, which limits the

findings of this study. Additionally, our numbers were too small to identify any association of EBV related clinical events with previous DMT exposure in MS patients. In conclusion, symptomatic EBV reactivation increases risk of neurological sequelae and LPD in MS-AHSCT. Regular monitoring for rising EBV viraemia, as recommended by Snowden et al [10] and Muraro et al [11], and serum electrophoresis for M-protein should be considered in the first 3 months post-AHSCT for MS. We recommend persistent high EBV viraemia > 500k DNA copies/ml as potential trigger for consideration of pre-emptive anti-CD20 therapy and potentially reduce associated morbidity. **Acknowledgements:** To our patients and their families and carers in supporting this study.

REFERENCES

\sim	\neg	-
1	1	n
\mathcal{L}	,	v

- 1. Pender MP, Burrows SR. Epstein–Barr virus and multiple sclerosis: potential opportunities for
- immunotherapy. Clinical & Translational Immunology **2014**; 3:e27. Available at:
- 379 http://doi.wiley.com/10.1038/cti.2014.25.
- Reich DS, Lucchinetti CF, Calabresi PA. Multiple Sclerosis. New England Journal of Medicine 2018;
- 381 378:169–180. Available at: http://www.nejm.org/doi/10.1056/NEJMra1401483.
- 382 3. Compston A, Coles A. Multiple sclerosis. The Lancet 2008; 372:1502–1517. Available at:
- http://linkinghub.elsevier.com/retrieve/pii/S0140673608616207.
- Lublin FD, Reingold SC, Cohen JA, et al. Defining the clinical course of multiple sclerosis: The 2013
- 385 revisions. Neurology **2014**; 83:278–286. Available at:
- 386 http://www.neurology.org/cgi/doi/10.1212/WNL.000000000000560.
- 5. Comi G, Radaelli M, Soelberg Sørensen P. Evolving concepts in the treatment of relapsing multiple
- 388 sclerosis. The Lancet **2017**; 389:1347–1356. Available at:
- 389 https://linkinghub.elsevier.com/retrieve/pii/S0140673616323881.
- 390 6. Muraro PA, Douek DC, Packer A, et al. Thymic output generates a new and diverse TCR repertoire
- after autologous stem cell transplantation in multiple sclerosis patients. The Journal of Experimental
- 392 Medicine **2005**; 201:805–816. Available at: http://www.jem.org/lookup/doi/10.1084/jem.20041679.
- 393 7. Muraro PA, Robins H, Malhotra S, et al. T cell repertoire following autologous stem cell transplantation
- for multiple sclerosis. J Clin Invest **2014**; 124:1168–1172. Available at:
- 395 http://www.ncbi.nlm.nih.gov/pubmed/24531550.
- 396 8. Cull G, Hall D, Fabis-Pedrini M, et al. Lymphocyte reconstitution following autologous stem cell
- transplantation for progressive MS. Multiple Sclerosis Journal Experimental, Translational and
- 398 Clinical **2017**; 3:205521731770016. Available at: https://doi.org/10.1177/2055217317700167.
- 399 9. Sormani MP, Muraro PA, Schiavetti I, et al. Autologous hematopoietic stem cell transplantation in
- 400 multiple sclerosis. Neurology **2017**; 88:2115–2122. Available at:
- 401 http://www.neurology.org/lookup/doi/10.1212/WNL.000000000003987.
- 402 10. Snowden JA, Saccardi R, Allez M, et al. Haematopoietic SCT in severe autoimmune diseases:
- 403 updated guidelines of the European Group for Blood and Marrow Transplantation. Bone Marrow
- Transplantation **2012**; 47:770–790. Available at: http://www.nature.com/articles/bmt2011185.
- 405 11. Muraro PA, Pasquini M, Atkins HL, et al. Long-term Outcomes After Autologous Hematopoietic Stem
- 406 Cell Transplantation for Multiple Sclerosis. JAMA Neurology **2017**; 74:459. Available at:
- 407 http://archneur.jamanetwork.com/article.aspx?doi=10.1001/jamaneurol.2016.5867.
- 408 12. Burt RK, Balabanov R, Snowden JA, Sharrack B, Oliveira MC, Burman J. Non-myeloablative
- 409 hematopoietic stem cell transplantation (HSCT) is superior to disease modifying drug (DMD) treatment
- 410 in highly active Relapsing Remitting Multiple Sclerosis (RRMS): interim results of the Multiple Sclerosis

