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1 **Title**

2 T2* placental MRI in pregnancies complicated with fetal congenital heart disease

3 **Authors**

4 Johannes K Steinweg ¹; Grace Tin Yan Hui ³; Maximilian Pietsch^{3,4}, Alison Ho ³; Milou
5 PM van Poppel ¹; David Lloyd ^{1,2}; Kathleen Colford ³; John M Simpson ^{1,2}, Reza Razavi
6 ^{1,2}, Kuberan Pushparajah ^{1,2}; Mary Rutherford ³, Jana Hutter ^{3,4}

7 **Affiliations**

- 8 1. Department of Cardiovascular Imaging, School of Biomedical Engineering &
9 Imaging Science, King's College London, London, United Kingdom
10 2. Department of Congenital Heart Disease, Evelina Children's Hospital, London,
11 United Kingdom
12 3. Centre for the Developing Brain, King's College London, London, United
13 Kingdom
14 4. Department of Biomedical Engineering, School of Biomedical Engineering &
15 Imaging Science, King's College London, London, United Kingdom

16 **Corresponding author**

17 johannes.steinweg@kcl.ac.uk

18 Johannes K Steinweg
19 Department of Cardiovascular Imaging
20 Wellcome/EPSRC Centre for Medical Engineering
21 School of Biomedical Engineering & Imaging Sciences
22 King's College London
23 3rd Floor Lambeth Wing
24 St. Thomas' Hospital
25 Westminster Bridge Road
26 London SE1 7EH
27 United Kingdom

28 **Abstract**

29 **Background**

30 Congenital heart disease (CHD) is one of the most important and common group of
31 congenital malformations in humans. Concurrent development and close functional
32 links between the fetal heart and placenta emphasise the importance of understanding
33 placental function and its influence in pregnancy outcomes. The aim of this study was
34 to evaluate placental oxygenation by relaxometry (T2*) to assess differences in
35 placental phenotype and function in CHD.

36 **Methods**

37 In this prospective cross-sectional observational study, 69 women with a fetus affected
38 with CHD and 37 controls, whole placental T2* was acquired using a 1.5-Tesla MRI
39 scanner. Gaussian Process Regression was used to assess differences in placental
40 phenotype in CHD cohorts compared to our controls.

41 **Results**

42 Placental T2* maps demonstrated significant differences in CHD compared to controls
43 at equivalent gestational age. Mean T2* values over the entire placental volume were
44 lowest compared to predicted normal in right sided obstructive lesions (RSOL) (Z-
45 Score 2.30). This cohort also showed highest lacunarity indices (Z-score -1.7), as a
46 marker of lobule size. Distribution patterns of T2* values over the entire placental
47 volume were positively skewed in RSOL (Z-score -4.69) and suspected, not confirmed
48 coarctation of the aorta (CoA-) (Z-score -3.83). Deviations were also reflected in
49 positive kurtosis in RSOL (Z-score -3.47) and CoA- (Z-score -2.86).

50 **Conclusion**

51 Placental structure and function appear to deviate from normal development in
52 pregnancies with fetal CHD. Specific patterns of altered placental function assessed by
53 T2* deliver crucial complementary information to antenatal assessments in the
54 presence of fetal CHD.

55 **Keywords**

56 Congenital Heart disease (CHD), Placenta, Magnetic resonance imaging (MRI), T2*
57 Mapping, Gaussian Process Regression (GPR), Machine learning

58

59 Introduction

60 Congenital heart disease (CHD) is a common group of congenital malformations with a
61 prevalence of up to 1% of all live births, a leading cause of neonatal and infant death
62 and a global burden in child health [1,2]. Underlying aetiology is considered
63 multifactorial combining genetic, epigenetic and environmental causes [3–5]. Recently,
64 placental vascular malperfusion has been reported at pathological examination in
65 association with CHD [6,7] and it has been postulated that this may contribute to the
66 neurodevelopmental abnormalities observed in children with CHD [8].

67 The placenta is the only connection between the fetal and maternal circulation. It plays
68 a pivotal role in fetal development due to its responsibility for all fetomaternal
69 exchange, including oxygen, carbon dioxide and nutrients, and essential immunological
70 and homeostatic functions. The placenta and fetal heart develop concurrently and
71 share similar developmental pathways – amplifying its unique role and vulnerability to
72 disturbances especially in the presence of fetal CHD [9,10]. A sophisticated in vivo
73 assessment of placental structure and function may inform understanding of complex
74 antenatal pathophysiology in pregnancies with fetal CHD and help to identify fetuses at
75 greater risk of adverse long term neurodevelopmental outcomes.

76 A common MR contrast mechanism employed to study tissue oxygen concentration is
77 produced by the blood oxygen level dependency (BOLD) effect, exploited in
78 quantitative T2 relaxometry (T2*). The parametric properties of deoxyhaemoglobin
79 allow for a faster T2* decay than its oxygenated counterpart [11,12]. Widely used in
80 functional brain MRI, it has also recently found application in placental MRI, allowing in-
81 vivo insights into the tissue properties without the use of exogenous contrast agents
82 [13–15].

83 Previous studies in humans have shown a linear negative correlation of placental mean
84 T2* values over gestational age (GA) and found it to be predictive for low birth weight
85 [15,16]. An increase in placental mean T2* was previously reported in response to
86 maternal oxygen administration [17,18]. Decreased mean T2* values can be induced
87 by vasoconstrictive agents in animal models and have been demonstrated in
88 pregnancies with fetal growth restriction in both animals and humans [19–21]. Reduced
89 whole placental mean T2* values for GA were recently also associated with
90 preeclampsia [22].

91 Mean T2* value calculated over the placental parenchyma, either assessed in selected
92 slices or the entire placental volume have been used for quantification in previous

93 studies. However, novel advances extend this to include spatial analysis using
94 histograms and texture-based measurements, first shown in rhesus monkeys [23,24].
95 This study aimed to evaluate whether CHD is associated with altered placental
96 phenotype and function, including signs of lower oxygenation and altered tissue
97 morphology, as estimated by high-resolution whole placental MR relaxometry (T2*) and
98 to establish methods to evaluate deviations from the expected values over GA.

99 **Methods**

100 In this prospective cross-sectional observational study, we recruited pregnant women
101 carrying a fetus diagnosed with congenital heart disease (CHD) from a tertiary fetal
102 cardiology service for CHD (St Thomas' Hospital, London, UK) alongside a control
103 group of women with uncomplicated pregnancies during the second and third trimester.
104 Inclusion criteria for both cohorts were singleton pregnancy, maternal age over 18
105 years and ability to consent in English language. Exclusion criteria for both cohorts
106 were multiple pregnancies, major maternal health issues, any treatment for
107 hypertension at time of scan as well as contraindications for MRI such as metallic
108 implants and claustrophobia. Exclusion criteria for controls also included diagnosis of
109 fetal or intrauterine growth restriction (FGR/IUGR) and low birth weight (SGA), signs of
110 preeclampsia or (gestational) diabetes at time of scan.

111 Prospectively specified data collection included demographic characteristics and
112 maternal pregnancy data at time of scan, as well as delivery notes and neonatal
113 outcomes. Maternal data included age and BMI at scan, parity, gravida, medical history
114 of smoking, (gestational) diabetes, hypertension, preeclampsia, HELLP syndrome,
115 thyroid disease and anxiety or depression as noted at time of scan and in their delivery
116 notes. Neonatal data included fetal and neonatal cardiac diagnosis, sex, GA at scan
117 and birth, birth weight, 5 min APGAR, neonatal outcome, genetic testing, suspicion of
118 genetic abnormality and place of birth as noted on the delivery or (cardiology)
119 discharge notes.

120 Centiles and standard scores for birth weight were calculated following the
121 INTERGROWTH-21st project [25]. SGA was defined as birthweight less than 10th
122 centile in keeping with common clinical practice. Our CHD cohort was classified into 7
123 groups based on the main antenatal diagnosis (Figure 1, Table 1). We divided our
124 cohort into left sided obstructive lesions, namely hypoplastic left heart syndrome
125 (HLHS) and coarctation of the aorta (CoA), right-sided obstructive lesions (RSOL)
126 defined as structural or functional obstruction of the right ventricular outflow tract,
127 disorders of mixing (e.g., transposition of the great arteries (TGA)), suspected vascular
128 rings (VR) and other major lesions comprising of common arterial trunk (CAT), partial
129 anomalous pulmonary venous drainage (PAPVD) and cardiac rhabdomyomas (CR).
130 The group of suspected CoA was further divided by those with postnatal confirmation
131 requiring surgery within the neonatal period (CoA+) and without postnatal CoA during
132 follow up (CoA-). All our infants are therefore followed up by our team at least until 1
133 year of age.

134 Ethics:

135 This study was approved by London Research Ethics Committees of the Health
136 Research Authority (HRA) of the Department of Health in the United Kingdom,
137 “Quantification of fetal brain growth and development using MRI” (REC:
138 07/H0707/105), “Fetal Imaging with Maternal Oxygen” (REC: 17/LO/0282) and “iFIND-
139 2. Further Imaging” (REC: 14/LO/1806). Controls were also included from the
140 “Placenta Imaging Project” (REC: 16/LO/1573).

141 MRI Acquisition:

142 All women were scanned using a clinical 1.5-Tesla Philips Achieva MRI with a 28-
143 channel torso coil under clinical monitoring and medical cover during the entire scan.
144 All mothers carrying a fetus with CHD were scanned in left lateral tilt. A multi-echo
145 gradient echo sequence covering the entire uterus in coronal orientation was acquired
146 with a resolution of 2.5mm isotropic, FOV=360x360, 50-88 slices, no SENSE, no half-
147 scan, TR=14s, TE=11.376ms / 57.313ms / 103.249ms / 149.186ms / 195.122ms, free
148 breathing, TA=1min.

