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## **Protein status in phenylketonuria: A scoping review**

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## **ABSTRACT**

**Background:** The physical and functional outcomes of lifelong treatment with a phenylalanine restricted diet for the management of Phenylketonuria (PKU) remain unknown. Given that the mainstay of dietary management consists of modifying the sources of ingested protein, various aspects of body protein status could be compromised.

**Objectives:** To examine the existing evidence regarding the protein status of people with PKU and identify nutritional and lifestyle variables that influence protein status.

**Eligibility criteria:** Studies reporting anthropometric, biochemical and/or functional measurements of body protein status in people with PKU were eligible.

**Source of evidence:** MEDLINE (Ovid), Embase (Ovid), CENTRAL, Web of Science and Scopus, and conference abstracts.

**Results:** Seventy studies were included in the review. The majority of studies assessing protein status based on anthropometric measurements observed no differences between people with PKU and controls, although deficits in muscle mass were reported within PKU cohorts. Findings for biochemical assessment of protein status were mixed and limited studies assessed protein status using functional measures. Factors such as participant age, sex, metabolic control, protein source, type of protein substitute, and pharmacological treatments were found to modulate protein status of people with PKU.

**Conclusions:** Findings were inconclusive regarding body protein status in people with PKU. The relationship between diet and protein status outcomes remains unclear and further research is warranted to determine the impact of dietary regimens on physical and functional outcomes, and to understand the best clinical assessments to reliably monitor the protein status in people with PKU.

**Keywords:** amino acid kinetics; biochemical markers; body composition; muscle function; phenylketonuria; protein status

## INTRODUCTION

Phenylketonuria (PKU; OMIM 261600) is an inherited metabolic disorder characterised by an accumulation of phenylalanine in the blood due to a deficiency in phenylalanine hydroxylase that converts phenylalanine to tyrosine. If left untreated, high serum phenylalanine levels result in irreversible neurocognitive disability. Dietary intervention remains the cornerstone management, and consists of limiting dietary phenylalanine intake through adherence to a low protein diet and inclusion of phenylalanine-free or low-phenylalanine protein substitutes to meet protein requirements (1,2).

Protein substitutes predominantly consist of L-amino acids (L-AA) and are an essential component of the dietary management of PKU. Studies that profile the blood amino acid kinetics in healthy individuals following ingestion of elemental amino acids have provided useful insight into increased amino acid oxidation rates and a decrease in whole-body protein retention compared with ingestion of whole protein sources (3–8). Accordingly, and as a compensatory measurement, dietary recommendations advise a correction factor of 20-40% in excess of the protein requirement guidelines for the general population (1,2,9). However, the scientific evidence underpinning protein requirement guidelines for people with PKU is limited and, as such, are based on extrapolations from studies that estimate protein requirements in healthy populations (10). Moreover, these studies utilised nitrogen balance methodology (11) that determines the minimum nitrogen requirement to balance nitrogen losses, as opposed to a requirement that serves to optimise health and functional outcomes in a given population group (12,13).

Whilst current dietary guidelines are effective in maintaining blood phenylalanine concentrations at levels to support neurological development, the impact of a lifelong PKU diet on physical and functional outcomes remains unknown. Concerns have been raised regarding the impact of standard dietary practices on growth, rates of obesity and bone health in people with PKU, with conflicting results reported (14–20). Limited attention has focused on the impact of a PKU diet on functional outcomes such as skeletal muscle mass (SMM) that serves as the major storage site for amino acids. Muscle mass is a key determinant of an individual's physical and functional ability, and plays an underappreciated metabolic role in

reducing risk of cardiometabolic diseases, including obesity, cardiovascular disease, diabetes and hyperlipidaemia (21). Moreover, the age-related decline in SMM begins as early as the fourth decade of life and continues to progress with age (22). However, it remains unknown whether existing protein requirement guidelines for people with PKU adequately offset age-related changes in protein metabolism across the adult lifespan in order to optimise functional and health outcomes (10).

The primary aim of this scoping review is to characterise the protein status of people with PKU across the lifespan, as determined by a combination of anthropometric, biochemical, and functional measurements. The secondary aim is to identify key nutritional and lifestyle variables that influence protein status in people with PKU. Understanding how PKU and associated dietary components facilitate or compromise the protein status of the individual is crucial to inform future research into personalising protein recommendations for individuals of all ages with PKU. In this regard, a scoping review is pertinent to enable the breadth of evidence to be mapped and synthesised given the wide focus of the research questions.

## **METHODS**

This scoping review was undertaken in accordance with the five-stage framework outlined by Arksey and O'Malley (23), and is reported in accordance with the PRISMA Extension for Scoping Reviews checklist (24). A full peer-reviewed protocol is available (25).

### **Identifying relevant studies**

#### **Information sources and search strategy**

MEDLINE (Ovid), Embase (Ovid), CENTRAL, Web of Science and Scopus were searched between June and July 2021 to identify relevant literature. No date restriction was applied and all articles up to July 2021 were included. Only English language articles were selected. Abstracts from relevant conferences held between 2010-2020 and reference lists of eligible full-text articles were manually searched for additional literature. Corresponding authors were contacted to request articles not available in full text. The search strategy was developed by

the research team, with guidance from a Librarian (see supplemental material for example of MEDLINE (Ovid) search strategy).

### **Eligibility criteria**

The Joanna Briggs Institute population, concept, context (PCC) strategy was used to develop the inclusion and exclusion criteria (25). Studies were considered for inclusion if they included participants with PKU, and all age categories and study designs were considered (*Population*). Pregnancy and the presence other co-morbidities that could influence protein intake were excluded. Given that the focus of this scoping review was on body protein status, eligible studies reported anthropometric, biochemical and/or functional measurements of protein status in people with PKU (*Concept*). Studies from all geographical areas were considered (*Context*).

### **Study selection**

All records were uploaded to Zotero 5.0 (George Mason University, USA). Following the removal of duplicate records, two independent reviewers (SF, MOK) applied the pre-defined eligibility criteria and independently assessed eligible titles and abstracts. In accordance with the published protocol (25), studies were excluded if they reported body weight, height, growth parameters (height, growth rate and head circumference), body mass index, fat mass (FM), or bone mineral density, with no other measurements of protein status. Full texts of eligible articles were retrieved and independently assessed by the same reviewers (SF, MOK). Studies were included if they reported at least one measurement of protein status (Table 1). Reasons for exclusion of full-text articles are outlined in Figure 1. Following each stage of the selection process, the reviewers compared results and reached a consensus.

**[Insert Table 1: Examples of anthropometric, biochemical and functional measurements of protein status]**

### **Data charting process and items extracted**

Data from eligible full-text articles were independently tabulated by the reviewers (SF, MOK), using an extended version of the data charting tool outlined in the protocol. The tool was piloted and modified to ensure all relevant information was extracted. Only anthropometric, biochemical and functional data relevant to the measurement of protein status (Table 1) were extracted. Factors modulating protein status or variables (either controlled or stratified by) were extracted if relevant to the assessment of protein status. The majority of studies included multiple outcome measurements but only the findings pertinent to protein status were tabulated. Additionally, primary outcomes and pharmacological interventions (eg. BH4 or sapropterin) were extracted. Due to limited availability, these data were not tabulated, but will be discussed herein. Protein intake data were extracted (see supplemental material).

### **Data Synthesis**

The data is synthesised and reported under three main sections: anthropometric, biochemical, and functional measurements of protein status and variables influencing protein status are discussed under each section. Age categories were defined as: ‘children’ <10 years of age, ‘adolescents’ 10-19 years of age, ‘adults’ >19 years of age, and ‘older adults’ >60 years of age. Prealbumin, otherwise known as transthyretin, will be referred to as prealbumin throughout the text.

**[Insert Figure 1: Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) study flow diagram outlining the process of study selection]**

## **RESULTS**

The electronic database search identified 1353 papers and an additional 94 were obtained through manual searching (Figure 1). Following the removal of duplicates, 860 titles and abstracts were assessed resulting in 102 studies for full text review. Thirty-two studies were subsequently excluded, with reasons for exclusion outlined in Figure 1. Full data extraction was performed on 70 studies and sample size ranged from one (7) to 174 participants (26). Of

the 70 studies which reported outcomes relevant to protein status, 24 reported anthropometric, 32 biochemical, and 14 included mixed outcome measurements of body protein status. No studies measured protein status in older adults (>60 years of age). Twenty-six studies included participants that were diagnosed through newborn screening or defined as being early diagnosed for PKU. Six studies included both early and late diagnosed, but provided no further information, and 38 studies did not report time of diagnosis.

The extent to which protein status was reported varied between the 70 studies. Giovannini et al (2014) was the only study to state a primary outcome measurement that was related to protein status. Nineteen studies focused on investigating protein status or specific marker(s) of protein status and, of these, 6 included anthropometric methods (27–32), 11 included biochemical methods (7,33–42), and 2 included multiple methods (43,44). The remaining 50 studies were not primarily focussed on assessing protein status but included relevant markers.

*What is the existing evidence of the protein status of people with PKU across the lifespan?*

## **Anthropometric assessments of protein status**

### *Study characteristics*

Study characteristics are summarised in Table 2. Thirty-seven studies, including 24 full text papers and 13 conference abstracts, reported anthropometric assessments of protein status. Seven were interventional studies but only three reported the study design; two had a retrospective design (45,46) and one was a 3-year prospective longitudinal study (27). The remaining 30 studies were non-interventional, of which 53% were cross-sectional (28,29,31,43,44,47–57). Nine studies did not state the study design; however, based on the reported methods, all were non-interventional (30,32,58–64).

One study was conducted in children only (47), 15 in children and adolescents (aged 2 months to 19 years) (27,44,48,49,54–56,58,59,62,65–69), 11 in adolescents and adults (age range 15–50 years) (43,45,46,53,57,61,63,64,70–72), and one in adults only (60). Seven studies



included a mixed age cohort (age range 4 to 54.6 years) (28–31,50–52). Two studies did not report participants age (32,73).