- International Stem cell Transp. Neurology **2018**; 90. Available at:
- http://n.neurology.org/content/90/15_Supplement/S36.004.abstract.
- 413 13. Daikeler T, Tichelli A, Passweg J. Complications of autologous hematopoietic stem cell transplantation
- for patients with autoimmune diseases. Pediatric Research **2012**; 71:439–444. Available at:
- 415 http://www.nature.com/doifinder/10.1038/pr.2011.57.
- 416 14. Snowden JA, Badoglio M, Labopin M, et al. Evolution, trends, outcomes, and economics of
- hematopoietic stem cell transplantation in severe autoimmune diseases. Blood Advances **2017**;
- 418 1:2742–2755. Available at:
- http://www.bloodadvances.org/lookup/doi/10.1182/bloodadvances.2017010041.
- 420 15. Patel S, Zuckerman M, Smith M. Real-time quantitative PCR of Epstein–Barr virus BZLF1 DNA using
- the LightCycler. Journal of Virological Methods **2003**; 109:227–233. Available at:
- 422 http://linkinghub.elsevier.com/retrieve/pii/S0166093403000764.
- 423 16. Semenova T, Lupo J, Alain S, et al. Multicenter Evaluation of Whole-Blood Epstein-Barr Viral Load
- Standardization Using the WHO International Standard. Journal of Clinical Microbiology **2016**; 54:1746
- 425 LP-1750. Available at: http://jcm.asm.org/content/54/7/1746.abstract.
- 426 17. Styczynski J, van der Velden W, Fox CP, et al. Management of Epstein-Barr Virus infections and post-
- 427 transplant lymphoproliferative disorders in patients after allogeneic hematopoietic stem cell
- 428 transplantation: Sixth European Conference on Infections in Leukemia (ECIL-6) guidelines.
- 429 Haematologica **2016**; 101:803–811. Available at:
- http://www.haematologica.org/cgi/doi/10.3324/haematol.2016.144428.
- 431 18. Dejaco C, Duftner C, Grubeck-Loebenstein B, Schirmer M. Imbalance of regulatory T cells in human
- 432 autoimmune diseases. Immunology **2006**; 117:289–300. Available at:
- 433 http://doi.wiley.com/10.1111/j.1365-2567.2005.02317.x.
- 434 19. Arellano G, Acuña E, Reyes LI, et al. Th1 and Th17 Cells and Associated Cytokines Discriminate
- among Clinically Isolated Syndrome and Multiple Sclerosis Phenotypes. Frontiers in immunology **2017**;
- 436 8:753. Available at: http://journal.frontiersin.org/article/10.3389/fimmu.2017.00753/full.
- 437 20. Jha S, Srivastava SY, Brickey WJ, et al. The Inflammasome Sensor, NLRP3, Regulates CNS
- 438 Inflammation and Demyelination via Caspase-1 and Interleukin-18. Journal of Neuroscience 2010;
- 439 30:15811–15820. Available at: http://www.jneurosci.org/cgi/doi/10.1523/JNEUROSCI.4088-10.2010.
- 440 21. Beynon V, Quintana FJ, Weiner HL. Activated Human CD4+CD45RO+ Memory T-Cells Indirectly
- Inhibit NLRP3 Inflammasome Activation through Downregulation of P2X7R Signalling. PLoS ONE
- 442 **2012**; 7:e39576. Available at: https://dx.plos.org/10.1371/journal.pone.0039576.
- 443 22. Fernández-Menéndez S, Fernández-Morán M, Fernández-Vega I, Pérez-Álvarez A, Villafani-Echazú J.
- 444 Epstein–Barr virus and multiple sclerosis. From evidence to therapeutic strategies. Journal of
- Neurological Sciences **2016**; 361:213–219. Available at:
- 446 https://www.clinicalkey.com.ezsecureaccess.balamand.edu.lb/service/content/pdf/watermarked/1-s2.0-
- 447 S0022510X16300132.pdf?locale=en_US.

