



## King's Research Portal

*Document Version*  
Peer reviewed version

[Link to publication record in King's Research Portal](#)

*Citation for published version (APA):*

Lorio, S., & Carmichael, D. (in press). MRI profiling of focal cortical dysplasia using multi-compartment diffusion models. *Epilepsia*.

### **Citing this paper**

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

### **General rights**

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Research Portal

### **Take down policy**

If you believe that this document breaches copyright please contact [librarypure@kcl.ac.uk](mailto:librarypure@kcl.ac.uk) providing details, and we will remove access to the work immediately and investigate your claim.

## **MRI profiling of focal cortical dysplasia using multi-compartment diffusion models**

Sara Lorio<sup>1,2</sup>, Sophie Adler<sup>1</sup>, Roxana Gunny<sup>3</sup>, Felice D'Arco<sup>3</sup>, Enrico Kaden<sup>4</sup>, Konrad Wagstyl<sup>5</sup>, Thomas S. Jacques<sup>6</sup>, Chris A. Clark<sup>1</sup>, J. Helen Cross<sup>1</sup>, Torsten Baldeweg<sup>1</sup>, David W Carmichael<sup>1,2,\*</sup>

<sup>1</sup>*Developmental Neurosciences, Great Ormond Street Institute of Child Health, University College London, United Kingdom*

<sup>2</sup>*School of Biomedical Engineering & Imaging Sciences, King's College London, St Thomas' Hospital, London, United Kingdom*

<sup>3</sup>*Great Ormond Street Hospital, London, United Kingdom*

<sup>4</sup>*Centre for Medical Image Computing, University College London, United Kingdom*

<sup>5</sup>*Brain Mapping Unit, Institute of Psychiatry, University of Cambridge, United Kingdom*

<sup>6</sup>*Developmental Biology and Cancer Programme, UCL Great Ormond Street Institute of Child Health, University College London and Department of Histopathology, Great Ormond Street Hospital for Children NHS Foundation Trust, London, United Kingdom*

**Running Head:** Characterisation of FCD with new diffusion MRI

**Key words:** Epileptogenic zone; Cortical dysplasia; multi-compartment diffusion models

Submission Type: **Research article**

Number of text pages: **20**

Word count of Main text: **3999**

Number of References: **46**

Number of Figures: **5 (coloured figures 5)**

Number of Tables: **1**

### **Corresponding Author**

Sara Lorio, PhD

Wellcome EPSRC Centre for Medical Engineering

King's College London

St Thomas' Hospital

London, SE1 7EH

Telephone: +44 (0) 20 7188 7188 ext: 53205

E-mail: [sara.lorio@kcl.ac.uk](mailto:sara.lorio@kcl.ac.uk)

ORCID iD: <https://orcid.org/0000-0002-1790-3586>

\*ORCID iD: <https://orcid.org/0000-0001-9972-0718>

## Abstract

**Objective:** Focal cortical dysplasia (FCD) lesion detection and sub-typing remain challenging on conventional MRI. New diffusion models such as the spherical mean technique (SMT) and neurite orientation dispersion and density imaging (NODDI) provide measurements that potentially produce more specific maps of abnormal tissue microstructure. This study aims to assess the SMT and NODDI maps for computational and radiological lesion characterisation compared to standard fractional anisotropy (FA) and mean diffusivity (MD).

**Methods:** SMT, NODDI, FA and MD maps were calculated for 33 paediatric patients with suspected FCD (18 histologically confirmed). Two neuro-radiologists scored lesion visibility on clinical images and diffusion maps. Signal profile changes within lesions and homologous regions were quantified using a surface-based approach. Diffusion parameter changes at multiple cortical depths were statistically compared between FCDIIa and IIb.

**Results:** Compared to FLAIR or T1w, lesions conspicuity on NODDI intracellular volume fraction (ICVF) maps was better/equal/worse in respectively 5/14/14 patients, while on SMT intra-neurite volume fraction (INVF) in 3/3/27. Compared to FA or MD, lesions conspicuity on the ICVF was better/equal/worse in 27/4/2, while on the INVF in 20/7/6. Quantitative signal profiling demonstrated significant ICVF and INVF reductions in the lesions, while SMT microscopic mean, radial and axial diffusivities were significantly increased. FCDIIb exhibited greater changes than FCDIIa. No changes were detected on FA, MD profiles.

**Significance:** FCD lesion-specific signal changes were found in ICVF and INVF but not in FA and MD maps. ICVF and INVF showed greater contrast than FLAIR in some cases and had consistent signal changes specific to FCD suggesting they could improve current pre-surgical paediatric epilepsy imaging protocols and can provide features useful for automated lesion detection.

**Key words:** Epileptogenic zone; cortical dysplasia; multi-compartment diffusion models

## Introduction

Focal cortical dysplasia (FCD) is a malformation of cortical development, and the most common cause of drug-resistant focal epilepsy in children<sup>1,2</sup>. It is characterised by disrupted tissue organisation with the presence of abnormal cells such as dysmorphic neurons and balloon cells<sup>1</sup>. FCD lesion detection, extent identification and microstructure characterisation on MRI are crucial for planning surgical treatment<sup>3,4</sup>, however the radiological assessment remains challenging<sup>5</sup>.

Diffusion MRI can probe tissue microstructure non-invasively by measuring the diffusion process of water molecules. The most commonly used metrics are based on diffusion tensor imaging (DTI) maps such as fractional anisotropy (FA) and mean diffusivity (MD). Previous studies reported reduced FA<sup>6-9</sup> and increased MD<sup>6,9</sup> values in white matter regions subjacent to MRI-visible FCD lesions. However, alterations in FA and MD distant from the FCD have also been reported<sup>6,10</sup>, and thus the general consensus is that these features are not specific for lesion classification<sup>6,11</sup>.

DTI-based metrics FA and MD cannot differentiate between the contributions to signal changes of fibre density/orientation dispersion and diffusion across intra- and extra-cellular compartments<sup>12-15</sup>. This lack of specificity hampers the neurobiological interpretation and is a confounder in the identification of patho-physiological phenomena in FCD<sup>6,15-17</sup> because similar signal variation can result from pathological changes or normal white matter structure<sup>18</sup>.