#### *How is body protein status measured?*

Methods to assess body protein status are presented in Table 3. The majority of studies utilised bioelectrical impedance analysis (BIA) (32,45,46,49–52,57,64,65,70–72), dual-energy X-ray absorptiometry (DEXA) (27,29,30,53,56,62,63,68) or skinfold techniques (31,44,55,58,66,69) for the anthropometric assessment of protein status. Protein status was also measured using deuterium dilution technique (73), total body electrical conductivity (48), peripheral quantitative computed tomography (43), total body potassium (59), and prompt gamma neutron capture analysis for total body nitrogen (TBN) (44). Two studies used more than one method to assess protein status; Dobbelaere et al (2003) used skinfolds and BIA, whilst Wilcox et al (2011) used DEXA and gamma neutron activation analysis. Evans et al (2018) measured fat-free mass (FFM) using both BIA and total body water by deuterium dilution technique (TBWDeut) to determine the validity of BIA as a measurement of body composition in children and adolescents. This study reported no difference in FFM between the two measurements and confirmed that FFM calculated from BIA was correlated with FFM calculated from TBWDeut. Three studies did not report the method used for determining protein status (54,61,67).

#### *Key findings from anthropometric assessments of protein status*

Table 4 describes the main findings for anthropometric outcome measurements related to protein status. The majority of studies that compared cohorts of people with PKU to healthy controls reported no significant differences in protein status, regardless of age (32,44,47,48,50–52,56,62,65,71,73). However, in one study, TBN was lower in participants with PKU and, within each age, children with PKU accrued 53g less TBN compared to controls, equating to a six-month lag in TBN in the PKU group (44). Longitudinally, the annual accretion of nitrogen was comparable between groups. Moreover, in a similarly aged PKU population, lower total body potassium z-scores were reported compared to controls (59). A reduction in muscle cross-sectional area (MCA) was also reported in adolescent and adult

participants with PKU, with 30% exhibiting a MCA below the third percentile (43). Dietary adherence in this study was variable, whereby 59% of participants were following a PKU diet plus protein substitutes and a further 7% of participants were following a PKU diet with no supplementation. In contrast, Paci et al (2018) reported a greater FFM and total body water content in children and adolescents with PKU compared to controls, although neither the methods of assessments nor protein intake data were reported.

There was greater variation regarding the assessment of protein status within cohorts of people with PKU. In children and adolescents, total body protein and muscle mass were normal (55), and consistent findings were reported in adults (60). Two studies reported a lower protein status in people with PKU, with 30.8% of child and adolescent participants recording a deficit in fat-free mass index (FFMI) (54), whilst in a mixed-age cohort, a 41.5% overall deficit in muscle mass was identified (31). Of those participants that reported a deficit in muscle mass, the majority (87.5%) were of normal weight-for-age and of adolescent age (78%) (31). Regarding the relationship between body weight and protein status, Rocha et al (2010) found that adolescents and adults with PKU who were classified as overweight and obese reported a lower FFM percentage compared to other participants. No data regarding protein intakes were reported in the above studies; however, Rocha et al (2010) observed no difference in the median intakes of natural protein and protein substitute in participants classified as overweight/obese compared to other participants with PKU. Three studies revealed positive correlations between FFM and bone mineral density (BMD) (56,57) and bone strength (43) in people with PKU, and FFM rather than FM had a greater effect on BMD. In one study, these findings were limited to females only (57).

### *Factors modulating anthropometric outcomes of protein status*

#### *Gender*

Several studies focused on factors known to influence protein status. Stratified by sex, Sailer et al (2020) found male children and adolescents to exhibit a lower percentage lean body mass (LBM) compared to healthy controls, whereas Allen et al (1995, 1996) found female children and adolescents to have lower total body FFM and TBN, respectively, when compared

to controls. Sailer et al (2020) reported no differences in total protein intake between people with PKU and controls, and no differences in total protein, natural protein and protein substitute intake were detected between males and females with PKU (see supplemental material). However, when considering percentage of total energy from protein and protein to energy ratio, protein intakes were lower in males with PKU compared with controls. These differences in protein intake were not observed in females. No sex comparisons of dietary protein intake were reported by Allen et al (1995, 1996). In contrast, one study reported no sex differences in body composition, although in the mild hyperphenylalaninemia (HPA) group, increased LBM was identified in pubertal compared to prepubertal participants (56).

Within a PKU cohort, adolescent and adult males reported a higher LBM and appendicular lean mass compared to females (53). However, no sex differences were observed in appendicular lean mass index (ALMI) or ALMI z-scores, although the mean ALMI among younger males was close to a cut-off point previously suggested to identify sarcopenia in older men. The mean total protein substitute consumed was greater in males compared to females but did not reach statistical significance. The intake of protein substitutes, expressed per kilogram of body weight or per kilogram of lean mass, were not statistically different between males and females.

### *Dietary factors*

The impact of natural protein and/or protein substitute intake on protein status has also been examined. In children and adolescents, Huemer et al (2007) reported that natural protein intake correlated with FFM, explaining 47.7% of the variance in FFM. This relationship was also observed in a mixed-age cohort. Jani et al. (2017) correlated high natural protein intake with high FFMI and low FMI:FFMI ratio in adults with PKU, whereas in children, protein substitute and total protein intake were associated with FFMI. In a mixed-aged PKU cohort, the percent of total protein intake, expressed relative to DRI, positively correlated with muscle mass (52). Moreover, a similar relationship between protein intake and protein status in children and adolescents was reported but in the context of percentage of FM rather than percentage of FFM; with a decline in percentage FM there was an assumed proportionate

increase in FFM (65). A negative correlation was reported between total protein, natural protein and protein substitute intake and percentage of FM, that presumably translates to a positive correlation with percentage of FFM (65). In contrast, no differences in body composition parameters were reported in adolescents and adults with PKU that were consuming vegan diets, vegan diets plus protein substitutes, non-restricted diets (72) or PKU-diets (64,72).

### *Metabolic control*

Metabolic control has been reported to impact protein status. Poorer metabolic control (i.e., higher phenylalanine levels) resulted in improved protein status (32,66). The cause for higher phenylalanine levels in participants included in these studies is unclear and could be related to increases in habitual dietary protein intake and / or reduced intake of protein substitutes. In contrast, one study reported no difference in LBM between the pubertal PKU sub-groups with high phenylalanine levels and participants with blood phenylalanine levels within target range (30). However, when analysed using LBM z-scores, a significant difference was reported with lower LBM z-scores in those participants with higher phenylalanine levels. No difference was reported in LBM z-scores between the pre-pubertal and the pubertal subgroup with recommended phenylalanine levels, indicating no impact of pubertal status on LBM z-scores. Choukair et al (2017) reported no relationship between MCA and classification of PKU or phenylalanine levels.

### *Type of protein substitute*

Two studies investigated LBM in children and adolescents with PKU that consumed either casein glycomacropeptide (cGMP) or L-AA (27,68). No significant differences between groups were found at baseline; however, Daly et al (2019) reported higher FFM values in those participants taking cGMP compared to L-AA after 36 months. Whilst not significant, Daly et al (2021) demonstrated improved LBM in people with PKU taking a cGMP-based protein substitute. No differences in body composition parameters were reported in adolescents and adults taking L-AA compared to cGMP (45,46).

### *Pharmacological management and other factors*

Two studies measured protein status outcomes in participants taking BH4 and reported no difference in percent FFM compared to participants on diet only (65), and in branchial muscle area compared to general population reference data (69). Genotype (29,53) nor socioeconomic status (66) were associated with protein status.

## **Biochemical assessments of protein status**

### *Study characteristics*

Forty-four eligible articles, 33 full text and 11 abstracts, included biochemical assessments of protein status (Table 2). Eighteen articles were intervention studies, of which seven reported the study design. Randomised control trials were conducted by Prince et al (1997), Giovannini et al (2014) and Ney et al (2016). Two studies were prospective (74,75), and two were retrospective longitudinal studies (45,46). The remaining 11 studies did not identify the study design; however, based on the reported methods, had an interventional design (7,36,39–41,69,72,76–79). Twenty-six studies were non-interventional and the majority utilised a cross-sectional design (44,56,57,80–82). Thirteen studies failed to state the study design, but were non-interventional based on the methods reported (33,34,38,42,62–64,83–88).

There was greater variation in participant age for studies including biochemical measurements of protein status than anthropometric measurements of protein studies. Six studies were conducted in children (age range 13.7 days to 10 years) (33,38,42,84,85,89), ten studies were in children and adolescents (age range 2.4 months to 18 years) (44,56,62,69,74,76,77,83,86,90), 15 in adolescents and adults (11–49 years) (7,39,40,45,46,57,63,64,70,72,75,79,91–93) and two in adults only (41,82). Seven studies included mixed-age cohorts (age range 7 months to 54 years) (26,36,37,80,81,94,95). Four studies failed to report participant age (34,78,87,88).

### *How is protein status measured using biochemical assessments?*

A range of biochemical measurements were used to assess protein status, and most studies included multiple measurement (Table 3). Serum albumin (36,41,42,56,62,63,69,72,74,78,85,86,95), prealbumin (26,34,37,57,80,81,84,87,89) or both (33,38,40,45,46,76,77,82,83,88,90,91,93,94) were the most common biochemical measurements reported. Total protein (36,40–42,56,63,72,76,78,80,81,85,86,91,93,94) and blood urine nitrogen (BUN) (33,40,45,74,75,78,85,86,93) were also frequently used. Three acute phase studies were included: two studies measured whole-body protein metabolism using valine and leucine stable isotopes (39,41), and one measured nitrogen excretion and leucine kinetics (7).