- 448 23. Pender MP. The Essential Role of Epstein-Barr Virus in the Pathogenesis of Multiple Sclerosis. The
- 449 Neuroscientist **2011**; 17:351–367. Available at:
- 450 http://journals.sagepub.com/doi/10.1177/1073858410381531.
- 451 24. Ascherio A, Munger KL. Environmental risk factors for multiple sclerosis. Part I: The role of infection.
- 452 Annals of Neurology **2007**; 61:288–299. Available at: http://doi.wiley.com/10.1002/ana.21117.
- 453 25. Harley JB, Chen X, Pujato M, et al. Transcription factors operate across disease loci, with EBNA2
- implicated in autoimmunity. Nature Genetics **2018**; 50:699–707. Available at:
- 455 http://www.nature.com/articles/s41588-018-0102-3.
- Loren AW, Porter DL, Stadtmauer EA, Tsai DE. Post-transplant lymphoproliferative disorder: a review.
- 457 Bone Marrow Transplantation **2003**; 31:145–155. Available at:
- 458 http://www.nature.com/articles/1703806.
- 459 27. Dotti G, Fiocchi R, Motta T, et al. Lymphomas occurring late after solid-organ transplantation: influence
- of treatment on the clinical outcome. Transplantation **2002**; 74:1095–102. Available at:
- http://www.ncbi.nlm.nih.gov/pubmed/12438953.
- 462 28. Nagle SJ, Reshef R, Tsai DE. Posttransplant Lymphoproliferative Disorder in Solid Organ and
- Hematopoietic Stem Cell Transplantation. Clinics in Chest Medicine **2017**; 38:771–783. Available at:
- https://linkinghub.elsevier.com/retrieve/pii/S0272523117300874.
- 465 29. Meijer E, Dekker AW, Weersink AJL, Rozenberg-Arska M, Verdonck LF. Prevention and treatment of
- 466 epstein-barr virus-associated lymphoproliferative disorders in recipients of bone marrow and solid
- organ transplants. British Journal of Haematology **2002**; 119:596–607. Available at:
- 468 http://dx.doi.org/10.1046/j.1365-2141.2002.03887.x.
- 469 30. Dierickx D, Habermann TM. Post-Transplantation Lymphoproliferative Disorders in Adults. New
- 470 England Journal of Medicine **2018**; 378:549–562. Available at:
- 471 http://dx.doi.org/10.1056/NEJMra1702693.
- 472 31. Meij P. Impaired recovery of Epstein-Barr virus (EBV)--specific CD8+ T lymphocytes after partially T-
- depleted allogeneic stem cell transplantation may identify patients at very high risk for progressive
- EBV reactivation and lymphoproliferative disease. Blood **2003**; 101:4290–4297. Available at:
- 475 http://www.bloodjournal.org/cgi/doi/10.1182/blood-2002-10-3001.
- 476 32. Nash R a, Dansey R, Storek J, et al. Epstein-Barr virus-associated posttransplantation
- 477 lymphoproliferative disorder after high-dose immunosuppressive therapy and autologous CD34-
- selected hematopoietic stem cell transplantation for severe autoimmune diseases. Biology of blood
- and marrow transplantation: journal of the American Society for Blood and Marrow Transplantation
- 480 **2003**; 9:583–91. Available at: http://www.ncbi.nlm.nih.gov/pubmed/14506660.
- 481 33. Daikeler T, Tzankov A, Hoenger G, et al. Minimal T-cell requirements for triggering
- 482 haemophagocytosis associated with Epstein–Barr virus-driven B-cell proliferation: a clinical case
- 483 study. Annals of the Rheumatic Diseases 2011; 70:1338 LP-1339. Available at:
- http://ard.bmj.com/content/70/7/1338.abstract.