New diffusion models such as neurite orientation dispersion and density imaging (NODDI)<sup>12</sup> and spherical mean technique (SMT)<sup>13,14</sup> account for orientation dispersion and fibre crossings within different tissue compartments, with the potentiality to be more specific to microstructural changes in FCD lesions<sup>15</sup>. These multi-compartment models require a greater

range of diffusion data, which until recently would have required clinically impractical scan times particularly for paediatric patients.

A preliminary study used NODDI in 5 patients with suspected FCD<sup>15</sup>, showing that the intracellular volume fraction (ICVF) map enhanced lesion contrast<sup>15</sup>. The SMT multi-compartment microscopic diffusion was used to detect altered intra- and/or extracellular neuro-pathological process in mouse brains affected by tuberous sclerosis complex<sup>13</sup>, the brain lesions of which share histopathological features with FCDIb<sup>19</sup>.

In this study, we aimed to determine whether the diffusion parameters from multi-compartment models demonstrated consistent changes in suspected FCD lesions, to determine whether they were more sensitive than FA and MD, and to test their ability to characterise tissue property differences between histological sub-types. To this end we used recent advances in MRI software and hardware to obtain data suitable for SMT and NODDI techniques in ~7minutes. We investigated a retrospective cohort of more than 30 patients with suspected FCD, utilising NODDI, microscopic diffusion tensor and multi-compartment microscopic diffusion SMT. All patients had MRI positive lesions on clinical 3D FLAIR or T1-weighted images.

First, two neuro-radiologists visually assessed the lesion conspicuity on the new diffusion parameters, FA, MD maps and optimised epilepsy protocol<sup>20</sup> (3D T1-weighted and 3D-FLAIR) to compare lesion contrast on different MR images. Second, changes in diffusion parameters were quantified using profiles across different cortical, subcortical depths and compared with homotopic healthy regions. Finally, we assessed if the new diffusion map profiles were specific to FCD histological subtypes.

## Methods

## Participants

A retrospective cohort of 33 paediatric patients (mean age=10±4years, range=2-21years, female=15) was identified for this research study from all those undergoing assessment for epilepsy surgery at Great Ormond Street Hospital (GOSH), following approval by the national research ethics service. The inclusion criteria were: patients with radiological and electro-clinical diagnosis compatible with FCD, patients who had 3T MRI at GOSH with the full epilepsy imaging protocol that included multi-shell diffusion. Patients younger than 2 years of age with MRI scans showing severe motion artefacts (i.e. indistinguishable adjacent gyri due to motion or severe ringing), or without the full protocol described in the following section were excluded. The 33 patients included in the study represent the radiologically defined group that was used to test visually and quantitatively signal changes on the multi-compartment diffusion maps.

## MR imaging

All patients were scanned on a 3T whole-body MRI system (Magnetom Prisma, Siemens Medical Systems, Germany), using a 20-channel receive head coil and body coil for transmission and 80mT/m magnetic field gradients. Three-dimensional structural T1-weighted (T1w) images were acquired using magnetisation prepared rapid gradient-echo (MPRAGE) (TR/TE=2300/2.74ms, FOV=256×256mm<sup>2</sup>, flip angle=8°, voxel size=1×1×1mm<sup>3</sup>), fluid attenuated inversion recovery (FLAIR) (TR/TE/TI=4000/395/1800ms, FOV=256×256mm<sup>2</sup>, flip angle=120°, voxel size=0.65×1×0.65mm<sup>3</sup>), and a diffusion-weighted protocol. The multi-direction diffusion sequence was included primarily to provide state-of-the-art white matter tractography data for patients going forward to surgery without requiring further imaging. This employed a diffusion-weighted spin-echo single shot 2D EPI acquisition, field of view=220×220mm<sup>2</sup>,

matrix size=110x110, in-plane voxel resolution=2.0mm, GRAPPA factor 2, phase-encoding (PE) partial Fourier=6/8. Multi-band radio frequency pulses allowed simultaneous multi-slice acquisition<sup>21,22</sup>. A multi-band factor of 2 was employed, halving the time required to obtain the 66 slices (2mm thickness with 0.2mm gap). Diffusion gradients were applied over two shells:  $b=1000, 2200\text{s/mm}^2$ , with 60 non-co-linear diffusion directions per shell, in addition to 13 interleaved  $b=0$  ( $b_0$ ) (non-diffusion weighted) images. The gradient strength and eddy current performance enabled monopolar diffusion encoding with  $TE=60\text{ms}$  and  $TR=3050\text{ms}$ , thereby limiting the total acquisition time to 7minutes 20s. For the correction of magnetic susceptibility-related distortions (see Diffusion processing) an additional single  $b_0$  acquisition was performed, with the PE direction flipped by  $180^\circ$  (in the anterior-posterior direction), all other parameters were unchanged.

### Diffusion processing and map estimation

We applied four different techniques to model the diffusion signal across brain tissue:

- model 1: DTI providing FA and MD maps;
- model 2: NODDI providing ICVF and orientation dispersion (OD) maps;
- model 3: SMT microscopic ( $\mu$ ) diffusion tensor estimating  $\mu$  axial diffusivity ( $\mu AD$ ),  $\mu$  radial diffusivity ( $\mu RD$ ),  $\mu FA$  and  $\mu MD$  maps;
- model 4: SMT multi-compartment microscopic diffusion computing intra-neurite volume fraction (INVF), intrinsic diffusivity, extra-neurite  $\mu RD$  and extra-neurite  $\mu MD$  maps.

The pre-processing of the diffusion-weighted data was the same for all models and was performed using FSL5.0 ([www.fmrib.ox.ac.uk/fsl](http://www.fmrib.ox.ac.uk/fsl)). To estimate and correct susceptibility-induced distortions, the diffusion-weighted data was combined with the PE-flipped  $b_0$



image<sup>23</sup> using the “topup” function. Then eddy-current and susceptibility distortions were removed using the “eddy” function before the brain extraction tool (BET)<sup>24</sup> was applied to skull-strip the brain volume.