#### *Key findings from biochemical assessments of protein status*

The key findings for biochemical assessments of protein status are summarized in Table 5. In children with PKU, no differences in albumin (33,38,42), total protein (42), retinol binding protein (33), BUN (33) or plasma essential amino acid (EAA) concentrations (except phenylalanine) were reported compared to healthy controls (89). In children and adolescents, studies reported albumin (56,62,83,85), total protein (56,83,85), creatinine (62), BUN (85) and/or plasma amino acid levels (44) were comparable to untreated HPA or controls. However, two studies identified lower concentrations of albumin (77,90), retinol binding protein (77), and higher amino acid ratios (90) in participants on a PKU diet compared to untreated HPA or controls. Furthermore, deficits in prealbumin (38,77,83,90) were found, and participants with low prealbumin levels were more likely to have an EAA deficiency (83). In contrast, Acosta et al (1999) and Prince et al (1997) observed prealbumin concentrations within normal reference ranges despite recording protein intakes below the recommended dietary allowance (RDA) (1980) and the Medical Research Council (1993) protein recommendations, respectively.

In two mixed-age cohorts, increased prealbumin concentrations were observed in PKU participants compared to controls, with no difference in albumin and total protein levels (88,94). In contrast, five studies reported low prealbumin levels in PKU participants (34,37,80,81,87); one study reporting prealbumin  $\leq 20$  mg/dL in 60% of participants (34) and

another reported prealbumin levels below the third percentile for 15% of PKU participants, despite average total and natural protein intakes (g/kg/day) of 117% and 48% of RDA, respectively (80). Total protein levels were similar between people with PKU and controls (80). Despite normal levels of total protein, abnormal albumin (high) and BUN (low) were identified in children and adolescents where total protein intake was  $62 \pm 15$  % of RDA (86). Low protein levels in adolescents and adults with low SMM, and normal protein levels in participants with high SMM were also observed (70). In contrast, Modan-Moses et al (2007) reported plasma albumin concentrations to be within range, and Prochazkova et al (2012) reported no significant differences in levels of prealbumin among a large mixed-age cohort of people with PKU and HPA.

Thompson et al (1990) and van Rijn et al (2007) investigated whole body protein metabolism in adolescents and adults with PKU compared to healthy participants. Both studies reported no differences in whole body protein metabolism between groups. In the study by van Rijn et al (2007), participants with PKU were prescribed total protein intakes (natural protein and protein substitute) to meet 120% of RDA. Half of the participants with PKU in the study by Thompson et al (1990) were consuming a normal diet, and plasma amino acid concentrations were at the lower end of the reference range compared to the participants on a PKU diet and controls. Thompson et al (1990) reported no association with plasma phenylalanine concentrations and protein synthesis rates.

### *Factors modulating biochemical outcomes of protein status*

#### *Type of protein substitute*

Protein substitutes also modulated various biochemical markers of protein status. Studies reported no difference in BUN (45,74,75,93), albumin (45,46,74) and prealbumin (45,46) concentrations in a mixed age cohort of people with PKU taking cGMP versus L-AA protein substitutes. However, van Calcar et al (2009) reported lower BUN with cGMP compared to L-AA protein substitutes. Ney et al (2016) reported higher albumin concentrations with the ingestion of cGMP than L-AA. When habitual protein substitutes were replaced with a protein substitute utilising prolonged release technology, Giovannini et al (2009) reported an

improvement in prealbumin concentrations within three days, and demonstrated a significant increase in prealbumin (90), plasma protein, albumin, and some amino acids (36) in participants who received the prolonged release protein substitute. These changes were not observed in participants taking only 80% of protein requirements from a prolonged-release protein substitute (36).

#### *Pharmacological management*

Studies that implemented BH4 therapy resulting in a corresponding increase in natural protein intake and reduction in protein substitute dose reported no changes in the majority of biochemical markers of protein status (69,76,78,91). However, an increase in prealbumin levels in children on BH4 treatment was reported by Singh et al (2010), despite total protein intakes remaining stable. In contrast, two studies reported lower BUN (78) and prealbumin (81) in participants on BH4 treatment. Adults treated with pegvaliase that recorded protein intakes that met or exceeded the RDA for protein reported no deficiencies in albumin, prealbumin and essential amino acids (82).

#### *Gender, age, metabolic control, dietary and other factors*

Sumanszki et al (2019) reported prealbumin concentrations among adolescents and adults with PKU to be higher in males compared to females. Age and metabolic control were reported to correlate with prealbumin levels across all age-categories. Desloovere et al (2014) stated a correlation between prealbumin and age but provided no supporting data. Lower prealbumin levels were reported in participants who were younger (37,81,83,84,94) and those with better metabolic control (80,81,83,84,94). In contrast, Rocha et al (2010) and Desloovere et al (2014) reported no correlations between prealbumin levels and median blood phenylalanine levels. Furthermore, no significant correlations were reported between prealbumin levels and the amount of protein substitute prescribed (34,37) or classification of PKU (37), and between prealbumin or total protein and time of diagnosis (81). Prealbumin levels were observed to correlate with haemoglobin levels (87). Age was reported to positively correlate with serum



total protein levels (81) and albumin concentrations (95), whereas findings for metabolic control were inconsistent (42,81).

No associations between dietary protein intake and biochemical markers of protein status were reported across studies (42,62), although studies were limited. Moreover, whether participants consumed a vegan diet, vegan plus protein substitutes, unrestricted or PKU diets did not alter total protein levels (72). However, plasma amino acid levels were higher in those individuals on a PKU diet and serum urea concentration was low-to-normal in those on restricted protein diets (72), consistent with findings presented by Das et al. (2010). The only study to measure biochemical markers of protein status in response to exercise was conducted by Mazzola et al (2015). At baseline, participants with PKU reported lower levels of BCAA compared to controls, with BCAA levels not modified post exercise in both PKU participants and controls.

### **Functional assessments of protein status**

#### *Study characteristics*

Three studies included functional assessments of protein status, with one study specifically related to muscle strength (43) and two studies including assessments of physical performance (71,79) (Table 3). One study was cross-sectional in design (43), and the other two studies did not state the study designs but were interventional according to the methodology. All studies included adolescents and adults with PKU, and made comparisons to healthy controls (71,79) or reference population data (43) (Table 2).

#### *How is protein status measured using functional assessments?*

Choukair et al., 2017 used handgrip strength as a functional measurement of protein status, whilst Sumanszki et al (2020) and Mazzola et al (2015) measured VO<sub>2</sub> max as a functional measurement of physical performance (Table 3).

#### *Key findings and factors modulating functional protein status outcomes*

Table 6 describes the main findings for functional assessments of protein status. Compared to a healthy population, handgrip strength was reduced in participants with PKU, with over one third of participants recording handgrip force below the third percentile (43). Regarding exercise performance, Sumanszki et al (2020) reported a lower VO<sub>2</sub> max and cumulative workload was lower in the PKU group compared to controls (71). In contrast, Mazzola et al (2015) reported no differences in VO<sub>2</sub> max between PKU and controls.

## **DISCUSSION**

This scoping review is the first to examine the literature that describes the body protein status of people with PKU, with a specific focus on anthropometric, biochemical and functional assessments of protein status. Overall, the findings for anthropometric, biochemical and functional measurements of protein status were inconsistent. Whilst majority of studies reported no discernible differences in anthropometric parameters when comparing participants with PKU to healthy controls, a significant minority of studies reported deficits in muscle mass among PKU cohorts and warrant further investigation. Two studies identified 30-40% of participants with PKU to have deficiencies in muscle mass (31,43). In one study, 87% of participants with reduced levels of muscle mass were of normal body weight, thus emphasising the importance of body composition analysis in addition to body weight (31). This has been discussed in the literature in the context of body fat assessment in people with PKU (96), and our findings also support the recommendation of monitoring FFM in people with PKU across the lifespan.

BIA was the most common method for assessing body composition, likely due its practicality in the clinical setting and cost effectiveness. BIA has been validated for FFM assessment in children and adolescents with PKU (28); however, it remains unvalidated in adults and older adults with PKU. Given the increasing adult PKU population, validation is warranted. Furthermore, data from the general population demonstrates that skeletal muscle mass has an important role in bone health (21) and these findings are now corroborated in people with PKU (43,56,57). Concerns have been raised regarding the impact of the PKU diet on bone health, which will be further exacerbated with ageing. Therefore, monitoring, and optimising muscle mass in people with PKU play an important role in bone health outcomes.

Biochemical measurements of protein status are routinely used in monitoring patients with PKU, but a lack of consistency in the markers utilised has been identified. Most studies measured albumin and/or prealbumin concentrations. Albumin levels were predominantly comparable to healthy controls or reference data, whereas the prealbumin levels were generally lower in people with PKU than healthy controls or reference data. Due to its short half-life, prealbumin is favoured over albumin for monitoring changes in nutritional status

(97), and therefore findings related to prealbumin concentrations may provide greater insight into the protein status of people with PKU than plasma albumin concentrations. However, both markers are influenced by disease states, and therefore their specificity to protein status may be limited (97,98). Measurements such as D3-creatinine dilution, 24-hour urinary creatinine and 3-methylhistidine concentrations have been recommended for monitoring protein status (97,99), but no studies have conducted these measurements in people with PKU, and their practicality for clinical use may be limited. Future research is warranted to determine the most effective biochemical markers for monitoring protein status in people with PKU. At the level of whole-body protein metabolism, two studies reported comparable outcomes to healthy controls and mean total protein intake exceeded RDA by 20%, which aligns with current dietary guidelines (1,2,9).

It is clear that the functional measurements of protein status remain scarce, with only one study measuring muscle strength. Two studies measured physical performance as an indirect measurement of protein status. In this regard, both physical performance and muscle strength were compromised in participants with PKU (43,57). Muscle mass does not directly translate to functional ability (100), and therefore outcomes from anthropometric studies in PKU cannot directly be extrapolated to functional ability. Accordingly, future studies are warranted to advance understanding regarding how PKU dietary guidelines translate to functional outcomes. Moreover, physical activity is known to modulate protein status in the general population, with exercise improving nitrogen retention (101) and postprandial rates of muscle protein synthesis (102). With the exception of one study that reported the impact of acute exercise on plasma BCAA concentrations in participants with PKU (79), no studies have directly investigated the impact of physical activity on protein status in people on a PKU diet.