485 34. Daikeler T, Tzankov A, Hoenger G, et al. Minimal T-cell requirements for triggering 486 haemophagocytosis associated with Epstein-Barr virus-driven B-cell proliferation: a clinical case study. 487 Annals of the Rheumatic Diseases 2011; 70:1338–1339. Available at: 488 http://ard.bmj.com/cgi/doi/10.1136/ard.2010.139246. 489 35. Brinkman DMC, de Kleer IM, ten Cate R, et al. Autologous stem cell transplantation in children with 490 severe progressive systemic or polyarticular juvenile idiopathic arthritis: Long-term followup of a 491 prospective clinical trial. Arthritis & Rheumatism 2007; 56:2410-2421. Available at: 492 http://doi.wiley.com/10.1002/art.22656. 493 36. Burns DM, Rana S, Martin E, et al. Greatly reduced risk of EBV reactivation in rituximab-experienced 494 recipients of alemtuzumab-conditioned allogeneic HSCT. Bone Marrow Transplantation 2016; 51:825-495 832. Available at: http://www.nature.com/articles/bmt201619. 496 37. van Esser JWJ. Epstein-Barr virus (EBV) reactivation is a frequent event after allogeneic stem cell 497 transplantation (SCT) and quantitatively predicts EBV-lymphoproliferative disease following T-cell-498 depleted SCT. Blood 2001; 98:972-978. Available at: 499 http://www.bloodjournal.org/cgi/doi/10.1182/blood.V98.4.972. 500 38. Tørring C, Andreasen C, Gehr N, Bjerg L, Petersen T, Höllsberg P. Higher incidence of Epstein-Barr 501 virus-induced lymphocyte transformation in multiple sclerosis. Acta Neurologica Scandinavica 2014; 502 130:90-96. Available at: http://doi.wiley.com/10.1111/ane.12249. 503 39. Martinez OM, Krams SM. The Immune Response to Epstein Barr Virus and Implications for 504 Posttransplant Lymphoproliferative Disorder. Transplantation 2017; 101:2009–2016. Available at: 505 http://insights.ovid.com/crossref?an=00007890-201709000-00018. 506 507 508 509 510 511 512 513 514 515 516 517 518 519 520 521 522 523 524

526

Baseline characteristics (n-36)		Patient Groups according to peak EBV DNA in copies/ml (n-29)	0 - 100,000	100,001 - 500,000	>500,000
Median age at time of AHSCT in years (range)	43.5 (36– 47)	No of patients (%)	16 (55.2)	3 (10.3)	10 (34.5)
Gender Male Female	19 (52.8%) 17 (47.2%)	M-Protein (n)	11	0	7
Disease Type (n; %) Relapsing Remitting MS Secondary Progressive MS Primary Progressive MS	22 (61.1%) 10 (27.8%) 4 (11.1%)	Median EBV DNA log value at peak (IQR)	4.8 (3.5-4.8)	5.5 (N/A)	6.25 (6.1-6.9)
Median number of previous DMT (range)	2 (0 – 6)	Median number of prior DMTs	2	3	2
Previous use of high efficacy DMT (n) Natalizumab Alemtuzumab Both	22 8 6	Symptomatic EBV (n)	1	0	7
Median EDSS (range)	6.0 (2.5 – 8.0)	LPD diagnosis (CT/Biopsy) (n)	0	0	3 by CT alone
Median follow up post AHSCT in days (range)	436 (188 – 785)	Neuro/autoimmune complications (n)	0	0	3
No. of patients with prior EBV exposure (n; %)	36 (100%)	Treated with Rituximab (n)	0	0	6
No. of patients with detectable EBV post AHSCT (n; %)	29 (80.5%)	Confirmed EBV resolution at last follow up (n)	7	2	7
No. of patients lost to long term follow up (n; %)	7 (19.5%)	Detectable EBV DNA at last follow up (n)	9	1	3
Median Time to EBV detection post AHSCT in days (IQR)	30 (23-46)	Median time for EBV resolution (IQR in days)	67 days (44-155)	47 days (N/A)	63 days (45 - 170)
Median Time to peak EBV DNA levels in days (IQR)	32 (31-53)	Median Time to peak EBV DNA levels in days (IQR)	40 days (25-85)	30 days (N/A)	39 days (32-43)