The diffusion tensor model was fitted to the corrected multi-shell data using “dtifit” with a weighted-least squares fit, and FA and MD maps were calculated.

The NODDI ICVF and OD maps were computed for the brain voxel using the NODDI Matlab Toolbox ([http://www.nitrc.org/projects/noddi\\_toolbox](http://www.nitrc.org/projects/noddi_toolbox))<sup>12</sup> with default settings. Briefly, NODDI models diffusion in each voxel as three independent compartments: intra-neurite, extra-neurite and free water compartment, assuming fixed compartment diffusivities<sup>12</sup>. The intra-neurite compartment characterises the space occupied by neurites and is modelled by a set of ‘sticks’. The extra-neurite compartment models water diffusing in the space around neurites, while the free water compartment represents the free water diffusion (i.e. CSF). The ICVF provides a measure of cell density as a fraction of the non-CSF compartment, whilst the OD estimates the orientation distribution of the intra-neurite compartment.

For the SMT models, the corrected diffusion-weighted data was smoothed with an isotropic Gaussian kernel of 2mm FWHM to remove Gibbs artefacts. The SMT toolbox 0.3 (<https://github.com/ekaden/smt>) was used to estimate the microscopic diffusion tensor and multi-compartment microscopic diffusion maps adjusted for the signal offset induced by Rician noise<sup>25</sup>. The model assumed variable diffusivity across the brain, with maximum value set to  $4 \times 10^{-3} \text{mm}^2/\text{s}$ <sup>13,14</sup>. The microscopic tensor maps are per-axon effective diffusion coefficients unconfounded by the intra-voxel fibre orientation distribution, such as  $\mu\text{AD}$ ,  $\mu\text{RD}$  and  $\mu\text{MD}$ . The multi-compartment microscopic diffusion maps represent estimates of diffusion features specific to the intra- and extra-neurite compartments without the confounding effects of complex fibre orientation distribution, including the INVF and

intrinsic diffusivity.

### *Visual assessment*

To compare the lesion visibility on different MRI contrasts, two neuro-radiologists (RG, FD'A) were presented with coregistered T1w, FLAIR, FA, MD, SMT and NODDI images of MR lesion positive patients. Following standard radiological practice, they compared the different contrasts in the same patient; this allowed them to visually assess lesion location and relative conspicuity in the different image types. A lesion visibility score from 1 to 4 (1=not visible, 2=subtle, 3=visible, 4=clearly visible) was assigned to each image type for each patient. Both the observers were not blinded to radiological and EEG reports. Intensity windowing was individually adapted to gain optimal contrast. The neuro-radiologists assessed the images independently and so were blinded to each other's ratings.

In order to determine the level of agreement between the two raters, intra-class correlation coefficient (ICC) was estimated using a two-way random effects model based on two raters and absolute agreement<sup>26</sup> implemented in IBM SPSS v25. The ICC values along with 95% confidence intervals (CI) and p-value were reported for each image on Table 1. Interpretation of the ICC values was as follows: <0.50, poor agreement; between 0.50 and 0.75, moderate, between 0.75 and 0.90 good; above 0.90, excellent<sup>26</sup>. The ICC values were corrected for chance of agreement.

Aiming to compare lesion conspicuity scores between T1w, FLAIR, and diffusion images across patients, we applied Friedman test to the mean score of the two radiologists computed for each map. Then we performed a multiple comparison test between the ranking means provided by the Friedman test for each group of images. We set statistical significance at  $p < 0.05$  after applying the Bonferroni correction for multiple comparisons.

### Quantitative cortical and subcortical sampling

Figure 1 shows the workflow for the sampling of the diffusion data. FreeSurfer software v5.3<sup>27</sup> was used to co-register the diffusion maps and FLAIR to the T1w, and to reconstruct cortical-subcortical surfaces. Both FLAIR and T1w images were employed to generate the accurate smooth mesh representations of the pial surface<sup>27,28</sup>.

To examine the intracortical and subcortical signal of the diffusion maps, we sampled the diffusion values from the pial surface at steps of 0.5mm down to 6mm, guided by a straight line providing vertex correspondence across surfaces<sup>29</sup>. We note that the GM/WM border was not used because of the tendency for lesions to cause tissue mis-classification owing to its defining feature being a loss of GM/WM differentiation. The diffusion parameter map surfaces (sampled at different depths) were smoothed using a 10mm FWHM Gaussian kernel. Finally we registered the sampled diffusion maps to an average symmetric space having an identical number of vertices for each hemisphere<sup>30</sup>. This allowed us to analyse diffusion changes between homologous regions and therefore control for differences in cortical thickness due to anatomical variability by using an anatomically matched internal control.

### Lesion masks

FCD lesions were identified on T1w and FLAIR images by an experienced paediatric neuro-radiologist. 3D binary masks were manually delineated for the 33 patients. The lesion masks were first registered onto the surface reconstructions and then to the symmetric template. This procedure provided a mask for the lesion and one for the homologous healthy tissue.

## Statistical analysis

### *Lesion profiling*

MRI profiles for each diffusion map were obtained by averaging the values within the patient's lesion mask and homologous region, separately, along each sampling surface. To investigate signal changes in FCD lesions, the profiles of the diffusion maps were statistically compared between lesion and homologous region for the radiologically defined groups (33 patients).

We used a paired t-test to evaluate diffusion changes within each parameter map at various sampling depths. Correction for multiple comparisons was applied using false discovery rate (FDR) at  $p < 0.05$ . The data normality, required for the t-test, was assessed using the Shapiro-Wilk test run on the MRI profiles of the lesions and homologous regions.

### *Histological subtype profiling*

To quantitatively study pathology-specific changes in the diffusion maps, we repeated the *lesion profiling* analysis described above in the sub-group of patients with histologically confirmed FCD lesions (4 FCDIIa and 14 FCDIIb). The diffusion maps that showed a significant difference across cortical depth in the histologically defined group were employed to estimate the asymmetry of the MRI profiling by computing the diffusion values difference between the lesion and homologous region at each sampling depth.