149 MRI data processing:

150 T2* maps were calculated using an in-house monoexponential fitting in MATLAB (The
151 MathWorks Inc, USA). The whole placenta was manually segmented by experienced
152 observers (JKS, GH, AH, JH) using ITK-SNAP [26], with exactly matched instructions
153 to segment the placenta conservatively, avoiding inclusion of both amniotic fluid and
154 maternal vasculature. Reproducibility of manual segmentations between observers
155 was assessed using the Sørensen–Dice coefficient [27].

156 The masks were automatically refined by excluding non-physiological values. Mean
157 T2*, skewness, kurtosis and lacunarity values were obtained using a purpose-build
158 python script [13,28]. Lacunarity values thereby reflect the spatial distribution of gaps of
159 a specific size within lobules [29] and a box-size matched to typical placental lobule
160 size.

161 To estimate placental development over GA we used Gaussian Process Regression
162 (GPR), a Bayesian non-parametric regression [30]. Clinical cohorts are inherently
163 heterogenous necessitating an estimation of uncertainty of probable distributions.
164 Accounting for this and a covariance between the data points GPR provides normative
165 group mean function, allowing point estimates and predictive confidence for each
166 observation. This allows the calculation of standard (Z-) scores for all measured test
167 data points, describing the distance to expected value following the normative group
168 mean function [31].

169 The control cohort was split into training and test data (0.7 to 0.3 ratio) and the test
170 subjects were used to train the model. The kernel function used was an additive
171 combination of Constant function, Radial Basis Functions and Noise kernel. The data
172 was scaled pre-training. Predictive posterior distributions were obtained for mean T2*,
173 lacunarity, skewness and kurtosis separately. Z-scores for the observed values were
174 derived for all CHD cohorts by estimation of mean deviation and median deviation from
175 GPR predicted value normalised by the prediction uncertainty.
176 For comparison of clinical parameters controls were restricted to GA at scan between
177 28 and 37 weeks to match GA for CHD cohort, categorical variables were compared
178 using Fisher's exact test, continuous variables were evaluated for normality by Shapiro
179 compared using Independent T-test (normal distribution), 2-tailed Mann-Whitney U test
180 (non-normal distribution). Dependence of results from clinical parameters was
181 evaluated by coefficient of determination. All statistical analysis and visualisation were
182 performed using SPSS Statistics v27 (IBM) and Jupyter Notebook, python3.
183

184 Results

185 Maternal and neonatal demographics are demonstrated in Table 1. Overall, 119
186 participants were enrolled in this study. Eight subjects from our CHD cohort were
187 excluded: for twin pregnancy (n=1), insufficient scan data (n=3) and insufficient
188 outcome data at time of analysis (n=4). Five controls were excluded for small for GA at
189 birth (n=3) and insufficient scan data (n=2). In total 69 women with pregnancies with
190 fetal diagnosis of CHD and 37 women with uncomplicated pregnancies were included
191 in the analysis (Figure 1). The median GA at scan was 31.3 weeks (IQR 2.21) and 31.2
192 weeks (IQR 8.11) respectively. Results of genetic testing from antenatal invasive
193 procedures or postnatal blood sample using at least array comparative genomic
194 hybridization (aCGH) were available for 46 (67%) subjects in our CHD cohort. 5/46
195 identified an abnormal result (two with Chr 2q31 deletion, one with a Chr 9 deletion and
196 gene copy number on Chr 2, one with TSC1 gene mutation, and one with mosaic
197 Monosomy X). All remaining patients caused no phenotypic suspicion for a genetic
198 abnormality after birth.

199 Seven newborns with CHD from our cohort died due to necrotising enterocolitis (1),
200 hypoxic ischemic encephalopathy (1), congenital diaphragmatic hernia (1) and cardiac
201 collapse or palliation (4). The CHD cohort had a higher incidence of low APGAR score
202 (<7) at 5 minutes (p=0.024) compared to our control cohort. GA at birth (p<0.001) and
203 birth weight (p<0.001) and birth head circumference (p=0.005) were significantly lower
204 in the CHD cohort, whose mothers were also younger (p=0.017). None of the other
205 clinical collected parameters achieved clinical significance.

206 We have reviewed medical records at time of scan and delivery. Our CHD cohort
207 included three women diagnosed with preeclampsia (4.3%). Two needed delivery at 32
208 weeks of gestation, one for severe HELLP and one for uncontrollable hypertension.

209 One woman delivered after induction early term at 37 weeks of GA.

210 In our CHD cohort 18 (26%) newborns were born <10th birth weight centile (SGA), 25
211 (36%) newborns were born with low head circumference (HC) <10th centile, 13
212 newborns (19%) were SGA and had low HC. Only one of these 13, would be defined
213 as growth restricted following the consensus-based definition in the newborn by Beune
214 et al. [32].

215 Placental histology was available in 12 pregnancies complicated by fetal CHD on
216 special clinical request mostly due to maternal pyrexia during delivery (Supplemental
217 Table 1). From the two placentas demonstrating signs of maternal vascular
218 malperfusion (MVM), one placenta showed also acute subchorionitis and maternal

219 inflammatory response (MIR) in keeping with ascending intra-uterine infection showing
220 infarcts of variable age, but no thrombi. The other placenta was from a patient
221 diagnosed with severe preeclampsia and HELLP syndrome leading to early delivery.
222 Placental histology was also available for 15 control subjects as part of other study
223 protocols (Supplemental Table 1).

224 **Qualitative assessments**

225 Placental T2* maps in our cohort of RSOL showed most marked differences compared
226 to age-matched controls as depicted in mid-parenchymal slices (Figure 2). Specifically,
227 short T2* values were noted in the entire placenta with additional and faster decay from
228 the centre to the periphery of the lobules. Furthermore, increased heterogeneity could
229 be observed in RSOL. Our cohorts of left sided obstructive lesions, disorders of mixing
230 and other lesions appeared only moderately different to controls at similar GA (Figure
231 2). Overall, our CHD cohort appeared to have generally lower signal intensity
232 throughout the placenta, advanced lobularity and higher granularity within the lobules
233 at a given GA compared to our control cohort.

234 **Quantitative assessments**

235 Interobserver variability of manually segmented placental masks showed good
236 correlation in 10 randomly selected with a Sørensen–Dice coefficient of 0.87.
237 Quantitative results from the control cohort illustrate decay in mean T2* with increasing
238 GA, in keeping with previous literature [15,29]. Lacunarity, kurtosis and skewness tend
239 to increase over GA in all our cohorts as previously shown in controls [28]. The
240 obtained posterior mean of the Gaussian process is given for all quantitative values in
241 Figure 3.
242 Mean T2* values over the entire placental volume were lowest compared to predicted
243 normal in RSOL (Z-Score 2.30) and our cohort with other major lesions [CAT, PAPVD,
244 CR] (Z-Score 2.31). Our CoA- cohort had a larger deviation from expected values (Z-
245 Score 1.39) than CoA+ (Z-score 0.24). Mean T2* values for HLHS (Z-score 0.63), VR
246 (0.09) and TGA (Z-score -0.11) were within one standard deviation (SD) from expected
247 results.
248 RSOL (Z-score -1.7), our group of other major lesions (Z-score -1.26) and CoA- (Z-
249 score -1.02) showed significantly higher lacunarity compared to expected results at GA
250 equivalent. HLHS (Z-score -0.18), TGA (Z-score -0.01) and suspected VR (Z-score
251 0.06) were similar to expected controls. CoA+ showed slightly lower lacunarity (Z-score
252 0.26).

253 Distribution of T2* values over the entire placental volume was positively skewed in
254 RSOL (Z-score -4.69) and CoA- (Z-score -3.83), followed by our group of other major
255 lesions (Z-score -1.75) and HLHS (Z-score -1.12). Suspected VR (Z-score -0.8), CoA+
256 (Z-score -0.66) distributions were positively skewed within one SD. Our TGA cohort
257 was closest to expected skewness (Z-score 0.02).

258 We found positive kurtosis of distribution most significantly in RSOL with highest mean
259 deviation (Z-score -3.47), followed by CoA- (Z-score -2.86) and our group of other
260 major lesions (Z-score -1.97). HLHS and CoA+ showed positive kurtosis with almost
261 one SD from expected results (Z-Scores -0.98), while suspected VR (Z-score -0.73)
262 and TGA (Z-Score -0.27) showed similar kurtosis of distribution compared to our
263 control cohort.

264 All placentas (19/69, 28%) with individual z-scores $> \pm 3$ for any of calculated results
265 were individually reviewed for correlation with clinical confounders, while we found
266 MVM in two placentas and SGA complicating one pregnancy, overall numbers did not
267 reach statistical significance.

268 Mean and median deviation from GP (Z-score) for all cohorts and measured values are
269 listed in Table 2. Individual heterogeneity is demonstrated in histograms of occurrence
270 fraction of T2* values in all voxels within the individual placenta from all participants in
271 Figure 4. These histograms are color-coded by results of genetic testing in
272 Supplemental Figure 1.

273 Maternal BMI at scan did not have an effect on mean T2* values ($R^2 = 0.004$). Maternal
274 lie supine or left lateral, 30 and 7 scans respectively, had no significant effect on mean
275 T2* values in our control group ($p = 0.44$). Placental position dichotomised in mostly
276 anterior or posterior was also not associated with mean T2* values ($p = 0.98$). There
277 was no linear correlation with mean T2* Z-scores at scan and weight ($R^2 = 0.13$) or
278 head circumference at birth ($R^2 = 0.11$).

