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DOI:

[10.1159/000502452](https://doi.org/10.1159/000502452)

*Document Version*

Peer reviewed version

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*Citation for published version (APA):*

Xu, Q. (2019). The renal collecting duct rises to the defence: Collecting duct as a critical renal protector. *Nephron*, 143(2), 148-152. <https://doi.org/10.1159/000502452>

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# The Renal Collecting Duct Rises to the Defence

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**Short Title:** Collecting duct as a critical renal protector

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## Keywords

Collecting duct · Retinoic acid · Protecting mechanisms · Acute kidney injury · Chronic kidney disease

## **Abstract**

When injury occurs, it implies that attack has overcome defence. Tubulointerstitial injury plays important roles in acute kidney injury (AKI) and chronic kidney disease (CKD), and is the common pathway leading to end-stage renal disease, but how the renal tubulointerstitium defends against attack is poorly understood. Emerging evidence suggests that collecting ducts (CDs), which modify urine from nephrons and drain into ureter, could be key defenders protecting tubulointerstitium from injury; furthermore, the canonical renal vitamin A signalling physiologically confined to CDs could be a key regulator of this protective machinery. This hypothesis can be tested by *in-vitro*, *in-vivo* and clinical studies, particularly by repressing or boosting key molecular regulators in CDs, to observe the resulting phenotypes in models of AKI and CKD. Further investigation of this hypothesis could lead to new strategies for diagnosis, prevention and treatment of AKI and CKD.

## **Background**

Kidneys and renal medicine are often regarded as the “victims of their own success”. Kidneys have remarkable functional reserves and renal medicine is among the most successful in replacement therapies when kidneys do fail [1]. Patients with end-stage renal disease (ESRD) on dialysis or kidney transplantation can maintain life, but these therapies are costly, donor organs are in short supply and mortality rates are high—young patients (20-49 years old) on these therapies are 16-25 times more likely to die in one year than the age-matched general population [2]. Rates of acute kidney injury (AKI) and chronic kidney disease (CKD) are increasing worldwide and the intercalated AKI-CKD syndrome continues to cause more ESRD and mortality [3,4]. The nephrology community has been promoting the “Oby25” initiative for AKI (zero preventable deaths by 2025)” [5], but few effective strategies exist to prevent AKI, CKD and the progression to ESRD.

Why are AKI and CKD such intractable problems? To answer this question, advice from past presidents of the American Society of Nephrology is worth noting: we must “emphasise innovation” [6] and “build new paths to kidney health” [7].

“Modern medicine can learn from traditional practices”, concluded GP-TCM, European Union’s first ever Coordination Action on modernisation of traditional Chinese medicine (TCM) under its Seventh Framework Programme [8]. As a nephrologist and the principal investigator of the consortium, I think that innovation in modern nephrology could be catalysed by the wisdom from *Yellow Emperor’s Inner Canon*, a TCM classic inscribed in the Memory of the World Register, United Nations Educational, Scientific and Cultural Organization [9], which says: “When there is sufficient protective *qi* inside, pathogenic factors have no way to hurt the body; when pathogenic factors cause damage, the internal *qi* must be deficient.” Herein, *qi* means the internal mechanisms of the body that defend against attack.

However, in nephrology most of the past work on AKI and CKD focused on how attacks damage, with relatively less consideration of the defending mechanisms in the kidney. In particular, although tubulointerstitial injury plays important roles in AKI [4] and progressive tubulointerstitial damage is the common pathway to ESRD [10], the protective mechanisms by which the renal tubulointerstitium defends against a variety of injurious stimuli is relatively less known.

### **Hypothesis**

Inspired by recent findings of my laboratory [11-14], I hypothesise that collecting ducts (CDs) are key to the defence of the tubulointerstitium and are regulated by the canonical vitamin A signalling mediated by retinoic acid (RA) and retinoic acid receptors (RARs).

### **Evidence supporting a key defence role for CDs**

Studies on AKI and CKD have traditionally focused on glomeruli, proximal tubules and the vasculature, leaving distal tubules and CDs largely neglected. In particular, CDs are the only tubules that span much of the kidney and are ideally placed to protect the whole tubulointerstitial compartment.