The MRI profiling asymmetry measures were compared between patients with histologically confirmed FCDIIa and IIb using two-sample t-test. Correction for multiple comparisons was applied using FDR at  $q < 0.05$ .

*Correlation between visual scores and lesion profiling*

The correlation between the mean visual score of the two radiologists computed for each diffusion map, and the asymmetry of the MRI profiling was estimated using the Spearman correlation coefficient as described in the Supplementary material. This was performed in the radiologically defined group (n=33).

**Results**Patients' clinical information

The patients' demographics and clinical information can be found in Table 2 in Supplementary material. A total of 25/33 patients underwent surgery: 18 were histologically diagnosed as FCD (14 FCDIb, 4 FCDIIa), one had minimally invasive surgery with thermal ablation hence no histology is available, one had polymicrogyria, two had glioneuronal tumours, and three were diagnosed with hippocampal sclerosis (two had standard temporal lobe resection involving the anterior temporal lobe, one had resection also in the cingulate which did not exhibit any histopathological abnormality). Seizure freedom was achieved in 21/25 cases at 1.5 years after surgery.

Visual assessment

Figure 2 shows examples of FCDIIa and Iib lesions clearly visible on ICVF, INVf,  $\mu$ MD,  $\mu$ RD and  $\mu$ AD maps. Compared to the best visualisation achieved in either FLAIR or T1w, the lesions conspicuity was visually assessed as being better/equal/worse: on the ICVF in respectively 5/14/14 individuals, on the INVf in 3/3/27, on the  $\mu$ RD in 3/1/29, on the  $\mu$ MD in 2/3/28, and on the  $\mu$ AD in 1/1/31 patients. Similarly to the ICVF map, the lesion conspicuity

on the T1w was better/equal/worse in 5/11/17 cases compared to FLAIR.

Compared to the best visualisation achieved between FA and MD maps the lesions conspicuity was visually assessed as being better/equal/worse: on the ICVF maps respectively in 27/4/2 cases, on the INVf in 20/7/6, on the  $\mu$ MD in 10/12/11, on the  $\mu$ RD in 17/10/6, and on the  $\mu$ AD in 11/10/12 patients.

The mean scores of lesion conspicuity for each image type and patient can be found in Table 3 in Supplementary material.

The ICC index showed a significant ( $p < 0.05$ ) moderate agreement ( $0.5 < \text{ICC} < 0.75$ ) for the clinical FLAIR MPRAGE, ICVF, and the majority of diffusion maps, as shown in Table 1. The  $\mu$ FA,  $\mu$ AD, FA and MD exhibited poor agreement ( $\text{ICC} < 0.5$ ), as reported in Table 1.

The lesion conspicuity was scored significantly different across image types ( $p < 0.001$ ). The post-hoc multiple comparison test applied to the Friedman ranking test showed that the mean ranking of FLAIR, T1w, and ICVF maps was not significantly different, while the mean ranking of FA, MD, other NODDI maps, multi-compartment microscopic and microscopic SMT was significantly reduced compared to FLAIR images ( $p < 0.05$ ) (see Fig. 3).

### Quantitative lesion profiling

We observed significant ( $p_{\text{FDR}} < 0.05$ ) diffusion value changes between the suspected FCD lesions and their homologous regions on the ICVF, INVf,  $\mu$ MD,  $\mu$ AD and  $\mu$ RD maps for the entire radiologically defined cohort (see Fig.4a). Significant signal reduction was found within the lesions on the ICVF and INVf maps at 2-5.5mm depth for both groups (see Fig.4a). The  $\mu$ MD,  $\mu$ AD and  $\mu$ RD were significantly increased in the lesion at 2-5mm cortical depth, with respect to the healthy homologous regions for both groups, as shown on Fig.4a.

No significant differences were observed for the other NODDI and SMT maps or on the DTI FA and MD images.

### Histological subtype profiling

The lesion profiling analysis performed on the histologically confirmed group showed similar results to the ones obtained for the radiologically defined group. Significant signal reduction was found within the lesions on the ICVF and IVNF maps at 2-5.5mm depth, while the  $\mu$ MD,  $\mu$ AD and  $\mu$ RD were significantly increased in the lesion with respect to the healthy homologous regions (see Fig.4b).

The asymmetry profile analysis, performed on those maps showing significant changes in lesion for both the radiologically and histologically confirmed groups, demonstrated that FCDIIb lesions had significant ( $p_{FDR} < 0.05$ ) increased asymmetry with respect to FCDIIa on the  $\mu$ MD and  $\mu$ RD (Fig. 5). The asymmetry profiling changes in type IIb involved all sampling depths. The asymmetry measures for type IIa showed subtle signal alterations at 1.5-4mm depth on the  $\mu$ RD, and at 1.5-3mm depth on the  $\mu$ MD.

FCDIIb demonstrated significant asymmetry changes with respect to FCDIIa on the ICVF and INVF maps (Fig. 5). The asymmetry profiles showed ICVF and INVF reduction in type IIb lesions with respect to homologous regions involving all sampling depths. While type IIa exhibited subtle ICVF asymmetry mainly located at 1.5-3mm depth and absence of signal changes on the INVF maps. No significant asymmetry differences between subtypes were found for the  $\mu$ AD.

As FA and MD did not show significant changes between the lesion and the healthy region in the histologically confirmed patients, those maps were not included in the histological subtype profiling analysis.

### Correlation between visual scores and lesion profiling

$\mu$ RD and  $\mu$ Extra-neurite RD maps had the highest correlation between visual scores and lesion profiling (respectively  $\rho=0.59$ ,  $\rho=0.56$   $p$ -value $<0.01$ ), while ICVF, INVF,  $\mu$ FA, FA and MD had correlation smaller than 0.5 at  $p$ -value $<0.05$  (see Table 4 in Supplementary material).

## **Discussion**

To our knowledge this is the first study evaluating the ability of multi-compartment diffusion maps based on SMT and NODDI models, to delineate and characterise suspected FCD lesions in a significant paediatric population with drug-resistant focal epilepsy. This was made possible by recent advances in MRI hardware, such as improved scanner gradient performance, and software, such as multi-band imaging sequences<sup>21,22</sup> that reduce the acquisition time of multi-shell diffusion data (~7 minutes) allowing for its incorporation into a clinical paediatric epilepsy protocol.