279 Discussion

280 In this study we have shown for the first time a comprehensive approach to placental
281 tissue characterisation in CHD. Using T2 relaxometry (T2*) and Gaussian Process
282 Regression (GPR) we were able to provide standard deviations for a range of placental
283 metrics across various CHD groups from predicted values derived by our model trained
284 on a control group. Mean T2* values over the entire placenta may not represent
285 regional differences adequately and can lead to misinterpretation of imaging findings.
286 Therefore, we also used histograms and evaluated skewness and kurtosis to show
287 specific pattern of distribution depending on CHD subtype, which in turn may represent
288 pathophysiological substrates.

289 Requiring minimal acquisition time (<1min) and minor modification to clinical scan
290 protocols, these baseline placental assessments have been included into all our clinical
291 fetal MRI scans.

292 While mean T2* deviation for RSOL (Z-score 2.30) is comparable with our group of
293 other cardiovascular lesions (Z-score 2.31), the latter shows much less deviation from
294 calculated normal distribution as described in skewness and kurtosis (Z-scores -1.75
295 and -1.97, respectively). Strikingly, right-sided obstructive lesions (RSOL) show highest
296 deviation from expected normal distribution represented in skewness (Z-score -4.69)
297 and kurtosis (Z-score -3.47).

298 Our findings of abnormal placental imaging appear CHD lesions specific. One might
299 speculate that common intrinsic developmental pathways of placenta and fetal heart
300 may play a larger role, warranting further investigations towards understanding the
301 pathophysiology. Future investigations in conjunction with assessments of fetal
302 circulation in the presence of CHD, a flourishing field in both ultrasound and MRI
303 research [33,34], may allow a more detailed interpretation of our results.

304 Given the severe implications after birth, the cohort of HLHS appears to have
305 surprisingly limited deviations from control in placental structure and function. This
306 suggests less co-dependence between the development and or antenatal effects of
307 this anomaly and the development and function of the placenta also requiring further
308 research.

309 Our findings are in keeping with data from a very large Danish cohort from Matthiesen
310 et al. also depicting an association of RSOL, but not left outflow tract obstructions
311 including HLHS, aortic valve stenosis, and coarctation of the aorta, with lower placental
312 weight [35]. Moreover Llorca et al. have suggested an imbalance in maternal and fetal

313 angiogenic factors may contribute to CHD and placental dysfunction most marked in
314 cohorts other than left sided lesions [36].

315 Fetuses with antenatal suspicion of CoA without confirmation after birth (CoA-) were
316 purposely not classified as controls in the design of this study. Firstly, presentation of
317 CoA may present several weeks (up to a year) after closure of the arterial duct [37].
318 None of the infants included here showed signs of coarctation at the time of manuscript
319 preparation. Secondly, there is a growing evidence that the observed ventricular
320 asymmetry is a feature of abnormal loading conditions leading to abnormal myocardial
321 deformation in fetuses with suspected coarctation [38]. In turn our CoA cohort spreads
322 relatively widely through the range of values provided by our placental assessments
323 (Supplemental Figure 2), which may reflect the wide clinical spectrum of CoA+
324 observed after birth, but also may indicate previously underexplored CHD entities
325 some of which do not require an extensive surgical arch repair within the first year of
326 life.

327 Studies of placental dysfunction and vascular malperfusion have shown altered
328 baseline conditions on assessment with T2 relaxometry accounting for a higher relative
329 response to maternal hyperoxygenation [15]. In our study we show similar altered
330 placental baseline conditions in case of fetal CHD. This is a critical insight to
331 understand the reported effects of short term maternal hyperoxygenation in CHD,
332 which are being explored to improve fetal oxygenation in CHD [39].

333 Altered placental baseline functions are of particular importance to understand the
334 complex intertwined fetomaternal environment in CHD and will also be essential to
335 understand further approaches on antenatal intervention such as maternal
336 hyperoxygenation [40].

337 Furthermore, structural assessments based on T2-weighted, and more recently also T1
338 imaging, depict changes over GA encompassing advanced lobulation with varied
339 lobule sizes, higher granularity, and substantial areas of low-signal intensity and an
340 increasing microstructural heterogeneity [41]. These changes are pronounced in FGR
341 or PE compared to healthy controls [22,42]. Although we did not report weighted MR
342 imaging in this study, observations from our T2* maps of increasing lacunarity over GA
343 is in keeping with the literature for our control and CHD cohort. Moreover, our CHD
344 cohorts also present higher lacunarity compared to healthy controls, most pronounced
345 in RSOL (Z-score -1.70).

346 As previously reported factors such as maternal age, BMI, fetal sex, parity, mode of
347 delivery and placental location were not correlated with T2* once corrected for GA [28].

348 **Limitations of this study**

349 Although we are based at a major referral centre for congenital heart disease in the
350 UK, this was a single centre study, with some patients delivering outside of our
351 hospital, having been referred for antenatal imaging. Our CHD groups are of moderate
352 sample size and may influence statistical significance.

353 Our placental scan protocol consistent of one single timepoint, therefore temporal
354 variance in placental T2* measurements, as recently suggested by other groups, might
355 not be accounted for in our data [43]. Recent literature describes also the possible
356 influence of maternal or placental position on T2* signals in higher field MRI [44]. While
357 we have scanned all our CHD patients in left lateral position, controls were also
358 scanned supine, resulting in higher statistical variance, 371.6 vs 776.5 respectively, but
359 not reaching statistical significance.

360 While in-utero MRI provides an excellent opportunity to provide early, in-vivo evidence
361 for specific placental phenotypes associated with CHD and hence a window of
362 observation into the intertwined relation between developing heart and placenta, it
363 cannot directly answer the question on causality and order of events.

364 Despite recent attempts to standardize definitions for FGR/IUGR, there is currently no
365 universal definition, which would include fetuses with congenital malformations such as
366 CHD [45]. Furthermore, the recent consensus-based definition of growth restriction on
367 the newborn excludes congenital and chromosomal abnormalities specifically, although
368 stating it may be applicable for this group due to the lack of any other option [32].

369 Previous literature, including a large population of 924,422 cohorts with over 5500 CHD
370 cases from Denmark also suggests an association of some CHD subtypes with lower
371 birth weight and head circumference [46]. Therefore, we did not exclude any
372 participants in our CHD cohort solely due diagnosis for FGR/IUGR or SGA.

373 Placental histopathology results were only available in 25% of all our assessments and
374 in CHD patients only with a specific clinical question at time of birth, such as maternal
375 pyrexia. Statistical analysis of features of MVM or fetal vascular malperfusion (FVM) or
376 Chorioamnionitis could therefore not be included. Similarly, only two placentas showed
377 infarcts in histopathology. These could be identified on the in-vivo imaging but did not
378 significantly alter the observed whole placental mean T2* results and have therefore
379 not been factored in the statistical analysis. Previous literature from postnatal placental
380 histopathology describe increased findings of FVM and MVM in the presence of
381 preeclampsia as well as CHD supporting the hypothesis of similar etiopathogenetic
382 factors contributing to the development of placental malformation and CHD [6].

383 While T2* values are an indicator of oxygen concentration, direct measurement was
384 not possible in our setting. Oxygen-haemoglobin dissociation curves derived from MRI
385 in animal models might be available for future studies to allow close estimation of
386 actual oxygen content in the fetoplacental circulation [47,48].
387 Including only five pregnancies with confirmed fetal chromosomal abnormalities in our
388 cohort did not allow for further statistical analysis of potential associations. This study
389 did not include volumetric assessments of the placenta given echo planar imaging is
390 associated with geometric distortions.
391 Future studies applied to CHD cohorts with more complex and time-consuming scan
392 protocols, including diffusion-weighted imaging or texture analysis using T1 and T2
393 weighted methods, may allow modelling of fetal and maternal peculiarities in circulation
394 and disentangle tissue characteristics and flow pathophysiology, as recently in CHD
395 cohorts [41,49–51]. Slator et al. have recently published a study with a focused
396 extensive research protocol on the microenvironments within the placenta depicting
397 heterogenous compartments from maternal and fetal side [52]. Using Velocity-
398 Selective Arterial Spin Labelled MRI Zun et al. also reported that global placental
399 perfusion significantly decreased and regional variation of placental perfusion
400 significantly increased over GA in fetuses with CHD [53,54].

401

402 **Conclusion**

403 This study describes in vivo differences in placental tissue phenotypes in healthy
404 controls and fetuses with antenatal diagnosis of congenital heart disease based on T2
405 relaxometry. Using machine learning we depict unique features of T2* value
406 distribution and their standard score from our normal cohort for a wide range of major
407 cardiac lesions, providing information on placental dysfunction complementary in the
408 antenatal assessment of CHD.

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420 not necessarily those of the NHS, the NIHR or the Department of Health.