Importantly, in addition to principal cells that regulate water homeostasis and intercalated cells that regulate acid-base balance, CDs are equipped with specialised pro-repair and pro-regenerative mesenchymal stem cells [15] and defence molecules, e.g. antimicrobial  $\beta$ -defensins [16] and anti-fibrotic microRNAs [17]. Expression of many genes crucial for renal development, e.g. *Pax2*, *Wnt4* and *Wnt7b*, is also confined to CDs in the adult kidney [18]. Could these genes adapt to protective roles? Furthermore, CDs are equipped with RA/RAR physiological signalling [11,12]. A defence role for RA/RARs is highlighted by the susceptibility to pyelonephritis and urolithiasis in rats fed a vitamin A-deficient diet [19]. Since RA is effective in treating many models of AKI and CKD [20-22], endogenous RA/RARs in CDs may have been evolutionarily selected for protection against injury. To pinpoint roles for RA/RARs in CD cells, my laboratory has catalogued RA/RAR-dependent mRNAs in CD cells at a pan-genomic level, which support a critical role for the RA/RAR signalling in maintaining gene expression implicated in defence against infection (*Ppbb*, *Lcn2*, *Upk3b*), inflammation (*Ppbb*, *Cpm*, *Muc20*) and fibrosis (*Bmp7*) [11]. Thus, RA/RAR signalling in CDs, including its target genes, may serve as an important regulator for defence against tubulointerstitial damage.

Emerging evidence suggests that renal RA/RAR signalling plays important roles in AKI and CKD. In resolving AKI, CDs are relatively spared and renal RA/RAR activity is increased [21]. Conversely, in progressive CKD induced by unilateral ureteral obstruction, renal RAR expression decreases [22], and in mice with diabetic nephropathy there is a kidney-specific impairment of RA/RAR signalling [23]. Work in my laboratory has shown that endogenous RA/RAR activity is conserved in both mouse and human CD cells, including cortical and medullary CD cells and CD-derived mesenchymal stem cells (Xu Q et al, unpublished data) [13]. Furthermore, albumin specifically and dose-dependently represses RA/RAR activity in CD cells, suggesting that albuminuria, the leading risk factor for CKD progression, may contribute to CKD progression by repressing RA/RAR activity in CDs (Xu Q et al, unpublished data) [13]. In addition, angiotensin II, aldosterone, endotoxin and high glucose, which all contribute to CKD, dose-dependently repress RA/RAR signalling in CD cells, while gentamicin and aristolochic acid, which are known to induce AKI, increase signalling in cultured CD cells (Xu Q et al, unpublished

data) [13]. Thus, RA/RAR signalling in CD cells appears to be a convergence point of regulation by AKI and CKD risk factors and may be a novel target for AKI and CKD prevention and treatment.

### **Central questions and key experiments proposed to test the hypothesis**

CDs are structurally and functionally important for the kidney. They serve to drain urine from nephrons to ureters and plays fundamental roles in maintaining fluid and electrolyte homeostasis. Thus, any experiments proposed to test the hypothesised new protective roles for CDs must first ensure that the classical structural and functional roles for CDs are preserved. To determine the protective roles for CDs, it is important to identify potential protective effectors in CDs and test their functions and such experiments must be designed carefully at the molecular, subcellular and cellular levels. To begin with, at the molecular level, I propose to test the roles for *Pax2*, *Wnt4*, *Wnt7b* and the RA/RAR signalling in CDs as protective effectors in AKI and CKD models [11-14,18], while other candidate genes and signals are to be identified. I would like to propose the following key questions: How do CD cells maintain resilience? How do CDs protect other tubulointerstitial cells? What are the roles for CD-specific gene expression, RA/RAR signalling and CD-derived microvesicles and mesenchymal stem cells in renal tubulointerstitial defence? How do these mechanisms interact? How is CD defence regulated in AKI and CKD? Is failure of CD defence a cause of more severe AKI, chronicity and CKD progression? Do CDs secrete defence molecules into the urine and express defence biomarkers in biopsied renal tissue? Can these biomarkers predict prognosis and guide treatment? Finally, can novel therapies be devised to prevent AKI, AKI transition to CKD, and CKD progression to ESRD by modulating CD defence?