Studies included in this review were conducted between 1980 until 2021, a period that has seen significant advances in the dietary management of PKU. It is now recognised that protein requirements should exceed the RDA by 20-40% to compensate for the reduced uptake and utilisation of protein substitutes. Due to inconsistencies in studies reporting protein intake data, no conclusions could be made on whether differences in findings could be attributed to intakes of natural protein and protein substitutes. However, in studies reporting protein

intake data, dietary patterns did not translate to protein status outcome measurements, whereby protein intakes both above and below RDA had mixed findings. A limited number of studies utilised the current protein recommendation for PKU dietary management of 120-140% of RDA.

Several studies investigated the association between protein intake and protein status outcomes. Two studies reported that higher total protein intakes, as measured by 24 hour recalls and food records, resulted in improved protein status in children, adolescents and adults with PKU (52,65). In contrast, Huemer et al (2007) reported natural protein intake to be an important predictor of protein status in children and adolescents with PKU. Jani et al (2017) observed a similar association in adults with PKU, whereas total protein and protein substitute intake were shown to be of greater importance in children. In the context of a traditional PKU diet, there is limited scope for large increases in natural protein, thus highlighting the importance of preventing over-restriction by reassessing natural protein tolerance (103). Moreover, with pharmacological treatments there are now opportunities to improve natural protein intake. In addition to the amount of protein substitute, the type of protein substitute can modulate the protein status of an individual (27,36,40,45,46,68,74,75,77,90,93); however, findings were inconclusive and further research is warranted. A correlation between metabolic control and protein status was identified, whereby participants with poor metabolic control exhibited improved body composition parameters and higher prealbumin levels, possibility attributed to increases in natural dietary protein intake. Therefore, those individuals who have good metabolic control may be at greater risk of compromised protein status.

In addition to protein intake, participant sex and age were observed to modulate body protein status outcomes in people with PKU, consistent with data from a non-PKU population (104–106). However, the impact of sex on anthropometric outcomes in participants with PKU was inconsistent, whereas age was found to positively correlate with prealbumin and total protein levels, which has previously been described in the general population (107). The majority of studies were conducted in children, adolescents and younger adults, whereas no studies included older adults. This omission is likely due to when newborn screening of PKU was

introduced, limiting numbers within this cohort. However, individuals who have received dietary treatment for PKU are now approaching older age and therefore understanding the impact of dietary management on ageing is a key focus question.

#### *Protocol deviations and study limitations*

This scoping review was undertaken in accordance with the published protocol (25), with the only deviation being redefining the age categories to include adolescents. Limited studies had a primary focus on protein status outcomes, and therefore the majority of findings were extracted from studies where protein status measurements formed part of a wider nutritional status assessment or were reported as secondary findings. Almost a third of studies were abstracts and provided limited data. The heterogeneity in studies and outcomes restricted the conclusions that could be drawn on variables modulating protein status. However, studies that specifically investigated associations between natural protein intake, amount and type of protein substitutes, metabolic control and protein status provided some useful insight into areas that require further consideration in optimising the health outcomes of people with PKU. Some studies were impeded by small sample sizes, in that half of the studies had sample sizes of 30 or less participants. Additionally, the same patient populations may have been included in multiple studies and this may have affected the interpretation of results.

## **CONCLUSION**

The maintenance of SMM and function with advanced age is critical in reducing the risk of chronic disease and obesity, as well as optimising bone health. These conditions have been highlighted as concerns in the PKU population. However, limited studies and clinical guidance have specifically focused on monitoring and optimising FFM in people with PKU. Accordingly, it is currently unknown what the functional outcomes are for people with PKU following a phenylalanine restricted diet whereby the majority of protein is derived from elemental amino acids. Further research is warranted to understand both, the impact of PKU diet on functional ability, and the significance of currently used anthropometric, biochemical and functional

markers when assessing protein status in people with PKU. None of the currently used markers adequately assess body protein status; thus, emphasising the urgent need for research to establish robust clinical and biochemical markers of protein status that can be reliably used to monitor the impacts of dietary interventions in people with PKU.

Dietary adherence is known to decline with age, and therefore, conducting research in adults with PKU with a focus on investigating the impact of dietary management of health outcomes can be challenging. This challenge may be a contributing factor to the limited number of studies of protein status conducted in adults with PKU. With the consensus for life long treatment and as a result, a growing adult and ageing PKU population on dietary management, there is an urgent need for research specifically in adults with PKU to ensure dietary management leads to optimal health outcomes across the life course.

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## **Conflict of interest**

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## **Author Contributions**

SF, RR, KW, OW and MOK were involved in conceptualisation and development of the protocol. Database and manual searches were conducted by SF, and study selection, full text review and data extraction were completed by SF and MOK. SF led on preparing the original draft with support from MOK who was lead supervisor for this review. RR, KW, OW and MOK provided critical revision of the draft. SF, RR, KW, OW and MOK revised and approved the final manuscript.



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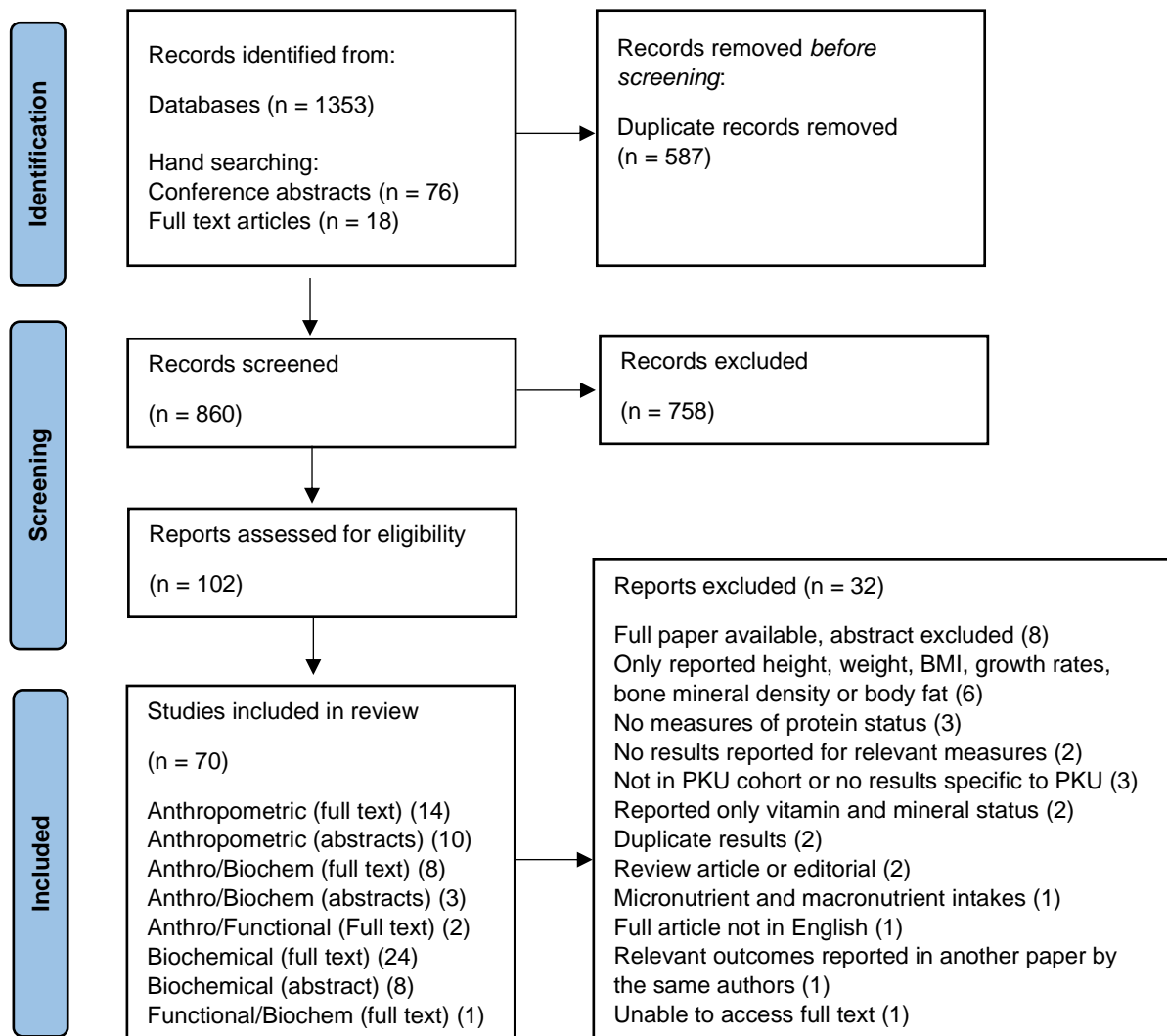
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**Figure 1.** Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) study flow diagram outlining the process of study selection



**Table 1: Examples of anthropometric, biochemical and functional measurements of protein status**

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|                       |  |
|-----------------------|--|
| <b>Anthropometric</b> | Body composition (fat free mass, lean body mass and / or skeletal muscle mass) via dual-energy X-ray absorptiometry (DEXA), bioelectrical impedance analysis (BIA), total body electrical conductivity (TOBEC), BodPod whole body air-displacement plethysmography or skinfolds.   |
| <b>Biochemical</b>    | 3-methylhistidine concentrations, albumin, pre-albumin, transthyretin, retinol-binding protein, urea production, blood urea nitrogen, urinary nitrogen, total body nitrogen, and whole-body protein metabolism. Plasma amino acids concentrations, urea production and creatinine where the author(s) have specifically used these as a measurement of protein status.   |
| <b>Functional</b>     | Hand-grip strength, the Short Physical Performance Test (SPPT; including tests of balance, gait speed, and timed sit-to-stand), one repetition max (1RM, or a five-repetition max for older adults), VO <sub>2</sub> max tests (or VO <sub>2</sub> peak for older adults) and other validated measurements of muscle function (i.e. isokinetic quadriceps strength using dynamometry and vertical jump performance using force platform technology). |