421 **Declaration of competing interests**

422 The authors declare that they have no competing interest.

423 **Keywords**

424 Congenital Heart disease (CHD), Placenta, Magnetic resonance imaging (MRI), T2*
425 Mapping, Gaussian Process Regression (GPR), Bayesian non-parametric regression,
426 Machine learning

427 **References**

- 428 [1] T. van der Bom, A.C. Zomer, A.H. Zwinderman, F.J. Meijboom, B.J. Bouma, B.J.M.
429 Mulder, The changing epidemiology of congenital heart disease, *Nature Reviews*
430 *Cardiology*. 8 (2011) 50–60. <https://doi.org/10.1038/nrcardio.2010.166>.
- 431 [2] M.S. Zimmerman, A.G.C. Smith, C.A. Sable, M.M. Echko, L.B. Wilner, H.E. Olsen,
432 H.T. Atalay, A. Awasthi, Z.A. Bhutta, J.L. Boucher, F. Castro, P.A. Cortesi, M.
433 Dubey, F. Fischer, S. Hamidi, S.I. Hay, C.L. Hoang, C. Hugo-Hamman, K.J.
434 Jenkins, A. Kar, I.A. Khalil, R.K. Kumar, G.F. Kwan, D.T. Mengistu, A.H. Mokdad,
435 M. Naghavi, L. Negesa, I. Negoi, R.I. Negoi, C.T. Nguyen, H.L.T. Nguyen, L.H.
436 Nguyen, S.H. Nguyen, T.H. Nguyen, M.R. Nixon, J.J. Noubiap, S. Patel, E.K.
437 Peprah, R.C. Reiner, G.A. Roth, M.-H. Temsah, M.R. Tovani-Palone, J.A. Towbin,
438 B.X. Tran, T.T. Tran, N.T. Truong, T. Vos, K. Vosoughi, R.G. Weintraub, K.G.
439 Weldegewergs, Z. Zaidi, B. Zheleva, L. Zuhlke, C.J.L. Murray, G.R. Martin, N.J.
440 Kassebaum, Global, regional, and national burden of congenital heart disease,
441 1990–2017: a systematic analysis for the Global Burden of Disease Study 2017,
442 *The Lancet Child & Adolescent Health*. 4 (2020) 185–200.
443 [https://doi.org/10.1016/S2352-4642\(19\)30402-X](https://doi.org/10.1016/S2352-4642(19)30402-X).
- 444 [3] D.K. Jarrell, M.L. Lennon, J.G. Jacot, Epigenetics and Mechanobiology in Heart
445 Development and Congenital Heart Disease, *Diseases*. 7 (2019) 52.
446 <https://doi.org/10.3390/diseases7030052>.
- 447 [4] Russell Mark W., Chung Wendy K., Kaltman Jonathan R., Miller Thomas A.,
448 *Advances in the Understanding of the Genetic Determinants of Congenital Heart*
449 *Disease and Their Impact on Clinical Outcomes*, *Journal of the American Heart*
450 *Association*. 7 (2018) e006906. <https://doi.org/10.1161/JAHA.117.006906>.
- 451 [5] S. Zaidi, M. Choi, H. Wakimoto, L. Ma, J. Jiang, J.D. Overton, A. Romano-
452 Adesman, R.D. Bjornson, R.E. Breitbart, K.K. Brown, N.J. Carriero, Y.H. Cheung,
453 J. Deanfield, S. DePalma, K.A. Fakhro, J. Glessner, H. Hakonarson, M.J. Italia,
454 J.R. Kaltman, J. Kaski, R. Kim, J.K. Kline, T. Lee, J. Leipzig, A. Lopez, S.M.
455 Mane, L.E. Mitchell, J.W. Newburger, M. Parfenov, I. Pe'er, G. Porter, A.E.
456 Roberts, R. Sachidanandam, S.J. Sanders, H.S. Seiden, M.W. State, S.
457 Subramanian, I.R. Tikhonova, W. Wang, D. Warburton, P.S. White, I.A. Williams,
458 H. Zhao, J.G. Seidman, M. Brueckner, W.K. Chung, B.D. Gelb, E. Goldmuntz,
459 C.E. Seidman, R.P. Lifton, De novo mutations in histone-modifying genes in
460 congenital heart disease, *Nature*. 498 (2013) 220–223.
461 <https://doi.org/10.1038/nature12141>.

- 462 [6] H. Miremberg, L. Gindes, L. Schreiber, A.R. Sternfeld, J. Bar, M. Kovo, The
463 association between severe fetal congenital heart defects and placental vascular
464 malperfusion lesions, *Prenatal Diagnosis*. 39 (2019) 962–967.
465 <https://doi.org/10.1002/pd.5515>.
- 466 [7] J. Binder, S. Carta, J.S. Carvalho, E. Kalafat, A. Khalil, B. Thilaganathan, Evidence
467 for uteroplacental malperfusion in fetuses with major congenital heart defects,
468 *PLOS ONE*. 15 (2020) e0226741. <https://doi.org/10.1371/journal.pone.0226741>.
- 469 [8] S.D. Schlatterer, J. Murnick, M. Jacobs, L. White, M.T. Donofrio, C. Limperopoulos,
470 Placental Pathology and Neuroimaging Correlates in Neonates with Congenital
471 Heart Disease, *Sci Rep*. 9 (2019). <https://doi.org/10.1038/s41598-019-40894-y>.
- 472 [9] G.J. Burton, E. Jauniaux, Development of the Human Placenta and Fetal Heart:
473 Synergic or Independent?, *Front. Physiol*. 9 (2018).
474 <https://doi.org/10.3389/fphys.2018.00373>.
- 475 [10] J.A. Courtney, J.F. Cnota, H.N. Jones, The Role of Abnormal Placentation in
476 Congenital Heart Disease; Cause, Correlate, or Consequence?, *Frontiers in*
477 *Physiology*. 9 (2018) 1045. <https://doi.org/10.3389/fphys.2018.01045>.
- 478 [11] L. Pauling, C.D. Coryell, The Magnetic Properties and Structure of Hemoglobin,
479 Oxyhemoglobin and Carbonmonoxyhemoglobin, *Proceedings of the National*
480 *Academy of Sciences*. 22 (1936) 210–216. <https://doi.org/10.1073/pnas.22.4.210>.
- 481 [12] S. Ogawa, T.-M. Lee, A.S. Nayak, P. Glynn, Oxygenation-sensitive contrast in
482 magnetic resonance image of rodent brain at high magnetic fields, *Magnetic*
483 *Resonance in Medicine*. 14 (1990) 68–78.
484 <https://doi.org/10.1002/mrm.1910140108>.
- 485 [13] J. Hutter, P.J. Slator, L. Jackson, A.D.S. Gomes, A. Ho, L. Story, J.
486 O’Muircheartaigh, R.P.A.G. Teixeira, L.C. Chappell, D.C. Alexander, M.A.
487 Rutherford, J. V. Hajnal, Multi-modal functional MRI to explore placental function
488 over gestation., *Magnetic Resonance in Medicine*. 81 (2019) 1191–1204.
489 <https://doi.org/10.1002/mrm.27447>.
- 490 [14] M.C. Schabel, V.H.J. Roberts, J.O. Lo, S. Platt, K.A. Grant, A.E. Frias, C.D.
491 Kroenke, Functional imaging of the nonhuman primate Placenta with endogenous
492 blood oxygen level–dependent contrast, *Magnetic Resonance in Medicine*. 76
493 (2016) 1551–1562. <https://doi.org/10.1002/mrm.26052>.
- 494 [15] M. Sinding, D.A. Peters, S.S. Poulsen, J.B. Frøkjær, O.B. Christiansen, A.
495 Petersen, N. Uldbjerg, A. Sørensen, Placental baseline conditions modulate the

- 496 hyperoxic BOLD-MRI response., *Placenta*. 61 (2018) 17–23.
497 <https://doi.org/10.1016/j.placenta.2017.11.002>.
- 498 [16]C. Wright, D.M. Morris, P.N. Baker, I.P. Crocker, P.A. Gowland, G.J. Parker, C.P.
499 Sibley, Magnetic resonance imaging relaxation time measurements of the
500 placenta at 1.5T, *Placenta*. 32 (2011) 1010–1015.
501 <https://doi.org/10.1016/j.placenta.2011.07.008>.
- 502 [17]I. Huen, D.M. Morris, C. Wright, G.J.M. Parker, C.P. Sibley, E.D. Johnstone, J.H.
503 Naish, R 1 and R 2 * changes in the human placenta in response to maternal
504 oxygen challenge, *Magnetic Resonance in Medicine*. 70 (2013) 1427–1433.
505 <https://doi.org/10.1002/mrm.24581>.
- 506 [18]A. Sørensen, D. Peters, C. Simonsen, M. Pedersen, B. Stausbøl-Grøn, O.B.
507 Christiansen, G. Lingman, N. Uldbjerg, Changes in human fetal oxygenation
508 during maternal hyperoxia as estimated by BOLD MRI., *Prenatal Diagnosis*. 33
509 (2013) 141–5. <https://doi.org/10.1002/pd.4025>.
- 510 [19]E. Ingram, D. Morris, J. Naish, J. Myers, E. Johnstone, MR imaging measurements
511 of altered placental oxygenation in pregnancies complicated by fetal growth
512 restriction, *Radiology*. 285 (2017) 953–960.
513 <https://doi.org/10.1148/radiol.2017162385>.
- 514 [20]G.E. Chalouhi, M. Alison, B. Deloison, R. Thiam, G. Autret, D. Balvay, C.A.
515 Cuenod, O. Clément, L.J. Salomon, N. Siauve, Fetoplacental Oxygenation in an
516 Intrauterine Growth Restriction Rat Model by Using Blood Oxygen Level–
517 Dependent MR Imaging at 4.7 T, *Radiology*. 269 (2013) 122–129.
518 <https://doi.org/10.1148/radiol.13121742>.
- 519 [21]E. Girsh, V. Plaks, A.A. Gilad, N. Nevo, E. Schechtman, M. Neeman, N. Dekel,
520 Cloprostenol, a Prostaglandin F(2alpha) Analog, Induces Hypoxia in Rat
521 Placenta: BOLD Contrast MRI, *NMR in Biomedicine*. 20 (2007) 304–325.
522 <https://doi.org/10.1002/nbm>.
- 523 [22]A.E.P. Ho, J. Hutter, L.H. Jackson, P.T. Seed, L. McCabe, M. Al-adnani, A.
524 Marnierides, S. George, L. Story, J. V Hajnal, M.A. Rutherford, L.C. Chappell, T2 *
525 Placental Magnetic Resonance Imaging An Observational Cohort Study,
526 Hypertension (Dallas, Tex. : 1979). (2020) 1–9.
527 <https://doi.org/10.1161/HYPERTENSIONAHA.120.14701>.
- 528 [23]A.J. Hirsch, V.H.J. Roberts, P.L. Grigsby, N. Haese, M.C. Schabel, X. Wang, J.O.
529 Lo, Z. Liu, C.D. Kroenke, J.L. Smith, M. Kelleher, R. Broeckel, C.N. Kreklywich,
530 C.J. Parkins, M. Denton, P. Smith, V. DeFilippis, W. Messer, J.A. Nelson, J.D.