### Visual assessment

Based on the multiple comparison statistical tests, the ICVF map provided comparable FCD lesion conspicuity to FLAIR and T1w images, and improved it with respect to MD and FA maps. At the individual level, the lesion contrast on ICVF was enhanced in ~10% of patients, therefore this map is most likely to provide information in combination with FLAIR to detect and demarcate FCD. Radiological evaluation is strengthened by observing signal abnormalities in several image modalities<sup>20</sup>, therefore it is beneficial to combine the contrast of these new diffusion maps with FLAIR and T1-weighted images. As in many patients



conventional imaging based on visual evaluation is unable to pinpoint the epileptogenic lesion<sup>31,32</sup>, it is crucial to test if these new image contrasts may increase detection rates<sup>33</sup> particularly in MRI negative patients.

The moderate agreement observed between the qualitative scoring of the two raters is concordant with previous literature studies reporting similar results for T2-weighted, FLAIR and T1w<sup>34</sup>. Differing assessments of the diffusion maps could potentially be explained by the relative lack of experience in viewing these contrasts and lower resolution of the diffusion maps (approximately half) compared to FLAIR and T1w images.

### Quantitative lesion profiling

In agreement with the visual analysis, quantitative investigation of ICVF, and additionally INVF,  $\mu$ MD,  $\mu$ RD and  $\mu$ AD parameters showed significant alterations in suspected and histologically confirmed FCD lesions at different sampling depths. In agreement with the previous study of 5 adult patients with suspected FCD<sup>15</sup>, we observed decreased ICVF signal in the lesions. Similarly, we found significant INVF decrease in suspected and confirmed FCD lesions. Both ICVF and INVF have been proposed as biomarkers of highly anisotropic structures, such as neurons nuclei and glia, as suggested by previous studies on multiple sclerosis<sup>35</sup>, Alzheimer's<sup>36</sup> and Parkinson's disease<sup>37</sup>. The within-lesion decreases are concordant with altered extracellular diffusion and increased extra-neurite volume measures performed on histology samples from surgical resections<sup>38</sup>. Those phenomena could also reflect the formation of additional diffusion barriers that may arise from loss of cortical stratification, deposits of extracellular matrix molecules or morphological changes of astrocytic processes usually associated with tissue remodelling due to astrogliosis<sup>38</sup>.

However, there are differences in the estimation of these two maps. In contrast to NODDI ICVF, the SMT multi-compartment INVF estimates the intrinsic diffusivity from the data

instead of assuming it to be equal to a fixed value<sup>12,13</sup>. The differences in the number of cases where the ICVF improved lesion conspicuity with respect to the INVf might also be explained by the fact that the diffusion data was smoothed before estimating the SMT maps to reduce their sensitivity to Gibbs ringing artefacts. The smoothing could enhance potential partial volume effects between subjacent structures.

The increase in both  $\mu$ RD and  $\mu$ AD is in agreement with an increase of  $\mu$ MD and explains the lack of signal changes on the  $\mu$ FA maps. Since  $\mu$ RD and  $\mu$ AD parameters are thought to be more specific to key features of brain microanatomy, they might be able to identify signal changes underlying a defect of neurogenesis or the presence of altered cells such as dysmorphic neurons and balloon cells<sup>13,14</sup>. As malformations of cortical development affect the tissue microstructure underlying cortical layers and white matter tracts<sup>38,39</sup>, the NODDI and SMT maps could now better characterise the presence of decreased myelinated axons and neurites resulting in increased extra-cellular space<sup>6</sup>, or the presence of abnormal cells.

In contrast to some other studies we did not observe any significant changes on FA and MD maps for neither the radiological nor the histologically confirmed FCD lesions. This might be explained by the fact that those DTI maps are affected by healthy variability in underlying tissue properties including neuronal density, fibre orientation dispersion, axonal diameter and degree of myelination, and ignore the presence of multiple tissue components, which can hinder signal changes induced by pathological phenomena in our sample size.

In this study, we were primarily interested in characterising the presence of consistent changes in quantitative diffusion maps via MRI profiling. Due to their ability to probe tissue biophysical properties *in-vivo*, they have potential to bridge the gap between radiological assessment and *ex-vivo* histology<sup>17</sup>. For this reason, we did not apply the profiling analysis to FLAIR and T1w images which has already been characterised<sup>16</sup>. While these clinical images

provide good contrast in some FCD lesions, the intensity is not quantitative, limiting their specificity to microstructural tissue properties<sup>40</sup>.

### Histological subtype profiling / neurobiological interpretation

Moreover, we showed that multi-compartment diffusion maps could help the characterisation of subtypes as FCDIIb lesions exhibited enhanced signal changes on the ICVF, INVf,  $\mu$ MD and  $\mu$ RD maps compared to FCDIIa, where signal alterations were subtle and affected layers closer to the pial surface. The presence of balloon cells in deep cortical and subcortical layers in FCDIIb might explain the altered cellular density measured by ICVF and INVf maps and the disrupted diffusivity quantified by  $\mu$ MD and  $\mu$ RD maps. These results are in agreement with previous studies analysing cellular water diffusivity on histological samples<sup>38</sup>. As our study has been carried out on 4 FCDIIa lesions, and 14 FCDIIb, further investigations on larger patient populations are necessary to validate the ability of multi-compartment diffusion images to capture histopathology subtype. Never-the-less the consistent signal changes we have demonstrated in modest numbers (despite being the largest study of its type) strongly motivate further evaluation.

### Limitations and outlook

As the clinical trend is to perform surgical resection on younger patients with clearly defined epileptogenic regions and seizures that affect both quality of life and development, it is crucial to non-invasively characterise FCD lesions for surgical planning, seizure freedom and neuro-developmental outcome<sup>17,41</sup>.