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*Retrieved from the published protocol (Firman et al., 2021)*

**Table 2: Study characteristics classified by measurement category.**

| Author(s) (Year)               | Study design        | Participants  | Intervention (duration of study)  |
|--------------------------------|---------------------|---|---|
| Country                        |                     | (N, age, sex)   |   |
| <b>ANTHROPOMETRIC MEASURES</b> |                     |   |   |
| <b>Daly et al. (2021)</b>      | Longitudinal        | 50 PKU (28 males, 22 females); 48 completed   | 2 types of protein supplements, allocation dependent on   |
| UK                             | prospective (3y)    | Age: AA 11.1y (range 5-15y); CGMP50 7.3y (range 5-15y); CGMP100 9.2y (range 5-19y).   | taste preference and phe levels:<br><br>(1) AA: protein substitute given as AA only;<br><br>(2) CGMP50: combination of CGMP-AA and AA;<br><br>(3) CGMP100: all their protein substitute as CGMP-AA. |
| <b>Daly et al. (2019)</b>      | Longitudinal        | 28 PKU (18 on cGMP-AA (9 male) and 10 on L-AA (7 male). Median  | Not applicable  |
| UK                             | prospective (36 mo) | age at study end: CGMP-A: 11.2y (range 8-19y) and L-AA: 15.9y (range 9-18y).  |   |
| <b>Evans et al. (2017)</b>     | Longitudinal        | 32 PKU on diet (D-PKU) (10 males, 22 females; Age $y \pm SD$ (range) $9.2 \pm 4.7y$ (0.83–18.0))  | Not applicable  |
| Australia                      | prospective         | 5 PKU treated with BH4 $\pm$ phe restricted diet (BH4-PKU) (3 males, 2 females; Age $y \pm SD$ (range) $8.8 \pm 4.6y$ (0.64–10.9)).<br><br>37 All-PKU (13 male, 24 female; Age $y \pm SD$ (range) $8.8 \pm 4.6$ (0.6–18.0)) |   |



|                             |                                  |   |                |
|-----------------------------|----------------------------------|---|----------------|
|                             |                                  | 21 controls (healthy sex and aged matched siblings): 8 males, 13 females, Age $y \pm SD 8.8 \pm 4.8y$   |                |
| <b>Paci et al. (2018)</b>   | Case-control                     | 25 PKU (aged 5-14y)   | Not applicable |
| Italy                       |                                  | 25 sex and aged matched controls  |                |
| <b>Rocha et al. (2013)</b>  | Case control                     | 89 PKU (age $14.4 \pm 6.6y$ ; 46% females)  | Not applicable |
| Portugal                    | cross-sectional                  | 78 controls (age $15.9 \pm 7.1y$ ; 58% females). 74.4% of controls were close relatives.  |                |
| <b>Huemer et al. (2007)</b> | Cross-sectional and longitudinal | 34 classical PKU (22 males, 12 females; mean age $8.7 \pm 3.9y$ , range 2 months to 15y)<br><br>34 healthy age- and sex-matched controls (mean age $9.2 \pm 3.7y$ , range 1 month to 16y) | Not applicable |
| <b>Sailer et al. (2020)</b> | Cross-sectional                  | 30 PKU (age 5-16y (mean $11.6y \pm 3.41$ ); 12 females, 18 males)   | Not applicable |
| USA                         |                                  | 30 control age- and sex- matched (mean age $11.75 \pm 3.49y$ ; 12 females, 18 males)  |                |
| <b>Weng et al. (2020)</b>   | Cross-sectional                  | 22 PKU (ages 8-27y, mean age $15.23 \pm 5.23 y$ ; 12 females, 10 males)   | Not applicable |
| Taiwan                      |                                  | 22 age- and gender-matched controls (ages 8-39y, mean $19.73 \pm 10.6y$ ; 12 females, 10 males)   |                |
| <b>Evans et al. (2018)</b>  | Cross-sectional                  | 16 PKU (7 males, 9 female). Median age 12.5y (range 5–20.6y)  | Not applicable |
| Australia                   |                                  |   |                |

|                                 |                 |   |                |
|---------------------------------|-----------------|---|----------------|
| <b>Stroup et al. (2018)</b>     | Cross-sectional | 15 (6 males and 9 females) (12 adults (aged 19-50y) and 3 adolescents (aged 15-17y))                            | Not applicable |
| USA                             |                 |   |                |
| <b>Jani et al. (2017)</b>       | Cross sectional | 86 PKU (59 children and 27 adults; 60.5% female)  | Not applicable |
| USA                             | study           | Age: Total sample: 16y (4 - 54.6y); Adults: 28.8 y (19.5-54.6), Children: 13.4y (4-18.9y))                      |                |
| <b>Torriente et al. (2017)</b>  | Cross-sectional | 12 with PKU (aged 3-18y). Both genders (number not specified)   | Not applicable |
| Cuba                            |                 |   |                |
| <b>Mazzola et al. (2016)</b>    | Cross-sectional | 27 PKU; 27 controls   | Not applicable |
| Brazil                          |                 | Aged (6-25y) and gender-matched (14 males and 12 females)   |                |
| <b>Kanufre et al. (2015)</b>    | Cross-sectional | 77 PKU (aged 5-25y)   | Not applicable |
| Brazil                          |                 |   |                |
| <b>Mexia et al. (2015)</b>      | Cross-sectional | 30 PKU; 57% female; mean age 12.2 ± 3.6y  | Not applicable |
| Portugal                        |                 |   |                |
| <b>Dobbelaere et al. (2003)</b> | Cross-sectional | 20 PKU (11 females; 9 males), age 8mo-7y (4.5 ± 1.6y/55 ± 19 mo).<br>Controls: Age (53 ± 19 mo) and sex matched | Not applicable |
| France                          |                 |   |                |
| <b>Nogueira et al. (2021)</b>   | Retrospective   | 53 PKU (aged 2-19y; (mean ± SD, 10.4 ± 4.6y); with 33 (62.3%) <12y old. 64% female (34/53)                      | Not applicable |
| Brazil                          | cohort study    |   |                |

|                               |              |  |   |
|-------------------------------|--------------|--|---|
| <b>Alfheaid et al. (2018)</b> | Not reported | 13 PKU and 10 health controls (matched for gender, age and BMI)  | Intervention not reported, measurements taken over 3 hrs after isocaloric meal. |
| UK                            |              |  |   |
| <b>Nalin et al. (2013)</b>    | Not reported | 23 PKU and 17 healthy, aged and gender matched controls  | Not applicable  |
| Brazil                        |              |  |   |
| <b>Adamczyk et al. (2011)</b> | Not reported | 45 PKU (aged $13.8 \pm 5.2$ y, range 4.9-21.9y), 25 males and 20 females   | Not applicable  |
| Poland                        |              | PKU subgroups =<br>1=pre-pubertal and normal phe levels, n=15<br>2a=pubertal and normal phe levels (n=18<br>2b=increased phe levels, n=12.<br>Compared to references for healthy children and adolescents <sup>a</sup> |   |
| <b>Wilcox et al. (2011)</b>   | Not reported | 42 PKU (33 females, 9 males) mean age $32.2 \pm 9.5$ y   | Not applicable  |
| Australia                     |              |  |   |
| <b>Bonifant et al. (2010)</b> | Not reported | 20 PKU   | Not applicable  |
| Australia                     |              | Data compared with normative data using matched controls   |   |
| <b>Rocha et al. (2010)</b>    | Not reported | 27 PKU (aged 18-38y)   | Not applicable  |
| Portugal                      |              |  |   |
| <b>Allen et al. (1995)</b>    | Not reported | 30 PKU (15 males, 15 females; aged 4.6-17.0y)  | Not applicable  |
| Australia                     |              | 76 controls (23 male, 42 females; aged 4.3-18.4y).<br>7 unaffected siblings and 69 non-familial  |   |

**BIOCHEMICAL MEASURES**

|                                 |  |  |   |
|---------------------------------|--|--|---|
| <b>Ney et al. (2016)</b>        | 2-stage, crossover RCT                             | 30 PKU (18 females and 12 males) included 5 minors (aged 15–17 y) and 25 adults (aged 18–49 y)   | 11-week protocol: 1 week education; 3 weeks GMP-MF or AA-MF; 3 weeks on usual routine with AA-MF (washout), then cross-over to 1 week education and 3 weeks with either GMP-MF or AA-MF   |
| <b>Giovannini et al. (2014)</b> | RCT  | 60 PKU (n=30 prolonged release group; n=30 in conventional substitute group); 55 completers (24 males, 31 females; age 9.2y (3.4y))<br>60 mild HPA (MHP) (26 males; age 9.3y (3.3y));<br>60 unaffected (26 males; age 9.2y (3.2y)) | Random allocation to prolonged-realise PHE-free protein substitute or current conventional substitute for 30 days.<br>Dose and frequency were tailored to needs of child.<br>Unable to blind participants.  |
| <b>Prince et al. (1997)</b>     | Phase 1: RCT<br>Phase 2: A historic control design | Phase 1: n=28 aged 4-10y<br>25 participants continued to phase 2 (completed the 5-y study).<br>Compared to non-PKU data from Armstrong and Stave 1973  | Phase 1: Duration 2 years.<br>Interventional: Random allocation to new AA mixture or standard mixture.<br>Phase 2: Compared the safety, efficacy, and acceptance of new treatment products developed with AA mixture tested in phase 1. Products nutritionally incomplete were taken with vitamin/mineral tablets |
| <b>Ahring et al. (2018)</b>     | Single-blinded, prospective, crossover             | 8 (7 female, 1 male), age 15–48 y (mean 33.25 ± SD 11.21)  | 24 h to 1 month<br>Intervention = 4 AA drink mixtures (DM)  |