531 Hennebold, M. Grafe, L. Colgin, A. Lewis, R. Ducore, T. Swanson, A.W. Legasse,
532 M.K. Axthelm, R. MacAllister, A.V. Moses, T.K. Morgan, A.E. Frias, D.N.
533 Streblow, Zika virus infection in pregnant rhesus macaques causes placental
534 dysfunction and immunopathology, *Nature Communications*. 9 (2018) 263.
535 <https://doi.org/10.1038/s41467-017-02499-9>.

536 [24]J.O. Lo, V.H.J. Roberts, M.C. Schabel, X. Wang, T.K. Morgan, Z. Liu, C.
537 Studholme, C.D. Kroenke, A.E. Frias, Novel Detection of Placental Insufficiency
538 by Magnetic Resonance Imaging in the Nonhuman Primate:, *Reproductive*
539 *Sciences*. (2017). <https://doi.org/10.1177/1933719117699704>.

540 [25]J. Villar, L. Cheikh Ismail, C.G. Victora, E.O. Ohuma, E. Bertino, D.G. Altman, A.
541 Lambert, A.T. Papageorgiou, M. Carvalho, Y.A. Jaffer, M.G. Gravett, M. Purwar,
542 I.O. Frederick, A.J. Noble, R. Pang, F.C. Barros, C. Chumlea, Z.A. Bhutta, S.H.
543 Kennedy, International Fetal and Newborn Growth Consortium for the 21st
544 Century (INTERGROWTH-21st), International standards for newborn weight,
545 length, and head circumference by gestational age and sex: the Newborn Cross-
546 Sectional Study of the INTERGROWTH-21st Project., *Lancet (London, England)*.
547 384 (2014) 857–68. [https://doi.org/10.1016/S0140-6736\(14\)60932-6](https://doi.org/10.1016/S0140-6736(14)60932-6).

548 [26]P.A. Yushkevich, J. Piven, H.C. Hazlett, R.G. Smith, S. Ho, J.C. Gee, G. Gerig,
549 User-guided 3D active contour segmentation of anatomical structures:
550 significantly improved efficiency and reliability., *NeuroImage*. 31 (2006) 1116–28.
551 <https://doi.org/10.1016/j.neuroimage.2006.01.015>.

552 [27]K.H. Zou, S.K. Warfield, A. Bharatha, C.M.C. Tempany, M.R. Kaus, S.J. Haker,
553 W.M. Wells, F.A. Jolesz, R. Kikinis, Statistical Validation of Image Segmentation
554 Quality Based on a Spatial Overlap Index, *Acad Radiol*. 11 (2004) 178–189.
555 [https://doi.org/10.1016/S1076-6332\(03\)00671-8](https://doi.org/10.1016/S1076-6332(03)00671-8).

556 [28]J. Hutter, L. Jackson, A. Ho, M. Pietsch, L. Story, L.C. Chappell, J. V. Hajnal, M.
557 Rutherford, T2* relaxometry to characterize normal placental development over
558 gestation in-vivo at 3T, *Wellcome Open Research*. 4 (2019) 166.
559 <https://doi.org/10.12688/wellcomeopenres.15451.1>.

560 [29]A.R. Backes, A new approach to estimate lacunarity of texture images, *Pattern*
561 *Recognition Letters*. 34 (2013) 1455–1461.
562 <https://doi.org/10.1016/j.patrec.2013.05.008>.

563 [30]F. Pedregosa, G. Varoquaux, A. Gramfort, V. Michel, B. Thirion, O. Grisel, M.
564 Blondel, A. Müller, J. Nothman, G. Louppe, P. Prettenhofer, R. Weiss, V.
565 Dubourg, J. Vanderplas, A. Passos, D. Cournapeau, M. Brucher, M. Perrot, É.

566 Duchesnay, Scikit-learn: Machine Learning in Python, *Journal of Machine*
567 *Learning Research*. 12 (2012) 2825–2830.

568 [31]A.F. Marquand, I. Rezek, J. Buitelaar, C.F. Beckmann, Understanding
569 Heterogeneity in Clinical Cohorts Using Normative Models: Beyond Case-Control
570 Studies, *Biological Psychiatry*. 80 (2016) 552–561.
571 <https://doi.org/10.1016/j.biopsych.2015.12.023>.

572 [32]I.M. Beune, F.H. Bloomfield, W. Ganzevoort, N.D. Embleton, P.J. Rozance, A.G.
573 van Wassenaer-Leemhuis, K. Wynia, S.J. Gordijn, Consensus Based Definition of
574 Growth Restriction in the Newborn, *J Pediatr*. 196 (2018) 71-76.e1.
575 <https://doi.org/10.1016/j.jpeds.2017.12.059>.

576 [33]G. Mielke, N. Benda, Cardiac output and central distribution of blood flow in the
577 human fetus., *Circulation*. 103 (2001) 1662–8.
578 <https://doi.org/10.1161/01.cir.103.12.1662>.

579 [34]M. Seed, J. F P van Amerom, S.-J. Yoo, B. Nafisi, L. Grosse-Wortmann, E. Jaeggi,
580 M.S. Jansz, C.K. Macgowan, Feasibility of quantification of the distribution of
581 blood flow in the normal human fetal circulation using CMR: a cross-sectional
582 study, *Journal of Cardiovascular Magnetic Resonance*. 14 (2012) 79.
583 <https://doi.org/10.1186/1532-429X-14-79>.

584 [35]N.B. Matthiesen, T.B. Henriksen, P. Agergaard, J.W. Gaynor, C.C. Bach, V.E.
585 Hjortdal, J.R. Østergaard, Congenital Heart Defects and Indices of Placental and
586 Fetal Growth in a Nationwide Study of 924 422 Liveborn Infants, *Circulation*. 134
587 (2016) 1546–1556. <https://doi.org/10.1161/CIRCULATIONAHA.116.021793>.

588 [36]E. Llurba, O. Sánchez, Q. Ferrer, K.H. Nicolaides, A. Ruíz, C. Domínguez, J.
589 Sánchez-de-Toledo, B. García-García, G. Soro, S. Arévalo, M. Goya, A. Suy, S.
590 Pérez-Hoyos, J. Alijotas-Reig, E. Carreras, L. Cabero, Maternal and foetal
591 angiogenic imbalance in congenital heart defects, *European Heart Journal*. 35
592 (2014) 701–707. <https://doi.org/10.1093/eurheartj/eh389>.

593 [37]C.E.G. Head, V.C. Jowett, G.K. Sharland, J.M. Simpson, Timing of presentation
594 and postnatal outcome of infants suspected of having coarctation of the aorta
595 during fetal life, *Heart*. 91 (2005) 1070–1074.
596 <https://doi.org/10.1136/hrt.2003.033027>.

597 [38]J.O. Miranda, L. Hunter, S. Tibby, G. Sharland, O. Miller, J.M. Simpson, Myocardial
598 deformation in fetuses with coarctation of the aorta: a case-control study,
599 *Ultrasound Obstet Gynecol*. 49 (2017) 623–629.
600 <https://doi.org/10.1002/uog.15939>.

- 601 [39]W. You, N.N. Andescavage, K. Kapse, M.T. Donofrio, M. Jacobs, C.
602 Limperopoulos, Hemodynamic Responses of the Placenta and Brain to Maternal
603 Hyperoxia in Fetuses with Congenital Heart Disease by Using Blood Oxygen-
604 Level Dependent MRI., *Radiology*. (2019) 190751.
605 <https://doi.org/10.1148/radiol.2019190751>.
- 606 [40]J. Co-Vu, D. Lopez-Colon, H.V. Vyas, N. Weiner, C. DeGross, Maternal
607 hyperoxygenation: A potential therapy for congenital heart disease in the fetuses?
608 A systematic review of the current literature, *Echocardiography*. 34 (2017) 1822–
609 1833. <https://doi.org/10.1111/echo.13722>.
- 610 [41]Q.N. Do, M.A. Lewis, A.J. Madhuranthakam, Y. Xi, A.A. Bailey, R.E. Lenkinski,
611 D.M. Twickler, Texture analysis of magnetic resonance images of the human
612 placenta throughout gestation: A feasibility study, *PLOS ONE*. 14 (2019)
613 e0211060. <https://doi.org/10.1371/journal.pone.0211060>.
- 614 [42]N. Andescavage, S. Dahdouh, M. Jacobs, S. Yewale, D. Bulas, S. Iqbal, A.
615 Baschat, C. Limperopoulos, In vivo textural and morphometric analysis of
616 placental development in healthy & growth-restricted pregnancies using magnetic
617 resonance imaging, *Pediatric Research*. 85 (2019) 974–981.
618 <https://doi.org/10.1038/s41390-019-0311-1>.
- 619 [43]A.M. El-Ali, W. Reynolds, V. Rajagopalan, V. Lee, J. Wallace, J. Schabdach, A.
620 Zehner, M. Gruss, J.H. Adibi, V. Schmithorst, A. Panigrahy, BOLD MR Imaging of
621 Placenta in Congenital Heart Disease: Correlation with Maternal Risk Factors,
622 Placental Pathology, and Maternal Serum Hormones [abstract], ISMRM 27th
623 Annual Meeting & Exhibition. (2019). <https://doi.org/10.1101/388074>.
- 624 [44]E.A. Turk, S.M. Abulnaga, J. Luo, J.N. Stout, H.A. Feldman, A. Turk, B. Gagoski,
625 L.L. Wald, E. Adalsteinsson, D.J. Roberts, C. Bibbo, J.N. Robinson, P. Golland,
626 P.E. Grant, W.H. Barth, Placental MRI: Effect of maternal position and uterine
627 contractions on placental BOLD MRI measurements, *Placenta*. (2020).
628 <https://doi.org/10.1016/j.placenta.2020.04.008>.
- 629 [45]S.J. Gordijn, I.M. Beune, B. Thilaganathan, A. Papageorgiou, A.A. Baschat, P.N.
630 Baker, R.M. Silver, K. Wynia, W. Ganzevoort, Consensus definition of fetal growth
631 restriction: a Delphi procedure, *Ultrasound in Obstetrics & Gynecology*. 48 (2016)
632 333–339. <https://doi.org/10.1002/uog.15884>.
- 633 [46]N.B. Matthiesen, T.B. Henriksen, J.W. Gaynor, P. Agergaard, C.C. Bach, V.E.
634 Hjortdal, J.R. Østergaard, Congenital Heart Defects and Indices of Fetal Cerebral

635 Growth in a Nationwide Cohort of 924 422 Liveborn Infants., *Circulation*. 133
636 (2016) 566–75. <https://doi.org/10.1161/CIRCULATIONAHA.115.019089>.