The consistency of changes in ICVF,  $\mu$ MD,  $\mu$ RD and  $\mu$ AD, and INVf for both the radiologically defined lesions and the subset with histological confirmation indicate that these maps could be useful for the visual identification of lesions and the integration into

algorithms for automated detection<sup>42</sup>. Further, some evidence of changes specific to FCD subtype was found that may indicate that these measures have the potential to reveal underlying tissue properties in FCD lesions non-invasively. Future studies are needed to validate these maps in larger cohorts including MRI-negative patients.

In this work, NODDI and SMT techniques were applied to investigate changes in both white and cortical grey matter. Those models rely on strong assumptions, if the tissue properties significantly differ from those constraints, the model can lead to erroneous interpretations, as might be the case in cortical grey matter<sup>43</sup>. Despite criticisms raised regarding the model assumptions<sup>43</sup>, NODDI is frequently used to study the human cortex<sup>44–46</sup>.

In conclusion, we have demonstrated that the multi-compartment diffusion maps showed changes in FCD lesions compatible with underlying disrupted tissue microstructure and could be valuable features for characterising the affected area and identifying the histological subtypes.

### **Study funding and acknowledgement**

This research was funded by the Henry Smith Charity and Action Medical Research (GN2214). This research was supported by the NIHR Great Ormond Street Hospital Biomedical Research Centre. The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR or the Department of Health. DC & SL are supported by the King's College London Wellcome/EPSRC Centre for Medical Engineering [WT 203148/Z/16/Z]. SA received funding from the Rosetrees Trust. TSJ receives funding from Great Ormond Street Children's Charity, The Brain Tumour Charity, Children with Cancer UK, Cancer Research UK and the Olivia Hodson Cancer Fund.

We would like to thank the Centre for Magnetic Resonance Research at the University of Minnesota for providing the multiband-EPI sequence (<http://www.cmrr.umn.edu/multiband>) used in this work.

None of the authors has any conflict of interest to disclose.

We confirm that we have read the Journal's position on issues involved in ethical publication and affirm that this report is consistent with those guidelines.

### **Key Points**

- We tested new advanced diffusion MRI models for the characterisation of FCD lesions.
- New diffusion maps of the intra-cellular volume fraction and per-axon mean, radial, axial diffusivity are more specific to lesion changes than FA and MD.
- The intra-cellular volume fraction and per-axon mean, radial diffusivity maps show different changes across FCD histological subtypes in 18 patients.
- These new diffusion maps have potential to improve pre-surgical epilepsy MRI protocols by enhancing the characterisation of FCD lesions.

## Tables

Maps	ICC		
	Mean value	p-value	95% confidence interval Lower – upper bound
<b>FLAIR</b>	0.75	$<10^{-5}$	0.51 – 0.88
<b>MPRAGE</b>	0.69	0.001	0.38 – 0.84
<b>ICVF</b>	0.51	0.012	0.05 – 0.75
<b>ODI</b>	0.52	0.021	0.03 – 0.76
<b><math>\mu</math>FA</b>	0.49	0.035	0.01 – 0.75
<b><math>\mu</math>AD</b>	0.48	0.031	0.1 – 0.74
<b><math>\mu</math>MD</b>	0.58	0.003	0.153 – 0.79
<b><math>\mu</math>RD</b>	0.64	0.001	0.29 – 0.82
<b>Diff</b>	0.79	$<10^{-5}$	0.56 – 0.9
<b><math>\mu</math>Extra-neurite MD</b>	0.66	0.001	0.32 – 0.83
<b><math>\mu</math>Extra-neurite RD</b>	0.77	$<10^{-5}$	0.51 – 0.89
<b>INVf</b>	0.67	0.001	0.33 – 0.83
<b>FA</b>	0.26	0.129	--
<b>MD</b>	0.45	0.007	0.01 – 0.73

**Table 1: Measure of inter-rater agreement on visual scores.** Two expert neuro-radiologists scored the FCD lesion visibility on FLAIR, MPRAGE, NODDI, microscopic and multi-compartment microscopic SMT, and standard DTI images. Inter-rater agreement was assessed using intra-class coefficient (ICC), corrected for chance of agreement. For each image we report the index value, the p-value and the confidence interval for  $\alpha=95\%$ . Significance level was set at  $p\text{-value}<0.05$ . ICVF=intracellular volume fraction, ODI = orientation dispersion index,  $\mu$ FA = microscopic fractional anisotropy,  $\mu$ AD = microscopic axial diffusivity,  $\mu$ MD =  $\mu$  mean diffusivity,  $\mu$ RD = microscopic radial diffusivity, Diff = intrinsic diffusivity,  $\mu$ Extra-neurite MD= extra-neurite microscopic mean diffusivity,  $\mu$ Extra-neurite RD= extra-neurite microscopic radial diffusivity, INVf = intra-neurite volume fraction.

## Figures

**Figure 1: Maps sampling workflow.** First the new diffusion maps were coregistered to the T1-weighted (T1w) data. Manual lesion masking and surface extraction at increasing depths were performed on T1w and FLAIR images. The diffusion maps were projected onto the surfaces and sampled from the pial surface. Then the surface sampling and the lesion masks were spatially registered to a symmetric template, allowing the symmetrical MRI profiling of the diffusion maps along the normal (black arrow) to the pial surface for the lesion and the homotopic region.

**Figure 2: Diffusion models and examples of diffusion parameters maps obtained in clinical setting.** Example of FLAIR, NODDI intracellular volume fraction (ICVF), SMT multi-compartment microscopic intra-neurite volume fraction (INVF), SMT microscopic MD, radial and axial diffusivities ( $\mu$ RD and  $\mu$ AD), standard DTI FA and MD maps for three patients. (left) FCD IIb patient with very-well delineated lesion on all images except FA. (centre) FCD IIa patient, lesion is poorly delineated by FLAIR, FA and MD but visible on ICVF and  $\mu$ RD maps. (right) FCD IIb patient, the lesion is poorly delineated by FLAIR, FA, MD, but very well visible on ICVF,  $\mu$ MD,  $\mu$ RD and  $\mu$ AD maps.