To test whether and how CD cells, including principal cells, intercalated cells and CD-derived mesenchymal stem cells, protect other renal cells, co-culture of these CD-derived cells and other cells in the presence of various types of stress and nephrotoxins could be a useful *in-vitro* model. Conditional, CD-specific deletion, silencing or overexpression of *Pax2*, *Wnt4*, *Wnt7b* and other genes

of interest in animal models will establish *in-vivo* proof of concept of the roles of these genes in the CDs in AKI and CKD. To determine whether RA/RAR signalling in CDs plays an important role in AKI and CKD, spatiotemporal changes of renal RA/RAR activities in different AKI and CKD models could be visualised and quantified in RA/RAR activity reporter mice [12,24]; and as illustrated in Fig. 1, AKI and CKD models could be induced in transgenic mice in which RA/RAR signalling in CDs is conditionally repressed or boosted.

To examine the specificity of any proposed protective role for CDs, it is important to compare roles for specific genes and signals in CDs versus other tubules. For example, repressing microRNA biogenesis in CDs but not in proximal tubules leads to renal fibrosis, supporting a specific role for CDs in supplying anti-fibrotic microRNAs [17,25]. In contrast, deleting the TGF- $\beta$  type II receptor in either CDs or proximal tubules leads to similar pro-fibrotic phenotypes, suggesting that the anti-fibrotic effect of the TGF- $\beta$  type II receptor in CDs is not unique [26,27].

Many genes (e.g. *Pax2*, *Wnt4* and *Wnt7b*) and signals (e.g. the RA/RAR signalling) in CDs that are potential protectors are indispensable in renal development. Thus, silencing genes or repressing RA/RAR signalling non-conditionally may lead to malformation of the foetal kidney and embryonic lethality thus preventing further studies on their roles in adult AKI and CKD. To test my hypothesis, reliable conditional gene knockout or transgenes to facilitate conditional gene expression changes specifically in adult renal CDs will be needed. However, CD-specific gene modification remains a challenge. Promoters of *Hoxb7*, *Aqp2* and *Atp6v1b1* are most commonly used to selectively regulate gene expression in CDs, but the promoter activity of *Hoxb7* is not strictly confined to CDs [28]; *Aqp2* is only expressed in CD principal cells and CD-derived mesenchymal stem cells [15]; while *Atp6v1b1* is only expressed in CD intercalated cells and some hybrid cells [29]. Thus, it remains a challenge to find pan-CD cell-specific promoters. Based on single-cell transcriptomic analysis of different mouse CD cells [29], to target all major CD cells during adulthood, the best approaches may still be inducible deletion or overexpression of target genes mediated by *Hoxb7* promoter or both *Aqp2* and *Atp6v1b1* promoters.

Clinically and in animal models, whether CD-derived urinary microvesicles in AKI and CKD could report activation or failure of the defence capacity and thus predict disease prognosis is worthy of investigation. CDs and other tubules micro-dissected from biopsies of patients or animal models of AKI and CKD could be subjected to in-depth analysis to guide discovery of novel mediators of CD defence. Finally, retinoids and non-retinoid therapies mobilising protective mechanisms in CDs could be explored for the prevention and treatment of AKI and CKD.

## **Conclusions**

A new role for CDs in protecting the renal tubulointerstitium is proposed, supplementing their role in controlling urinary fluid and electrolyte composition. Further investigation of this hypothesis could lead to a paradigm shift in understanding kidney health and may lead to new strategies for diagnosis, prevention and treatment of AKI and CKD. Although this paper focuses on the kidney, CDs are also important in protecting the heart [30]. Given that non-immune defence mechanisms are generally overlooked in modern medicine, it is hoped that this special article will inspire not only nephrologists, but also other physicians and medical scientists alike, to perceive health and disease in light of the balance and imbalance between attack and defence.