637 [47]R. Avni, O. Golani, A. Akselrod-Ballin, Y. Cohen, I. Biton, J.R. Garbow, M.
638 Neeman, MR Imaging–derived Oxygen-Hemoglobin Dissociation Curves and
639 Fetal-Placental Oxygen-Hemoglobin Affinities, *Radiology*. 280 (2016) 68–77.
640 <https://doi.org/10.1148/radiol.2015150721>.

641 [48]L. Sun, D. Marini, B. Saini, E. Schrauben, C.K. Macgowan, M. Seed,
642 Understanding Fetal Hemodynamics Using Cardiovascular Magnetic Resonance
643 Imaging, *FDT*. 47 (2020) 354–362. <https://doi.org/10.1159/000505091>.

644 [49]J. Hutter, A.A. Hartevelde, L.H. Jackson, S. Franklin, C. Bos, M.J.P. Osch, J.
645 O’Muircheartaigh, A. Ho, L. Chappell, J. V. Hajnal, M. Rutherford, E. De Vita,
646 Perfusion and apparent oxygenation in the human placenta (PERFOX), *Magnetic
647 Resonance in Medicine*. 83 (2020) 549–560. <https://doi.org/10.1002/mrm.27950>.

648 [50]A. Melbourne, R. Aughwane, M. Sokolska, D. Owen, G. Kendall, D. Flouri, A.
649 Bainbridge, D. Atkinson, J. Deprest, T. Vercauteren, A. David, S. Ourselin,
650 Separating fetal and maternal placenta circulations using multiparametric MRI,
651 *Magnetic Resonance in Medicine*. 81 (2019) 350–361.
652 <https://doi.org/10.1002/mrm.27406>.

653 [51]N.S. Dellschaft, G. Hutchinson, S. Shah, N.W. Jones, C. Bradley, L. Leach, C.
654 Platt, R. Bowtell, P.A. Gowland, The haemodynamics of the human placenta in
655 utero, *PLOS Biology*. 18 (2020) e3000676.
656 <https://doi.org/10.1371/journal.pbio.3000676>.

657 [52]P.J. Slator, J. Hutter, L. McCabe, A.D.S. Gomes, A.N. Price, E. Panagiotaki, M.A.
658 Rutherford, J.V. Hajnal, D.C. Alexander, Placenta microstructure and
659 microcirculation imaging with diffusion MRI, *Magnetic Resonance in Medicine*. 80
660 (2018) 756–766. <https://doi.org/10.1002/mrm.27036>.

661 [53]Z. Zun, G. Zaharchuk, N.N. Andescavage, M.T. Donofrio, C. Limperopoulos, Non-
662 Invasive Placental Perfusion Imaging in Pregnancies Complicated by Fetal Heart
663 Disease Using Velocity-Selective Arterial Spin Labeled MRI., *Scientific Reports*. 7
664 (2017) 16126. <https://doi.org/10.1038/s41598-017-16461-8>.

665 [54]Z. Zun, C. Limperopoulos, Placental perfusion imaging using velocity-selective
666 arterial spin labeling, *Magn Reson Med*. 80 (2018) 1036–1047.
667 <https://doi.org/10.1002/mrm.27100>.

668

	Median	IQR	Min	Max	p
Control cohort (n=37)					
Maternal age at scan (a)	34.65	4.65	27.69	44.03	
Maternal BMI at scan (kg/m ²)	30.03	8.67	18.75	42.59	
Gestational age at scan (w)	30.86	8.36	19.57	37.86	
Fetal sex (female/total)	0.62				
Gestational age at birth (w)	40.14	2.14	38.14	42.14	
Birth weight (g)	3583	534	2780	4450	
BW Z-score	0.9558	1.2147	-1.0550	2.1966	
BW Centile	83.04	39.16	14.57	98.60	
CHD cohort (n=69)					
Maternal age at scan (a)	32.63	8.39	18.60	40.79	0.002†
Maternal BMI at scan (kg/m ²)	28.39	5.63	18.81	40.35	0.094†
Gestational age at scan (w)	31.57	2.64	28.43	36.29	0.069†
Fetal sex (female/total)	0.58				0.836
Gestational age at birth (w)	38.71	1.57	32.71	41.71	<0.001
Birth weight (g)	3080	964	1610	5150	<0.001
BW Z-score	-0.1230	1.7762	-2.9547	3.7986	0.003
BW Centile	48.20	62.20	0.16	99.99	0.003
Diagnosis	N	%			
Left-sided obstructive lesions	10	14.5	HLHS		
	13	18.8	COA- (suspected, not confirmed)		
	8	11.6	COA+ (confirmed neonatal)		
Right-sided obstructive lesions	10	14.5	TOF, PA, PS, TA, Ebstein anomaly		
Disorders of mixing	5	7.2	TGA		
Suspected vascular rings	17	24.6	DAA, RAA (+/- ALSA)		
Other major lesions	6	8.7	CAT, PAPVD, CR		

670 **Table 1: Maternal and pregnancy characteristics.**

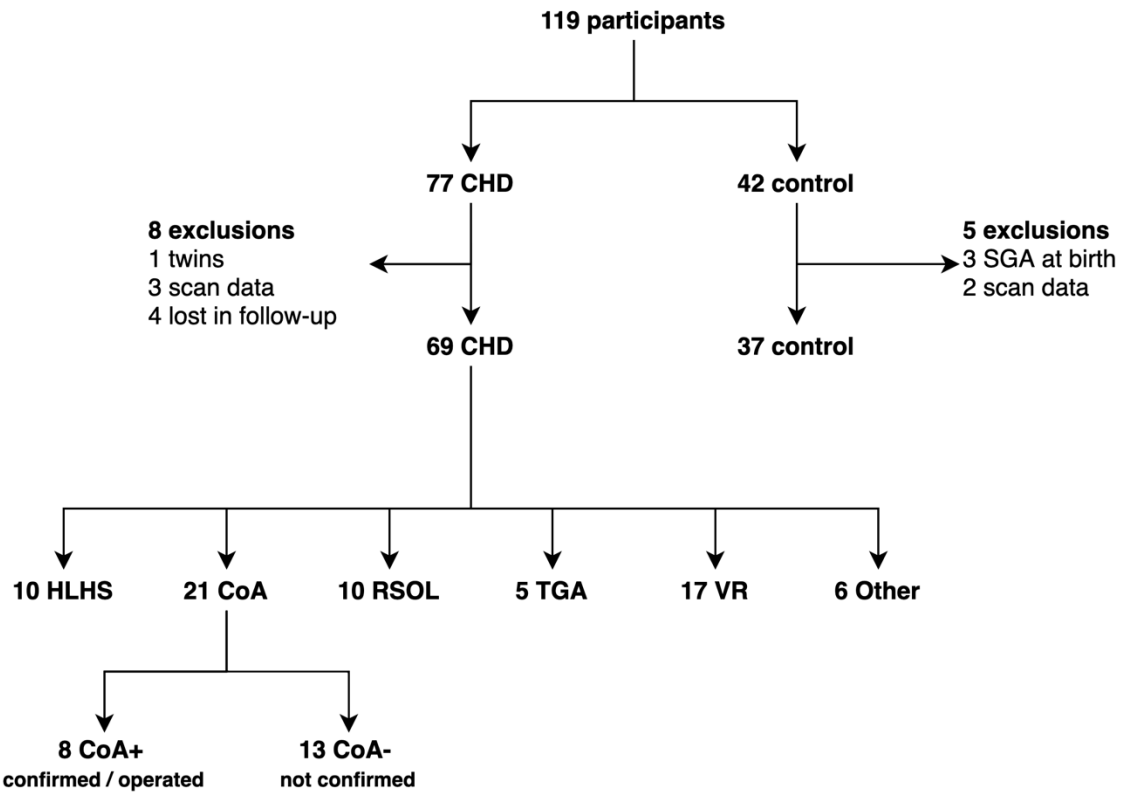
671 (†) For statistical analysis controls were restrained to GA range at scan (28 – 37

672 weeks, N=23).