|                           |  |   |   |
|---------------------------|--|---|---|
|                           | intervention study                           |   | DM1: Lacprodan CGMP-20; DM2: FSAA (equivalent AA profile as DM1); DM3: Lacprodan CGMP-20 and synthetic AA; DM4: FSAA (equivalent AA profile as DM3 but without Phe. 4 visits per patient, random allocation. Bloods at T0, T15, T30, T60, T120 and T240 min after meal. Test meal: total protein content was 25% of 1g /kg/day. |
| <b>Zaki et al. (2016)</b> | Prospective, self-controlled, clinical trial | 10 PKU (6 males, 4 females), aged 4-16y. Median IQR age 6.73 (5.02-11.79y)  | Two phases: Phase I was 9 weeks (50% GMP (cheese spread) and 50% AAF) and Phase II was 9 weeks (100% AAF).  |
| <b>Kose et al. (2019)</b> | Single center, case-control                  | 112 PKU (59 males, 53 females). Age: 136.8 ± 82.1 months (range: 18 to 377 mo)<br><br>PKU categorised into two groups:<br><br>Low dietary adherence (n=71, 41 females, 30 males; age 138.9 ± 80.1 mo (18-377 mo)); High dietary adherence (n=41; 12 female, 29 males; age 133.1 ± 84.4 mo(18-207 mo))<br><br>36 healthy controls (18 males, 18 females). Age: 119.7 ± 37.3 mo (73 to 206 mo). | Not applicable  |

|                                  |                 |   |                |
|----------------------------------|-----------------|---|----------------|
| <b>Prochazkova et al. (2012)</b> | Prospective     | 174 patients (113 children, 61 adults)  | Not applicable |
| Czech Republic                   |                 |   |                |
| <b>Viau et al. (2021)</b>        | Cross-sectional | 18 (mean age, SD 38.2 ± 8.8), 11 females, 7 males   | Not applicable |
| USA                              |                 |   |                |
| <b>Andrade et al. (2017)</b>     | Cross-sectional | 42 PKU (23 males, 19 females); Median age 10y (range 2-36y) 40 age and sex-matched controls                         | Not applicable |
| Spain                            |                 |   |                |
| <b>Crujeiras et al. (2015)</b>   | Cross-sectional | 156 PKU (46.8% male; range age: 7 months–42y old; 27.4% >18y)   | Not applicable |
| Spain                            |                 |   |                |
| <b>van Vliet et al. (2019)</b>   | Retrospective   | 12 with TT1 (mean age 13.5 ± 9.9, 75% male, 25% female)<br>92 with PKU (mean age 24.5 ± 13.9, 45% male, 55% female) | Not applicable |
| The Netherlands                  |                 |   |                |
| <b>Rocha et al. (2010)</b>       | Retrospective   | 69 treated PKU; 30 females (43.5%) and 39 males (56.5%) aged 1–27y (mean = 10y; SD = 6.47y)                         | Not applicable |
| Portugal                         |                 |   |                |
| <b>Gokmen-Ozel et al. (2009)</b> | Audit           | 34 PKU, 17 female, 17 male, median age of 15y (range 7–54y); 13 participants aged >18y of age.                      | Not applicable |
| UK                               |                 |   |                |
| <b>Kose et al. (2016)</b>        | Not reported    | 112 PKU<br>17 controls (age- and sex-matched)   | Not applicable |
| Turkey                           |                 |   |                |

|                                 |              |  |   |
|---------------------------------|--------------|--|---|
| <b>Desloovere et al. (2014)</b> | Not reported | 35 PKU   | Not applicable  |
| Belgium                         |              |  |   |
| <b>Schulpis et al. (2013)</b>   | Not reported | 54 PKU divided into groups based on metabolic control  | Not applicable  |
| Greece                          |              | Group A: 24 (12 males, 12 females, mean age $6.78 \pm 1.5$ y)<br>Group B: 30 (15 males, 15 females, mean age $5.0 \pm 3.2$ y)<br>50 age-and sex-matched controls (25 males, 25 females, mean age $7.68 \pm 2.3$ y) |   |
| <b>Douglas et al. (2013)</b>    | Not reported | 57, unclear how many participants were responders  | Sapropterin for 1 month, followed by phe challenge and classified as Definitive (DR) or Provisional (PR) Responders. Non-responders (NR) discontinued drug. Protein status assessed after 1 year  |
| USA                             |              |  |   |
| <b>Singh et al. (2010)</b>      | Not reported | Stage 1: 10 (9 males and 1 female) mean (SD) age 8.7y (2.5y);<br>Stage 2: 6 (6 males).   | Stage 1: BH4 response testing. BH4 administered OD 20 mg/kg/d. If plasma phe decreased by at least 30% after 1 wk = responsive. Dietary phe tolerance determined via milk challenge.<br>Stage 2: Protein substitute and protein intake adjusted to keep phenylalanine in target.<br>Follow-up for 24 months as per standard care. |
| USA                             |              |  |   |

|                                 |              |   |  |
|---------------------------------|--------------|---|--|
| <b>Giovannini et al. (2009)</b> | Not reported | 28 treated HPA (aged 6–18 y)<br>56 untreated mild HPA<br>56 controls matched for age and gender.  | Slow-release protein substitute given to 28 treated HPA and dietary intake and biochemical parameters reassessed after 3 days.   |
| Italy                           |              |   |  |
| <b>Rocha et al. (2009)</b>      | Not reported | 60 PKU  | Not applicable   |
| Portugal                        |              |   |  |
| <b>van Calcar et al. (2009)</b> | Not reported | 11 subjects (age range: 11–31y; 7 males and 4 females)  | Two dietary treatments of 4 d each: AA diet (days 1–4) and GMP diet plus multivitamin as GMP nutritionally incomplete (days 5–8). Dietary menu (24-h) repeated on all days of diet treatment.<br><br>Meals were timed to usual routine and PA was permitted but limited. Bloodspots taken d1-2. All blood samples drawn daily 3 h after the start of breakfast or 2.5 h after eating breakfast (days 3–8). |
| USA                             |              |   |  |
| <b>van Rijn et al. (2007)</b>   | Not reported | 6 PKU ( $27 \pm 7$ y; 3 females and 3 males)<br>6 controls ( $32 \pm 4$ y, 4 females and 2 males) | Isocaloric meal for 1 day providing 0.8g protein/kg/day, plus additional 20% from PKU3 supplement<br><br>Baseline bloods and breath samples taken.<br><br>T0-T60min whole-body $\text{NaH}_2\text{CO}_3$ production was measured using a primed constant infusion of $\text{NaH}_2\text{CO}_3$ .<br><br>Regular breath samples taken.  |
| The Netherlands                 |              |   |  |



T60-T420 min: NaH<sub>13</sub>CO<sub>3</sub> infusion replaced with L-[1-<sup>13</sup>C]-valine bolus followed by a continuous infusion  
 Regular blood and breath samples taken. At 1200 h, the meal period was started by consumption of the first meal and continued for 4 h by consumption of a meal every 60 min. After the start of the meal period, blood and breath samples were taken every 30 min for 3 h and during the last hour samples were taken every 15 min.

|                                 |              |  |   |
|---------------------------------|--------------|--|---|
| <b>Giovannini et al. (2006)</b> | Not reported | 13 PKU (7 females, 6 males; mean age 14y, range 5-26y)   | Participants randomised to new AA mixture for 6 mo:<br>Group 1: 100% daily N needs<br>Group 2: 80% daily N needs<br>Blood markers measured at baseline and 6mo.       |
| Italy                           |              |  |   |
| <b>Arnold et al. (2002)</b>     | Not reported | 38 PKU (mean age was 8.9y)   | Not applicable  |
| USA                             |              |  |   |
| <b>Arnold et al. (2001)</b>     | Not reported | 41 PKU (24 males, 17 females); age 1-16y; Age- and gender-matched patients (non-familial). Prealbumin controls were not available. | Not applicable  |
| USA                             |              |  |   |
| <b>Mönch et al. (1996)</b>      | Not reported | Study 3: 10 (aged 12-23y)<br>Study 4: 1 adult with PKU (female)  | Study 3: Day 1: AA mixture in two divided portions; Day 2: AA mixture in three divided portions<br>Study 4: <sup>13</sup> C-L-leucine (3 mg/kg) was given as a single |
| Germany                         |              |  |   |

|  |              |  |  |
|--|--------------|--|--|
|  |              |  | bolus together with the AA mixture taken at breakfast. Day 1: AA mixture taken in one single dose at breakfast. 5 days later: repeated with dividing the AA mixture into three portions per day.   |
| <b>Acosta et al. (1999)</b><br>USA         | Not reported | 35 with PKU (15 females and 20 males). Subjects entered the study at 13.7 ( $\pm 1.9$ SEM) days of age.<br><br>Data was compared to normal reference data                                  | Not applicable   |
| <b>Graffin et al. (1995)</b><br>USA        | Not reported | 6 PKU (3 males, 3 females, aged 3-16y)   | Not applicable   |
| <b>Thompson et al. (1990)</b><br>Australia | Not reported | 10 classical PKU (8 male, 2 females; mean age 19.3 y, range 14-24)<br>2 HPA (1 male, 1 female; ages 22 and 45 y)<br>6 age-matched normal controls (all male, mean age 20.8 y, range 19-23) | Radioisotope infusion: Isotope: priming bolus doses of L-[1- <sup>13</sup> C]leucine and sodium [1- <sup>13</sup> C]bicarbonate followed by continuous infusion of L-[1- <sup>13</sup> C]leucine was then given over the next 4-6 h (6 h PKU and HPA; 4 h controls).<br><br>Blood and expired air samples were collected at 15- to 20-min intervals in the final 2 h of each infusion and at 1/2-h intervals from 2-4 h in PKU participants. |
| <b>Nord et al. (1988)</b><br>USA           | Not reported | 50 children with PKU<br><br>13 children with HPA   | Not applicable   |