## **Acknowledgement**

The author would like to thank Prof. Leon G Fine (Cedars-Sinai Medical Center, USA), Dr. Jeffrey Kopp (NIH, USA), Prof. Jill T Norman and Prof. Patricia Wilson (University College London, UK), Prof. Frederick Tam (Imperial College London, UK), Prof. Donald Fraser (Cardiff University, UK), as well as colleagues at King's College London (Dr. Alexandros Papadimitriou, Prof. Bruce M Hendry, Prof. Claire Sharpe and Prof. Sir Robert Lechler) for useful discussions. This Special Article is dedicated to the late Professor Peter John Hylands, former Head of Institute of Pharmaceutical Science, and former Co-Director of King's Centre for Integrative Chinese Medicine, King's College London, for his irreplaceable mentorship, invaluable collaboration and inspirational encouragement and support.

## **Disclosure Statement**



The author has no conflicts of interest to declare.

### **Funding Sources**

This work has been supported by Kidney Research UK and the European Union.

### **Author Contributions**

Dr Qihe Xu conceptualised and wrote this paper.

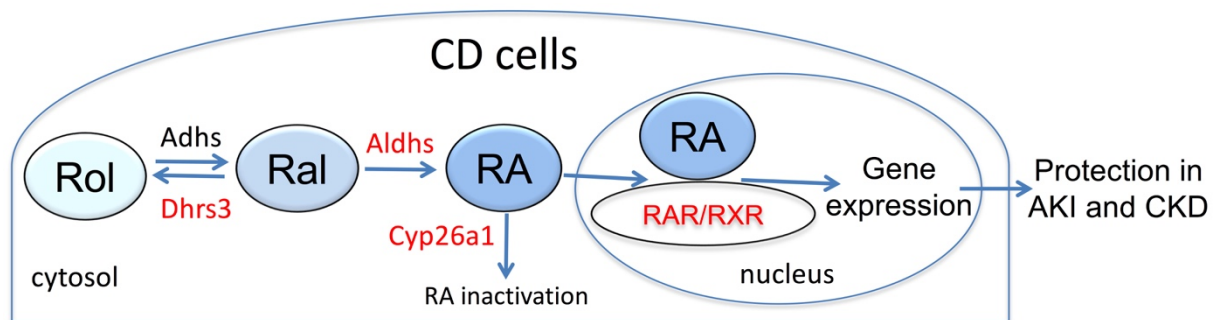
### **References**

- 1 The Lancet Editorial. Research in kidney disease: an acute and chronic history. *Lancet*. 2015 May 16;385(9981):1918.
- 2 Steenkamp R, Pyart R, Fraser S. UK Renal Registry 20th Annual Report: Chapter 5 Survival and cause of death in UK adult patients on renal replacement therapy in 2016: National and centre-specific analyses. *Nephron*. 2018;139 Suppl 1: 117-50.
- 3 GBD 2013 Mortality and Causes of Death Collaborators. Global, regional, and national age-sex specific all-cause and cause-specific mortality for 240 causes of death, 1990-2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet*. 2015 Jan 10;385(9963):117-71.
- 4 Chawla LS, Eggers PW, Star RA, Kimmel PL. Acute kidney injury and chronic kidney disease as interconnected syndromes. *N Engl J Med*. 2014 Jul 3;371(1):58-66.
- 5 Mehta RL, Cerdá J, Burdmann EA, Tonelli M, García-García G, Jha V, et al. International Society of Nephrology's Oby25 initiative for acute kidney injury (zero preventable deaths by 2025): a human rights case for nephrology. *Lancet*. 2015 June 27;385(9987):2616-43.
- 6 Molitoris BA. ASN Presidential Address 2013: Innovation and individualization—the path forward for nephrology. *J Am Soc Nephrol*. 2014 May;25(5):893-7.
- 7 Moe SM. ASN Presidential Address 2014: Moving past nephrology's midlife crisis. *J Am Soc Nephrol*. 2015 Apr;26(4):791-5.