Mean deviation from GP [Z-score] +/- SD of Z-scores within group									
	Control	CHD	HLHS	COA-	COA+	RSOL	TGA	Other	VR
N	9	69	10	13	8	10	5	6	17
Mean T2*	-0.56 +/-1.34	0.93* +/-2.26	0.64 +/-2.03	1.39* +/-2.91	0.24 +/-1.34	2.30*** +/-2.07	-0.11 +/-1.86	2.31** +/-2.01	0.09 +/-2.06
Skewness	-0.08 +/-0.59	-1.99* +/-4.27	-1.12 +/-1.58	-3.83 +/-7.18	-0.66 +/-0.73	-4.69* +/-5.56	0.02 +/-0.73	-1.75** +/-1.25	-0.80 +/-2.70
Kurtosis	-0.53 +/-1.01	-1.67 +/-2.34	-0.98 +/-1.39	-2.86* +/-3.25	-0.98 +/-0.67	-3.47** +/-3.02	-0.27 +/-1.50	-1.97** +/-0.93	-0.73 +/-1.66
Lacunarity	-0.15 +/-1.27	-0.54 +/-1.55	-0.18 +/-1.21	-1.02 +/-1.69	0.26 +/-0.53	-1.70* +/-1.99	-0.01 +/-0.98	-1.26 +/-2.35	0.06 +/-0.93
Median deviation from GP [Z-score] (IQR within group)									
	Control	CHD	HLHS	COA-	COA+	RSOL	TGA	Other	VR
N	9	69	10	13	8	10	5	6	17
Mean T2*	-0.78 (2.05)	0.70* (2.93)	0.57 (2.37)	0.89* (4.05)	0.25 (2.06)	2.36*** (3.32)	-0.41 (3.68)	2.10** (3.77)	-0.59 (2.76)
Skewness	0.01 (0.71)	-0.47* (2.19)	-0.49 (2.94)	-0.96 (4.38)	-0.62 (0.98)	-2.12* (10.23)	0.73 (2.32)	-1.89** (2.40)	0.02 (0.83)
Kurtosis	-0.80 (1.67)	-1.10 (1.95)	-0.75 (2.07)	-1.72* (3.35)	-0.86 (1.04)	-2.15** (5.64)	0.41 (2.65)	-1.83** (1.83)	-0.63 (1.23)
Lacunarity	0.01 (1.95)	-0.18 (1.67)	-0.09 (1.28)	-0.68 (2.49)	0.17 (0.82)	-1.43* (2.22)	0.29 (1.84)	-0.37 (3.84)	0.05 (1.35)

673 **Table 2: Results Gaussian process Regression, groupwise.**

674 Entire CHD cohort (CHD), Hypoplastic left heart syndrome (HLHS), Coarctation
675 suspected not confirmed (CoA-), Coarctation confirmed operated (CoA+), Right-sided
676 obstructive lesions (RSOL), Disorders of mixing (TGA), Other lesions, Suspected
677 vascular rings (VR). Significance calculated by Independent t-test (Mean T2*) or Mann-
678 Whitney U (Skewness, Kurtosis, Lacunarity) in comparison to test control cohort, *
679 $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$.



680

681

Figure 1 - Study cohort.

682

CHD subtypes classified by main antenatal diagnosis: Hypoplastic left heart (HLHS),
 683 coarctation of the aorta (CoA), right-sided obstructive lesions (RSOL), transposition of
 684 the great arteries (TGA), suspected vascular ring (VR) and other complex lesions.

685

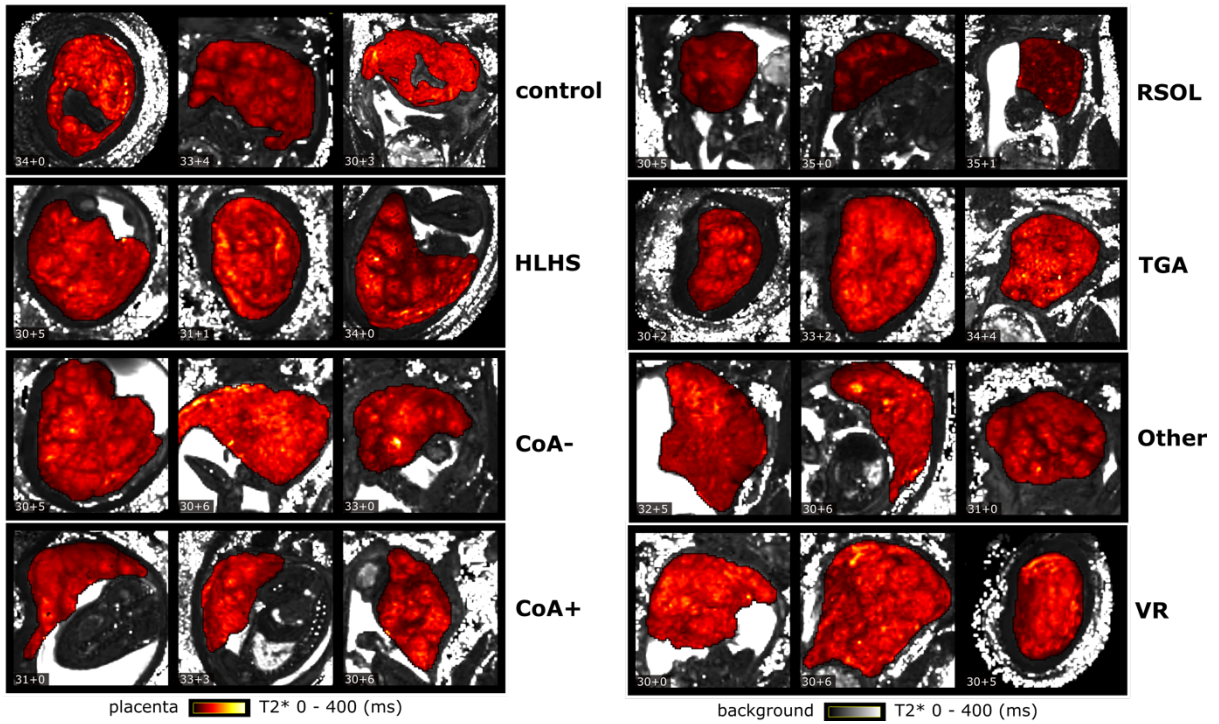
Controls were excluded for low birth weight (SGA) and insufficient scan data,

686

pregnancies complicated by fetal CHD had to be excluded for twin pregnancy and loss

687

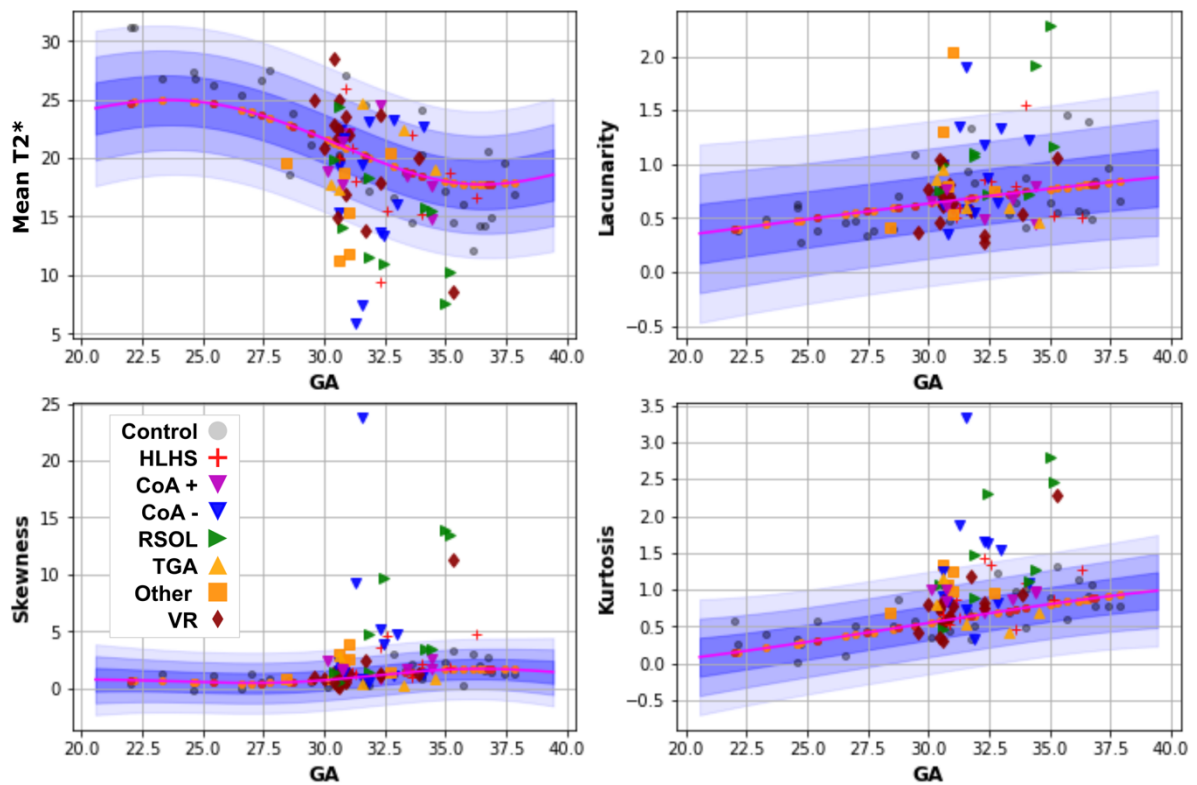
in follow-up at time of analysis.



688

689 **Figure 2: Qualitative Imaging**

690 Illustration of mid-stack slice in coronal orientation from T2* maps from 3 individual
 691 placentas for all included cohorts. Segmented placental parenchyma is highlighted in
 692 red/yellow scales.