Abbreviations:

529

- 531 AHSCT- Autologous Haematopoietic Stem cell transplant; CT- computed tomography; DMT-
- 532 Disease modifying therapy; EBV- Epstein Barr virus; EDSS- Kurtzke Expanded Disability
- 533 Status Scale; IQR- Interquartile range; LPD- Lymphoproliferative disease; M-Protein:
- Monoclonal paraprotein or gammopathy; MS- Multiple Sclerosis

Figure Legends

537 Figure 1: Trend of Lymphocyte count recovery following ATG in MS patients.

Legend: This figure shows trends of lymphocyte count from baseline to recovery post AHSCT for MS patients. Majority of patients had normal baseline lymphocyte counts pre-HSCT and became lymphopenic post ATG with counts recovering towards d+28 and >85% of patients recovered counts by d+56, with some overshooting from their baseline, possibly reflective of EBV related lymphoproliferation in some of these patients.

AHSCT: Autologous Haematopoietic Stem Cell Transplantation; ATG: Anti-Thymocyte Globulin; d+: Days post AHSCT; MS: Multiple Sclerosis.

Figure 2: Paraprotein, EBV & Lymphocyte trends in two MS patients with significant neurological sequelae post AHSCT.

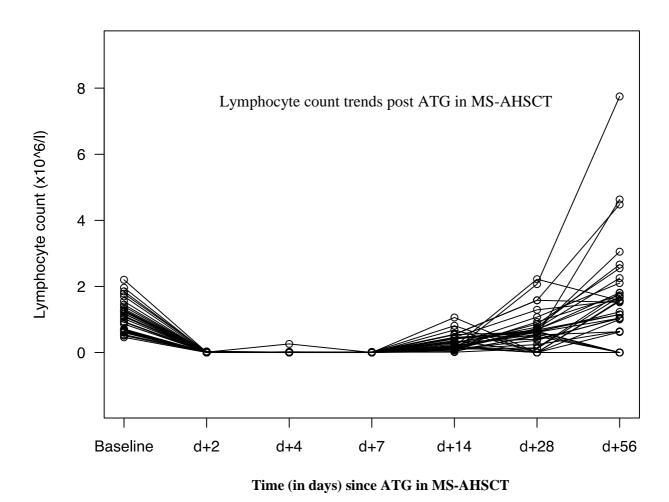
Legend: This figure demonstrates trends of EBV copies (log), paraprotein levels (g/lt) and Lymphocyte levels (counts x10^6/ml) in two MS patients with significant neurological symptoms following EBV reactivation. Both patients had significant EBV viraemia (log>5.2 or >500,000 copy number) and developed significant paraproteinaemia, which was only noted after persistent unexplained neurological symptoms. The trend reversed following anti-CD20 (rituximab) therapy, with limited recovery in neurological symptoms.

D: Days post AHSCT; EBV-R: Epstein Barr Virus reactivation; MS: Multiple sclerosis; AHSCT: Autologous Haematopoietic Stem Cell Transplants.