|  |   |  |                |
|--|---|--|----------------|
| <b>Shenton et al. (1983)</b>                     | Not reported  | 20 treated PKU (age 2-9y)  | Not applicable |
| UK   |   | 58 controls (age 1-15y). Control group = children with mild neurological disease, or children for elective operations or other admissions. |                |
| <b>Pena et al. (2018)</b>                        | Systematic review & meta-analysis of observational & interventional studies | 72 participants included in the meta-analysis  | Not applicable |
| Portugal and UK <sup>b</sup>                     |   |  |                |
| <b>ANTHROPOMETRIC + BIOCHEMICAL MEASUREMENTS</b> |   |  |                |
| <b>Boros et al. (2015)</b>                       | Case-control  | 27 PKU (aged 16-44y). Compared to age and sex matched reference values   | Not applicable |
| Hungary  |   |  |                |
| <b>Allen et al. (1996)</b>                       | Cross sectional with longitudinal cohort                                    | 37 PKU (aged 7.3 ± 2.0y, range 3.9-11y; 21 male, 16 female)<br>27 control children (aged 8.1 ± 1.9y, range 4-11.5y; 15 male, 12 female)    | Not applicable |
| Australia  |   |  |                |
| <b>Sumanszki et al. (2019)</b>                   | Cross-sectional   | 80 PKU (41 premenopausal women, 39 males (aged 18-49y))  | Not applicable |
| Hungary  |   |  |                |

|                                  |   |  |          |   |
|----------------------------------|---|--|----------|---|
| <b>Doulgeraki et al. (2014)</b>  | Cross-sectional                         | 48 PKU (25 males, 23 females; mean age $10.9 \pm 3.43y$ )<br>32 mild mHPA (18 male, 14 females; mean age $10.85 \pm 3.6y$ )<br>57 age and sex-matched controls | Greece   | Not applicable  |
| <b>Pena et al. (2021)</b>        | Retrospective longitudinal <sup>c</sup> | 11 PKU (8 females, 3 males). Mean age at CGMP-AA onset 28y (range 15-43y).   | Portugal | cGMP-AA (mean of 29 months)<br>cGMP-AA either fully or partially replaced L-AA; cGMP-AA: 100%, n = 4, 50% to <100%, n=4, <50%, n = 3.   |
| <b>Pinto et al. (2017)</b>       | Retrospective, longitudinal             | 11 PKU (8 females, 3 males) with PKU had a mean age of $27 \pm 10y$ (2 patients <18y)  | Portugal | cGMP-AA (either completely or partially) replaced AA, depending on patient preference when refusing to take AA (mean of $13 \pm 7$ mo). |
| <b>Das et al. (2013)</b>         | Not reported                            | 51 PKU (age range 16–44y, mean $\pm$ SD $26.6 \pm 6.6y$ ; 32 females, $26.5 \pm 6.2y$ ; 19 males, $26.8 \pm 7.7y$ )  | Germany  | All patients not taking AAM at the beginning of the study subsequently agreed to supplement their original diet with an AAM.            |
| <b>Das et al. (2010)</b>         | Not reported                            | 51 PKU (age 17–44y, 31 females, 20 males)  | Germany  | Not applicable  |
| <b>Modan-Moses et al. (2007)</b> | Not reported                            | 31 PKU (18 females, 13 males; mean age $25 \pm 5.3y$ (range 19–41))  | Israel   | Not applicable  |

|   |                       |  |  |
|---|-----------------------|--|--|
| <b>Lambruschini et al. (2005)</b>               | Not reported          | 14 PKU for BH4 treatment; 11 responders (aged 0.2-12.2y, 7 females, 4 males). Compared to age- and sex-specific percentiles for healthy population.              | BH4 treatment for 1 year. Initial dose 5mg/kg/day. Phe restricted diet was progressively liberalised by adding 200mg Phe/day/week for 2 mo, while formula was gradually reduced until complete removal was achieved. |
| <b>Hillman et al. (1996)</b>                    | Not reported          | 11 PKU (mean age 10.9 ± 4.2y; 5 male, 6 female),<br>64 controls (mean age 11.4 ± 4.2y; 32 male, 32 female) (11 were matched to the PKU children for sex and age) | Not applicable   |
| <b>ANTHROPOMETRIC + FUNCTIONAL MEASUREMENTS</b> |                       |  |  |
| <b>Choukair et al. (2017)</b>                   | Cross-sectional study | 56 PKU (16 male, 40 female), aged 26.0 ± 8.9y (range, 11.8–41.5y).<br>700 reference population data also used  | Not applicable   |
| <b>Sumanski et al. (2020)</b>                   | Not reported          | 12 PKU males (median age 26 (18-41y))<br>10 healthy controls (median age 26 (24-27y))  | Day 1: Stress stimuli: cold pressor test (CPT) and isometric handgrip test (HGT).<br>Day 2: peak treadmill test to exhaustion.   |
| <b>FUNCTIONAL + BIOCHEMICAL MEASUREMENTS</b>    |                       |  |  |
| <b>Mazzola et al. (2015)</b>                    | Not reported          | 9 PKU (7 males and 2 females; age 21 ± 4y)<br>17 controls (12 males, 5 females; aged 22 ± 4y)  | 2 days intervention.<br>Day 0: BMR test in fast state. VO2 peak test.<br>Minimum 1 week interval.<br>Day 1: Blood sampling (Moment 1 (M1), breakfast and   |

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30-min rest, aerobic exercise session (30 min at a prescribed VO<sub>2</sub>) and blood sampling (M2).

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Note: Authors in italics are abstract only papers. Not applicable: Not applicable, none interventional; PKU: phenylketonuria; HPA: hyperphenylalaninemia; Phe: phenylalanine; MUAC: Mid-upper arm circumference; UAMA: upper arm muscle area; ALMI: appendicular lean mass index; ALM: appendicular lean mass; PA: phase angle; BCM index: body cell mass index; NDR: nitrogen deposition rate; LBMDR: lean body mass deposition rate; MCA: muscle cross-sectional area; MM: muscle mass; LTM: lean tissue mass; TBW: total body water; FFMI: fat-free mass index; FMI: fat mass index; BUN: blood urine nitrogen; MF: medical food; PPS: Pretrial Protein Substitute; LPS: Liquid Protein Substitute.

<sup>a</sup> Płudowski P et al (2005) Reference values for the indicators of skeletal and muscular status of healthy polish children. *J Clin Densitom* 8:164–177

<sup>b</sup> Studies in the meta-analysis by Pena et al (2018) included participants from Denmark and USA.

<sup>c</sup> Study includes data from Pinto et al. (2017) but an extended follow-up period of 2.9 years if patients remained on CGMP-AA.

**Table 3: Protein status outcome measures**

| Reference              | Anthropometric |     |           |       |                             |                      |                     |      |                     | Biochemical |            |               |                         |                 |                     |                               |             |            |                    | Functional                                 |                     |               |                    |
|------------------------|----------------|-----|-----------|-------|-----------------------------|----------------------|---------------------|------|---------------------|-------------|------------|---------------|-------------------------|-----------------|---------------------|-------------------------------|-------------|------------|--------------------|--|---------------------|---------------|--------------------|
|                        | DEXA           | BIA | Skinfolds | TOBEC | Deuterium Isotopic Dilution | Total body potassium | Total body nitrogen | pQCT | Method not reported | Albumin     | Prealbumin | Total protein | Retinol-binding protein | Urea production | Blood urea nitrogen | Whole-body protein metabolism | Amino acids | Creatinine | Nitrogen excretion | <sup>13</sup> C-leucine in CO <sub>2</sub> | Method not reported | Grip strength | VO <sub>2max</sub> |
| Mazzola et al, 2016    |                | •   |           |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Adamczyk et al, 2011   | •              |     |           |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Weng et al, 2020       |                | •   |           |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Rocha et al, 2013      |                | •   |           |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Dobbelaere et al, 2003 |                | •   | •         |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Huemer et al, 2007     |                |     |           | •     |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Daly et al, 2021       | •              |     |           |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Nogueira et al, 2021   |                |     | •         |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Sailer et al, 2020     |                | •   |           |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Jani et al, 2017       | •              |     |           |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Stroup et al, 2018     | •              |     |           |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Evans et al, 2017      |                | •   |           |       |                             |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |
| Evans et al, 2018      |                | •   |           |       | •                           |                      |                     |      |                     |             |            |               |                         |                 |                     |                               |             |            |                    |  |                     |               |                    |