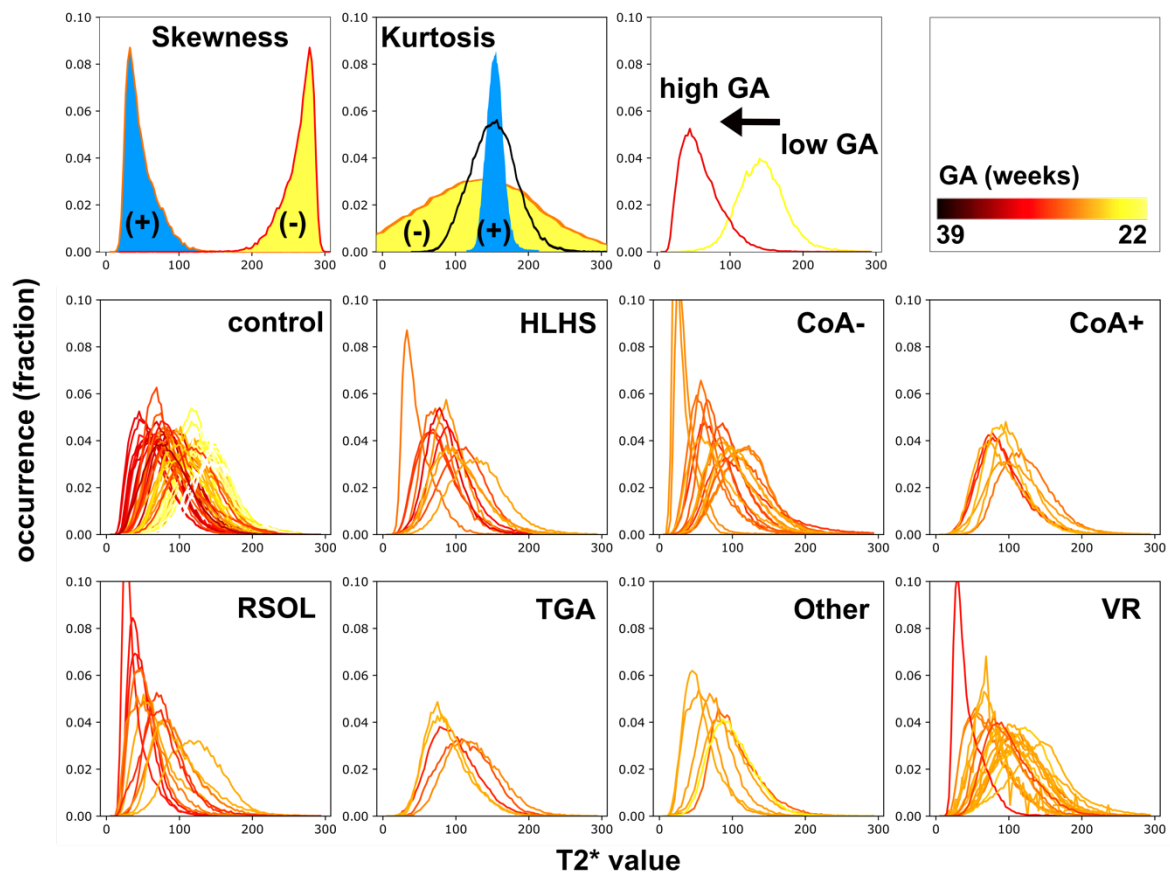


693

694 **Figure 3 – Gaussian Process Regression results**

695 The posterior probability for Mean T2*, Lacunarity, Skewness and Kurtosis over GA for
 696 all considered controls in red, the corridors corresponding to Z-scores of 1, 2 and 3 are
 697 illustrated in blue. Cohorts: Control (test subset), Hypoplastic left heart syndrome
 698 (HLHS), Coarctation of the aorta confirmed (COA+) and not confirmed (COA-), right-
 699 sided obstructive lesions (RSOL), disorders of mixing (TGA), other major lesions and
 700 suspected vascular rings (VR).

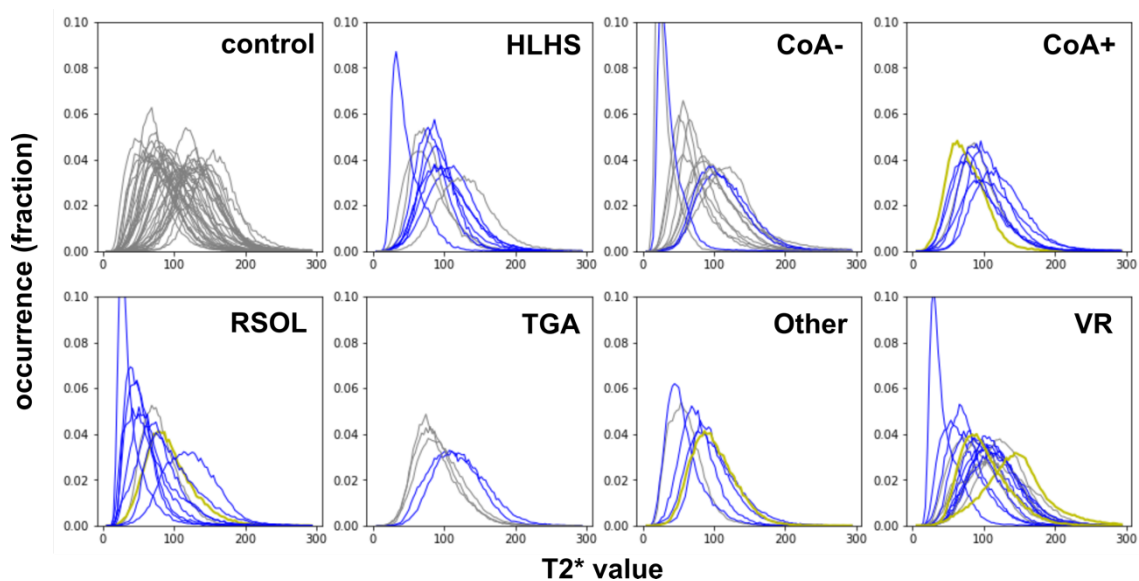
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702
703

Figure 4 - Individual Histograms

704 Individual Histograms, derived from about 70000 voxels per subject binned in 100,
 705 depicting occurrence fraction of T2* values over the entire placental volume from all
 706 participants, coloured by GA at scan. Groups from top left to bottom right. Control,
 707 Hypoplastic left heart syndrome (HLHS), coarctation of the aorta not confirmed (CoA-)
 708 and confirmed (COA+), right-sided obstructive lesion (RSOL), Transposition of the
 709 great arteries (TGA), Other lesions and suspected vascular ring (VR).



710

711 **Supplemental Figure 1 – Histograms with genetic test results**

712 Individual Histograms, as shown in Figure 4. Color coded by results of genetic testing.

713 Abnormal genetics (yellow), normal genetics (blue), not tested (grey), Hypoplastic left

714 heart syndrome (HLHS), coarctation of the aorta not confirmed (CoA-) and confirmed

715 (COA+), right-sided obstructive lesion (RSOL), Transposition of the great arteries

716 (TGA), Other lesions and suspected vascular ring (VR).

717

718

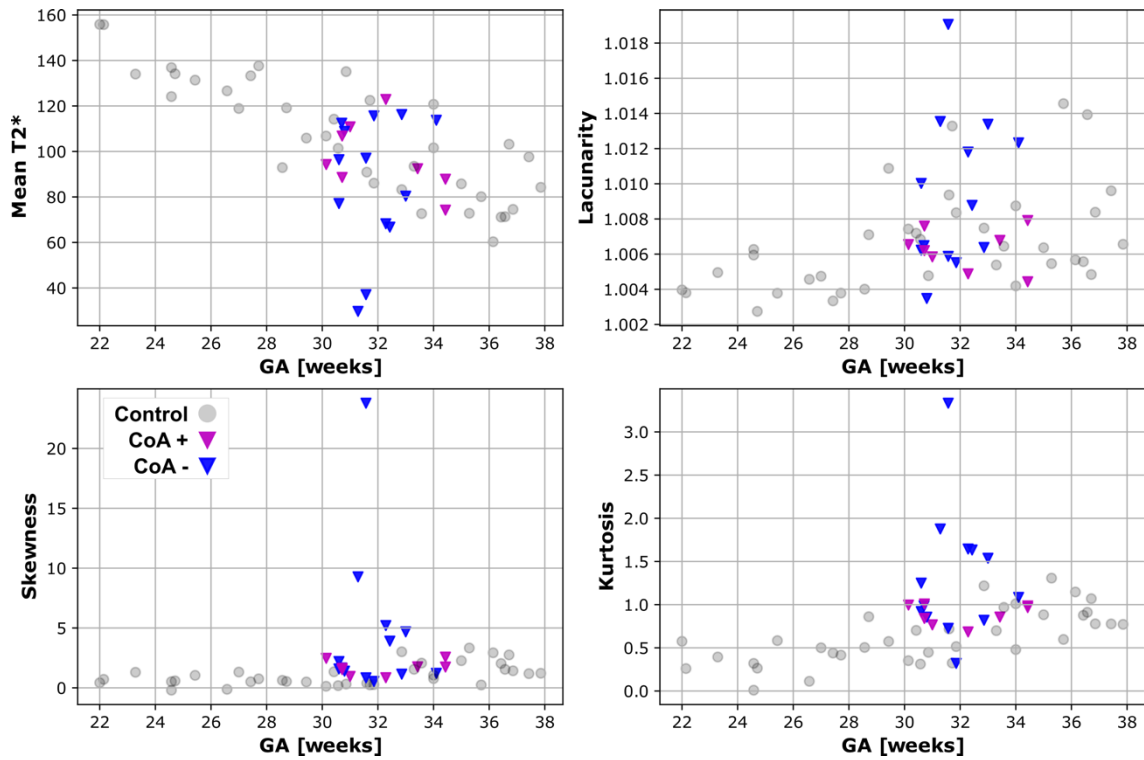
N (%)	Control	CHD
Available placental histology	15/37 (40%)	12/69 (17%)
MVM	1	2
With infarct	1	2
(Sub-)Chorionitis	8	7
With MIR	4	3
With FIR	5	4

719 **Supplemental Table 1 – Placental Histology**

720 Table shows available placental histology results. Maternal vascular malperfusion

721 (MVM), maternal inflammatory response (MIR), fetal inflammatory response (FIR).

722



723

724 **Supplemental Figure 2 – Coarctation of the aorta cohort and controls**

725 Mean T2*, Lacunarity, Skewness and Kurtosis over GA for all included controls in grey,

726 confirmed Coarctation of the aorta (CoA+) in pink and not confirmed (CoA-) in blue.

727