**Figure 3: Multiple comparison test on the mean ranks of the image scores.** The graph shows that the T1w, and NODDI intracellular volume fraction (ICVF) have mean ranks not significantly different (blue) from FLAIR, while the other diffusion maps have mean ranks significantly different (red) from FLAIR images. Circles represent mean rank values, error bars correspond to standard deviations. ODI = orientation dispersion index,  $\mu$ FA = microscopic fractional anisotropy,  $\mu$ AD = microscopic axial diffusivity,  $\mu$ MD = microscopic mean diffusivity,  $\mu$ RD = microscopic radial diffusivity, extra-neurite MD= extra-neurite microscopic mean diffusivity, extra-neurite RD= extra-neurite microscopic radial diffusivity,

INVF = intra-neurite volume fraction.

**Figure 4: MRI-profiling of FCD lesions and homotopic regions.** (a) MRI-profiling of FCD lesions (red) and homotopic regions (green) for the new diffusion maps showing significant profile changes at false discovery rate (FDR)  $<0.05$  (\*), and for MD, FA maps not providing significant differences for the whole patients cohort. The sampling depth is reported as a distance from the pial surface (0mm). (b) MRI-profiling of FCD lesions (red) and homotopic (green) regions on histologically confirmed patients showing significant (false discovery rate (FDR)  $<0.05$ ) profile changes for all sampling depths on new diffusion maps in contrast to MD, FA maps. ICVF = intra-cellular volume fraction, INVF = intra-neurite volume fraction,  $\mu$ MD =  $\mu$  mean diffusivity,  $\mu$ RD =  $\mu$  radial diffusivity,  $\mu$ AD =  $\mu$  axial diffusivity, MD = mean diffusivity, FA = fractional anisotropy. Error bars correspond to standard deviations computed over the cohort of patients.

**Figure 5: MRI-profiling of the asymmetry for FCD IIa and IIb patients.** Significant asymmetry differences at false discovery rate (FDR)  $<0.05$  (\*), between histologically confirmed FCD IIb (black) and FCD IIa (red) patients were found on the ICVF, INVF,  $\mu$ MD and  $\mu$ RD. The asymmetry was estimated by computing the difference vertex-wise between the lesion and homotopic region values using the maps showing significant changes between the two regions. The sampling depth is reported as a distance from the pial surface (0mm). ICVF = intra-cellular volume fraction, INVF = intra-neurite volume fraction,  $\mu$ MD =  $\mu$  mean diffusivity,  $\mu$ RD =  $\mu$ Radial diffusivity,  $\mu$ AD =  $\mu$ Axial diffusivity. Error bars correspond to standard deviations computed over the cohort of patients.



## References

1. Blümcke I, Thom M, Aronica E, et al. The clinicopathologic spectrum of focal cortical dysplasias: a consensus classification proposed by an ad hoc Task Force of the ILAE Diagnostic Methods Commission. *Epilepsia*. 2011; 52:158–74.
2. Harvey AS, Cross JH, Shinnar S, et al. Defining the spectrum of international practice in pediatric epilepsy surgery patients. *Epilepsia*. 2008; 49:146–55.
3. Duncan JS. Imaging in the surgical treatment of epilepsy. *Nat Rev Neurol*. 2010; 6:537–50.
4. Téllez-Zenteno JF, Hernández Ronquillo L, Moien-Afshari F, et al. Surgical outcomes in lesional and non-lesional epilepsy: a systematic review and meta-analysis. *Epilepsy Res*. 2010; 89:310–8.
5. Kim YH, Kang H-C, Kim D-S, et al. Neuroimaging in identifying focal cortical dysplasia and prognostic factors in pediatric and adolescent epilepsy surgery. *Epilepsia*. 2011; 52:722–7.
6. Eriksson SH, Rugg-Gunn FJ, Symms MR, et al. Diffusion tensor imaging in patients with epilepsy and malformations of cortical development. *Brain J Neurol*. 2001; 124:617–26.
7. Fonseca VCMs, Yasuda CLM, Tedeschi GG, et al. White matter abnormalities in patients with focal cortical dysplasia revealed by diffusion tensor imaging analysis in a voxelwise approach. *Epilepsy*. 2012; 3:121.
8. Wang Y, Zhou Y, Wang H, et al. Voxel-based automated detection of focal cortical dysplasia lesions using diffusion tensor imaging and T2-weighted MRI data. *Epilepsy Behav EB*. 2018; 84:127–34.
9. Liu W, Yan B, An D, et al. Perilesional and contralateral white matter evolution and integrity in patients with periventricular nodular heterotopia and epilepsy: a longitudinal diffusion tensor imaging study. *Eur J Neurol*. 2017; 24:1471–8.
10. Dumas de la Roque A, Oppenheim C, Chassoux F, et al. Diffusion tensor imaging of partial intractable epilepsy. *Eur Radiol*. 2005; 15:279–85.
11. Rugg-Gunn FJ, Eriksson SH, Symms MR, et al. Diffusion tensor imaging of cryptogenic and acquired partial epilepsies. *Brain J Neurol*. 2001; 124:627–36.
12. Zhang H, Schneider T, Wheeler-Kingshott CA, et al. NODDI: practical in vivo neurite orientation dispersion and density imaging of the human brain. *NeuroImage*. 2012; 61:1000–16.
13. Kaden E, Kelm ND, Carson RP, et al. Multi-compartment microscopic diffusion imaging. *NeuroImage*. 2016; 139:346–59.
14. Kaden E, Kruggel F, Alexander DC. Quantitative mapping of the per-axon diffusion coefficients in brain white matter. *Magn Reson Med*. 2016; 75:1752–63.