Figure 3: ROC curve estimates for peak EBV viraemia levels and significant clinical events in MS post AHSCT.

Legend: ROC curve demonstrating significant correlation between high EBV levels and clinical events (LPD & neurological events) in MS-AHSCT patients, with highest sensitivity and specificity noted with peak EBV viraemia of >500,000 copies/ml (p-0.0004).

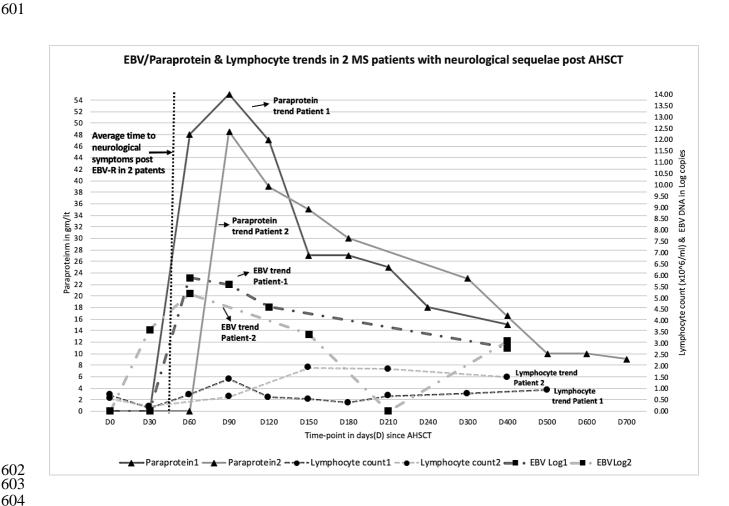
EBV: Epstein Barr Virus; LPD- Lymphoproliferative disorder; MS-AHSCT: Multiple Sclerosis patients treated with autologous haematopoietic stem cell transplants; ROC: receiver operating characteristics



Abbreviations:

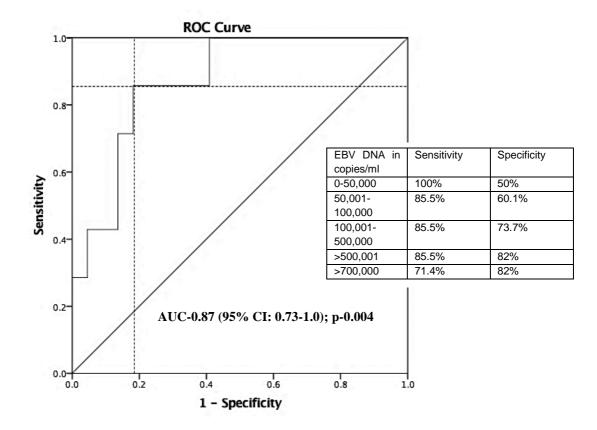
AHSCT: Autologous Haematopoietic Stem Cell Transplantation; ATG: Anti-Thymocyte Globulin; d+: Days post AHSCT; MS: Multiple Sclerosis.

Figure 2. Paraprotein, EBV & Lymphocyte trends in two MS patients with significant neurological sequelae post AHSCT



Abbreviations:

D: Days post AHSCT; EBV-R: Epstein Barr Virus reactivation; MS: Multiple sclerosis; AHSCT: Autologous Haematopoietic Stem Cell Transplants.



Abbreviations:

EBV: Epstein Barr Virus; LPD- Lymphoproliferative disorder; MS-AHSCT: Multiple Sclerosis patients treated with autologous haematopoietic stem cell transplants; ROC: receiver operating characteristics

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.

If this message is not eventually replaced by the proper contents of the document, your PDF viewer may not be able to display this type of document.

You can upgrade to the latest version of Adobe Reader for Windows®, Mac, or Linux® by visiting http://www.adobe.com/go/reader_download.

For more assistance with Adobe Reader visit http://www.adobe.com/go/acrreader.