- 8 European Commission [Internet]. Results in brief. Modern medicine can learn from traditional practices [cited 2019 June 24]. Available from: [https://cordis.europa.eu/result/rcn/91212\\_en.html](https://cordis.europa.eu/result/rcn/91212_en.html)
- 9 United Nations Education, Scientific and Cultural Organization [Internet]. Huang Di Nei Jing (Yellow Emperor's Inner Canon) [cited 2019 June 24]. Available from: <http://www.unesco.org/new/en/communication-and-information/memory-of-the-world/register/full-list-of-registered-heritage/registered-heritage-page-4/huang-di-nei-jing-yellow-emperors-inner-canon/>
- 10 Meola M, Samoni S, Petrucci I. Clinical scenarios in chronic kidney disease: Chronic tubulointerstitial diseases. In Meola M, Petrucci I, Ronco C (eds): Ultrasound Imaging in Acute and Chronic Kidney Disease. Contrib Nephrol. Basel, Karger, 2016;188:pp108-19.
- 11 Wong YF, Wilson PD, Unwin RJ, Norman JT, Arno M, Hendry BM, et al. Retinoic acid receptor-dependent, cell-autonomous, endogenous retinoic acid signaling and its target genes in mouse collecting duct cells. PLoS One. 2012 Sep 26;7(9):e45725.
- 12 Wong YF, Kopp JB, Roberts C, Scambler PJ, Abe Y, Rankin AC, et al. Endogenous retinoic acid activity in principal cells and intercalated cells of mouse collecting duct system. PLoS One. 2011 Feb 4;6(2): e16770.
- 13 Xu Q. Canonical vitamin A signalling in collecting duct cells controls expression of anti-fibrotic microRNAs and is differentially regulated by AKI/CKD mediators. UK Kidney Week 2018. Poster No. 213. [cited 2019 June 24]. Available from: <https://www.ukkw.org.uk/abstracts-from-uk-kidney-week/>
- 14 Xu Q. Renal collecting ducts: A frontline in protecting against tubulointerstitial injury? UK Kidney Week 2018. Poster No. 214. [cited 2019 June 24]. Available from: <https://www.ukkw.org.uk/abstracts-from-uk-kidney-week/>

- 15 Li J, Ariunbold U, Suhaimi N, Sunn N, Guo J, McMahon JA, et al. Collecting duct-derived cells display mesenchymal stem cell properties and retain selective in vitro and in vivo epithelial capacity. *J Am Soc Nephrol*. 2015 Jan;26(1):81-94.
- 16 Lehmann J, Retz M, Harder J, Krams M, Kellner U, Hartmann J, et al. Expression of human beta-defensins 1 and 2 in kidneys with chronic bacterial infection. *BMC Infect Dis*. 2002 Sep 18;2:20.
- 17 Hajarnis S, Yheskel M, Williams D, Brefort T, Glaudemans B, Debaix H, et al. Suppression of microRNA activity in kidney collecting ducts induces partial loss of epithelial phenotype and renal fibrosis. *J Am Soc Nephrol*. 2018 Feb;29(2):518-31.
- 18 Little MH, Kairath P. Does renal repair recapitulate kidney development? *J Am Soc Nephrol*. 2017 Jan;28(1):34-46.
- 19 Munday JS, McKinnon H, Aberdein D, Collett MG, Parton K, Thompson KG. Cystitis, pyelonephritis, and urolithiasis in rats accidentally fed a diet deficient in vitamin A. *J Am Assoc Lab Anim Sci*. 2009 Nov;48(6):790-4.
- 20 Xu Q, Lucio-Cazana J, Kitamura M, Ruan X, Fine LG, Norman JT. Retinoids in nephrology: promises and pitfalls. *Kidney Int*. 2004 Dec;66(6):2119-31.
- 21 Chiba T, Skrypnik NI, Skvarca LB, Penchev R, Zhang KX, Rochon ER, et al. Retinoic acid signaling coordinates macrophage-dependent injury and repair after AKI. *J Am Soc Nephrol*. 2016 Feb;27(2):495-508.
- 22 Long YB, Qin YH, Zhou TB, Lei FY. Association of retinoic acid receptors with extracellular matrix accumulation in rats with renal interstitial fibrosis disease. *Int J Mol Sci*. 2012 Oct 31;13(11):14073-85.
- 23 Starkey JM, Zhao Y, Sadygov RG, Haidacher SJ, Lejeune WS, Dey N, et al. Altered retinoic acid metabolism in diabetic mouse kidney identified by O isotopic labeling and 2D mass spectrometry. *PLoS One*. 2010 Jun 14;5(6):e11095.