15. Winston GP, Micalef C, Symms MR, et al. Advanced diffusion imaging sequences could aid assessing patients with focal cortical dysplasia and epilepsy. *Epilepsy Res.* 2014; 108:336–9.
16. Hong S-J, Bernhardt BC, Caldairou B, et al. Multimodal MRI profiling of focal cortical dysplasia type II. *Neurology.* 2017; 88:734–42.
17. Adler S, Lorio S, Jacques TS, et al. Towards in vivo focal cortical dysplasia phenotyping using quantitative MRI. *NeuroImage Clin.* 2017; 15:95–105.
18. Yang G, Tian Q, Leuze C, et al. Double diffusion encoding MRI for the clinic. *Magn Reson Med.* 2018; 80:507–20.
19. Hirfanoglu T, Gupta A. Tuberous Sclerosis Complex With a Single Brain Lesion on MRI Mimicking Focal Cortical Dysplasia. *Pediatr Neurol.* 2010; 42:343–7.
20. Bernasconi A, Cendes F, Theodore WH, et al. Recommendations for the use of structural magnetic resonance imaging in the care of patients with epilepsy: A consensus report from the International League Against Epilepsy Neuroimaging Task Force. *Epilepsia.* 2019; 60:1054–68.
21. Setsompop K, Cohen-Adad J, Gagoski BA, et al. Improving diffusion MRI using simultaneous multi-slice echo planar imaging. *NeuroImage.* 2012; 63:569–80.
22. Setsompop K, Gagoski BA, Polimeni JR, et al. Blipped-controlled aliasing in parallel imaging for simultaneous multislice echo planar imaging with reduced g-factor penalty. *Magn Reson Med.* 2012; 67:1210–24.
23. Andersson JLR, Skare S, Ashburner J. How to correct susceptibility distortions in spin-echo echo-planar images: application to diffusion tensor imaging. *NeuroImage.* 2003; 20:870–88.
24. Smith SM. Fast robust automated brain extraction. *Hum Brain Mapp.* 2002; 17:143–55.
25. Gudbjartsson H, Patz S. The Rician Distribution of Noisy MRI Data. *Magn Reson Med.* 1995; 34:910–4.
26. McGraw KO, Wong SP. Forming inferences about some intraclass correlation coefficients. *Psychol Methods.* 1996; 1:30–46.
27. Fischl B, Dale AM. Measuring the thickness of the human cerebral cortex from magnetic resonance images. *Proc Natl Acad Sci U S A.* 2000; 97:11050–5.
28. Ségonne F, Pacheco J, Fischl B. Geometrically accurate topology-correction of cortical surfaces using nonseparating loops. *IEEE Trans Med Imaging.* 2007; 26:518–29.
29. Polimeni JR, Fischl B, Greve DN, et al. Laminar analysis of 7T BOLD using an imposed spatial activation pattern in human V1. *NeuroImage.* 2010; 52:1334–46.
30. Greve DN, Van der Haegen L, Cai Q, et al. A surface-based analysis of language lateralization and cortical asymmetry. *J Cogn Neurosci.* 2013; 25:1477–92.

31. Bernasconi A, Bernasconi N, Bernhardt BC, et al. Advances in MRI for ‘cryptogenic’ epilepsies. *Nat Rev Neurol.* 2011; 7:99–108.
32. Von Oertzen J, Urbach H, Jungbluth S, et al. Standard magnetic resonance imaging is inadequate for patients with refractory focal epilepsy. *J Neurol Neurosurg Psychiatry.* 2002; 73(6):643–7.
33. Wellmer J, Quesada CM, Rothe L, et al. Proposal for a magnetic resonance imaging protocol for the detection of epileptogenic lesions at early outpatient stages. *Epilepsia.* 2013; 54:1977–87.
34. Bernasconi A, Antel SB, Collins DL, et al. Texture analysis and morphological processing of magnetic resonance imaging assist detection of focal cortical dysplasia in extra-temporal partial epilepsy. *Ann Neurol.* 2001; 49:770–5.
35. Granberg T, Fan Q, Treaba CA, et al. In vivo characterization of cortical and white matter neuroaxonal pathology in early multiple sclerosis. *Brain.* 2017; 140:2912–26.
36. Parker TD, Slaterry CF, Zhang J, et al. Cortical microstructure in young onset Alzheimer’s disease using neurite orientation dispersion and density imaging. *Hum Brain Mapp.* 2018; 39:3005–17.
37. Kamagata K, Zalesky A, Hatano T, et al. Gray Matter Abnormalities in Idiopathic Parkinson’s Disease: Evaluation by Diffusional Kurtosis Imaging and Neurite Orientation Dispersion and Density Imaging. *Hum Brain Mapp.* 2017; 38:3704–3722.
38. Vargova L, Homola A, Cicanic M, et al. The diffusion parameters of the extracellular space are altered in focal cortical dysplasias. *Neurosci Lett.* 2011; 499:19–23.
39. Colombo N, Salamon N, Raybaud C, et al. Imaging of malformations of cortical development. *Epileptic Disord Int Epilepsy J Videotape.* 2009; 11:194–205.
40. Lorio S, Kherif F, Ruef A, et al. Neurobiological origin of spurious brain morphological changes: A quantitative MRI study. *Hum Brain Mapp.* 2016; 37:1801–15.
41. Choi SA, Kim SY, Kim H, et al. Surgical outcome and predictive factors of epilepsy surgery in pediatric isolated focal cortical dysplasia. *Epilepsy Res.* 2018; 139:54–9.
42. Adler S, Wagstyl K, Gunny R, et al. Novel surface features for automated detection of focal cortical dysplasias in paediatric epilepsy. *NeuroImage Clin.* 2017; 14:18–27.
43. Lampinen B, Szczepankiewicz F, Mårtensson J, et al. Neurite density imaging versus imaging of microscopic anisotropy in diffusion MRI: A model comparison using spherical tensor encoding. *NeuroImage.* 2017; 147:517–31.
44. Colgan N, Siow B, O’Callaghan JM, et al. Application of neurite orientation dispersion and density imaging (NODDI) to a tau pathology model of Alzheimer’s disease. *NeuroImage.* 2016; 125:739–44.
45. Eaton-Rosen Z, Melbourne A, Orasanu E, et al. Longitudinal measurement of the developing grey matter in preterm subjects using multi-modal MRI. *NeuroImage.* 2015; 111:580–9.

46. Nazeri A, Mulsant BH, Rajji TK, et al. Gray Matter Neuritic Microstructure Deficits in Schizophrenia and Bipolar Disorder. *Biol Psychiatry*. 2017; 82:726–36.