- 24 Bilbija D, Haugen F, Sagave J, Baysa A, Bastani N, Levy FO, et al. Retinoic acid signalling is activated in the post-ischemic heart and may influence remodelling. *PLoS One*. 2012 Sept 28;7(9):e44740.
- 25 Wei Q, Bhatt K, He HZ, Mi QS, Haase VH, Dong Z. Targeted deletion of Dicer from proximal tubules protects against renal ischemia-reperfusion injury. *J Am Soc Nephrol*. 2010 May;21(5):756-61.
- 26 Gewin L, Bulus N, Mernaugh G, Moeckel G, Harris RC, Moses HL, et al. TGF- $\beta$  receptor deletion in the renal collecting system exacerbates fibrosis. *J Am Soc Nephrol*. 2010 Aug;21(8):1334-43.
- 27 Nlandu-Khodo S, Neelisetty S, Phillips M, Manolopoulou M, Bhave G, May L, et al. Blocking TGF- $\beta$  and  $\beta$ -catenin epithelial crosstalk exacerbates CKD. *J Am Soc Nephrol*. 2017 Dec;28(12):3490-503.
- 28 Pritchett TL, Bader HL, Henderson J, Hsu T. Conditional inactivation of the mouse von Hippel-Lindau tumor suppressor gene results in wide-spread hyperplastic, inflammatory and fibrotic lesions in the kidney. *Oncogene*. 2015 May 14;34(20):2631-9.
- 29 Chen L, Lee JW, Chou CL, Nair AV, Battistone MA, Păunescu TG, et al. Transcriptomes of major renal collecting duct cell types in mouse identified by single-cell RNA-seq. *Proc Natl Acad Sci U S A*. 2017 Nov 14;114(46):E9989-98.
- 30 Fujiu K, Shibata M, Nakayama Y, Ogata F, Matsumoto S, Noshita K, et al. A heart-brain-kidney network controls adaptation to cardiac stress through tissue macrophage activation. *Nat Med*. 2017 May;23(5):611-22.



**Fig. 1. A hypothesised protective role for the canonical vitamin A signalling in CDs and proposed points for experimental intervention in order to test the hypothesis [11,12, 20].** Vitamin A (RoI) is metabolised to RA via two oxidation steps. While the first step is reversibly catalysed by alcohol dehydrogenases (Adhs) 1-4 and dehydrogenase/reductase (SDR family) member 3 (DhRs3), respectively, the second step converting retinaldehyde (Ral) into RA is catalysed by aldehyde dehydrogenase family 1 subfamily a1-3 (Aldh1a1, a2 and a3) and is irreversible. RA binds RARs in heterodimers of retinoid X receptors (RXR) and RAR to regulate gene expression. In RAR/RXR heterodimers, RXRs are silent. Thus RA-activated RAR/RXR signalling is also simplified as RA/RAR signalling. By tetracycline-inducible, CD-specific promoter-driven overexpression of a dominant negative mutant RAR (to antagonise RARs) and an RA-metabolising enzyme, e.g. Cyp26a1 (to eliminate RA), the RA/RAR signalling in CDs can be repressed on demand—this approach will be suitable to examine roles for the RA/RAR signalling in CDs when it is relatively high; by conditional, CD-specific overexpression of *Aldhs* (to increase RA synthesis) or deletion of *DhRs3* (to increase RA precursors), the RA/RAR activity in CDs can be boosted—this approach will be suitable to examine roles for the RA/RAR signalling in CDs when it is repressed. Proposed points of intervention are highlighted by red fonts.