|                          |                |   |                |  |  |   |  |   |   |  |   |   |   |  |  |  |  |  |  |  |                |
|--------------------------|----------------|---|----------------|--|--|---|--|---|---|--|---|---|---|--|--|--|--|--|--|--|----------------|
| Allen et al, 1995        |                |   | •              |  |  |   |  |   |   |  |   |   |   |  |  |  |  |  |  |  |                |
| Bonifant et al, 2010     |                |   |                |  |  | • |  |   |   |  |   |   |   |  |  |  |  |  |  |  |                |
| Daly et al, 2019         | •              |   |                |  |  |   |  |   |   |  |   |   |   |  |  |  |  |  |  |  |                |
| Rocha et al, 2010        |                |   |                |  |  |   |  |   | • |  |   |   |   |  |  |  |  |  |  |  |                |
| Nalin et al, 2013        |                | • |                |  |  |   |  |   |   |  |   |   |   |  |  |  |  |  |  |  |                |
| Wilcox et al, 2011       | •              |   |                |  |  |   |  | • |   |  |   |   |   |  |  |  |  |  |  |  |                |
| Alfheaid et al, 2018     |                |   |                |  |  | • |  |   |   |  |   |   |   |  |  |  |  |  |  |  |                |
| Torriente et al, 2017    |                |   |                |  |  | • |  |   |   |  |   |   |   |  |  |  |  |  |  |  |                |
| Mexia et al, 2015        |                |   |                |  |  |   |  |   |   |  |   | • |   |  |  |  |  |  |  |  |                |
| Boros et al, 2015        |                |   | •              |  |  |   |  |   |   |  |   |   |   |  |  |  |  |  |  |  | •              |
| Paci et al, 2018         |                |   |                |  |  |   |  |   |   |  |   | • |   |  |  |  |  |  |  |  |                |
| Kanufre et al, 2015      |                |   |                |  |  | • |  |   |   |  |   |   |   |  |  |  |  |  |  |  |                |
| Allen et al, 1996        |                |   |                |  |  | • |  |   |   |  | • |   |   |  |  |  |  |  |  |  | •              |
| Doulgeraki et al, 2014   | •              |   |                |  |  |   |  |   |   |  |   | • |   |  |  |  |  |  |  |  | • <sup>a</sup> |
| Pena et al, 2021         |                |   | •              |  |  |   |  |   |   |  |   | • | • |  |  |  |  |  |  |  | •              |
| Hillman et al, 1996      | •              |   |                |  |  |   |  |   |   |  |   | • |   |  |  |  |  |  |  |  | •              |
| Lambruschini et al, 2005 |                |   |                |  |  | • |  |   |   |  |   | • |   |  |  |  |  |  |  |  |                |
| Pinto et al, 2017        |                |   | •              |  |  |   |  |   |   |  |   | • | • |  |  |  |  |  |  |  |                |
| Modan-Moses et al, 2007  | • <sup>a</sup> |   |                |  |  |   |  |   |   |  |   | • |   |  |  |  |  |  |  |  | •              |
| Das et al, 2010          |                |   | • <sup>b</sup> |  |  |   |  |   |   |  |   |   |   |  |  |  |  |  |  |  | •              |



|                              |  |                |  |  |  |  |  |  |   |                |   |   |   |   |  |   |   |   |                |  |  |   |
|------------------------------|--|----------------|--|--|--|--|--|--|---|----------------|---|---|---|---|--|---|---|---|----------------|--|--|---|
| <i>Sumanszki et al, 2019</i> |  | •              |  |  |  |  |  |  |   |                | • |   |   |   |  |   |   |   |                |  |  |   |
| Das et al, 2013              |  | • <sup>a</sup> |  |  |  |  |  |  |   | • <sup>a</sup> |   | • |   | • |  |   |   | • | • <sup>a</sup> |  |  |   |
| Choukair et al, 2017         |  |                |  |  |  |  |  |  | • |                |   |   |   |   |  |   |   |   |                |  |  | • |
| Sumanszki et al, 2020        |  | •              |  |  |  |  |  |  |   |                |   |   |   |   |  |   |   |   |                |  |  | • |
| Singh et al, 2010            |  |                |  |  |  |  |  |  |   | •              | • | • |   |   |  |   |   |   |                |  |  |   |
| van Vliet et al, 2019        |  |                |  |  |  |  |  |  |   | •              | • | • |   |   |  |   |   |   |                |  |  |   |
| van Calcar et al, 2009       |  |                |  |  |  |  |  |  |   | •              | • | • |   |   |  | • |   |   |                |  |  |   |
| Andrade et al, 2017          |  |                |  |  |  |  |  |  |   |                | • | • |   |   |  |   |   |   |                |  |  |   |
| Arnold et al, 2001           |  |                |  |  |  |  |  |  |   | •              | • | • |   |   |  |   |   | • |                |  |  |   |
| Gokmen-Ozel et al, 2009      |  |                |  |  |  |  |  |  |   | •              |   |   |   |   |  |   |   |   |                |  |  |   |
| Viau et al, 2021             |  |                |  |  |  |  |  |  |   | •              | • |   |   |   |  |   |   | • |                |  |  |   |
| Shenton et al, 1983          |  |                |  |  |  |  |  |  |   | •              | • |   |   |   |  |   |   |   |                |  |  |   |
| Arnold et al, 2002           |  |                |  |  |  |  |  |  |   |                | • |   |   |   |  |   |   |   |                |  |  |   |
| van Rijn et al, 2007         |  |                |  |  |  |  |  |  |   | •              |   | • |   |   |  | • | • |   |                |  |  |   |
| Thompson et al, 1990         |  |                |  |  |  |  |  |  |   |                |   |   |   |   |  | • | • |   |                |  |  |   |
| Acosta et al, 1999           |  |                |  |  |  |  |  |  |   | •              | • |   | • |   |  | • |   |   |                |  |  |   |
| Giovannini et al, 2014       |  |                |  |  |  |  |  |  |   | •              | • |   | • |   |  |   |   | • |                |  |  |   |
| Schulpis et al, 2013         |  |                |  |  |  |  |  |  |   | •              |   | • |   |   |  |   |   |   |                |  |  |   |
| Rocha et al, 2010            |  |                |  |  |  |  |  |  |   |                | • |   |   |   |  |   |   |   |                |  |  |   |
| Prince et al, 1997           |  |                |  |  |  |  |  |  |   |                | • |   |   |   |  |   |   | • |                |  |  |   |

|  |   |    |   |   |   |   |   |   |   |    |    |    |   |   |    |   |    |   |   |   |   |   |   |   |
|--|---|----|---|---|---|---|---|---|---|----|----|----|---|---|----|---|----|---|---|---|---|---|---|---|
| Mönch et al, 1996                            |   |    |   |   |   |   |   |   |   |    |    |    |   |   |    |   |    |   | • | • |   |   |   |   |
| Crujeiras et al, 2015                        |   |    |   |   |   |   |   |   |   |    | •  | •  |   |   |    |   |    |   |   |   |   |   |   |   |
| Kose et al, 2019                             |   |    |   |   |   |   |   |   |   | •  | •  | •  |   |   |    |   |    |   |   |   |   |   |   |   |
| Nord et al, 1988                             |   |    |   |   |   |   |   |   |   | •  |    | •  |   |   |    |   |    | • |   |   |   |   |   |   |
| <i>Kose et al, 2016</i>                      |   |    |   |   |   |   |   |   |   | •  | •  |    |   |   |    |   |    |   |   |   |   |   |   |   |
| <i>Prochazkova et al, 2012</i>               |   |    |   |   |   |   |   |   |   |    | •  |    |   |   |    |   |    |   |   |   |   |   |   |   |
| <i>Giovannini et al, 2006</i>                |   |    |   |   |   |   |   |   |   | •  |    | •  |   |   |    |   |    | • |   |   |   |   |   |   |
| <i>Desloovere et al, 2014</i>                |   |    |   |   |   |   |   |   |   |    | •  |    |   |   |    |   |    | • |   |   |   |   |   |   |
| <i>Giovannini et al, 2009</i>                |   |    |   |   |   |   |   |   |   | •  | •  |    | • |   |    |   |    | • |   |   |   |   |   |   |
| <i>Graffin et al, 1995</i>                   |   |    |   |   |   |   |   |   |   | •  |    | •  |   |   |    |   | •  |   |   |   |   |   |   |   |
| <i>Rocha et al, 2009</i>                     |   |    |   |   |   |   |   |   |   |    | •  |    |   |   |    |   |    |   |   |   |   |   |   |   |
| <i>Douglas et al, 2013</i>                   |   |    |   |   |   |   |   |   |   | •  |    | •  |   |   |    |   | •  |   |   | • |   |   |   |   |
| Ahring et al, 2018                           |   |    |   |   |   |   |   |   |   |    |    |    |   |   |    |   | •  |   | • |   |   |   |   |   |
| Pena et al, 2018                             |   |    |   |   |   |   |   |   |   |    |    |    |   |   |    |   | •  |   |   |   |   |   |   |   |
| Zaki et al, 2016                             |   |    |   |   |   |   |   |   |   | •  |    |    |   |   |    |   | •  |   | • |   |   |   |   |   |
| Ney et al, 2016                              |   |    |   |   |   |   |   |   |   | •  | •  | •  |   |   |    |   | •  |   | • |   |   |   |   |   |
| Mazzola et al, 2015                          |   |    |   |   |   |   |   |   |   |    |    |    |   |   |    |   |    | • |   |   |   |   |   | • |
| <b>Total of measures included in studies</b> | 9 | 15 | 7 | 1 | 2 | 1 | 2 | 1 | 3 | 27 | 23 | 17 | 3 | 2 | 10 | 2 | 18 | 4 | 1 | 1 | 1 | 1 | 2 |   |

Authors in italics are abstract only papers; DEXA: Dual-energy X-ray absorptiometry; BIA: Bioelectrical impedance analysis; TOBEC: total body electrical conductivity; IVNAA: Gamma neutron activation analysis; pQCT: Peripheral quantitative computed tomography

<sup>a</sup> Included in methods, but results not reported

<sup>b</sup> Reports body composition measured by BIA, but unclear if FM or FFM measured

**Table 3: Anthropometric measurements of protein status and key findings**

| Author (Year)              | Method used       | Modulating factors         | Key findings: Protein status outcomes   | Variable  | Group 1  | Group 2  | Group 3   |
|----------------------------|-------------------|----------------------------|---|---|--|--|---|
| <b>Daly et al. (2021)</b>  | DEXA              | Type of protein substitute | No significant differences in LBM between the Tx groups, although a trend for improved LBM was observed in the CGMP100 group. All body composition parameters increased over 3 years. | Lean mass at baseline (kg)<br>Lean mass at end of study (kg)<br>Lean mass change (kg) | <b>AA (n=19):</b><br>26.7 (16.9-34.2)<br>32.6 (5.9-40.5)<br>+5.9 (9.0-6.3) | <b>GMP50 (n=13):</b><br>16.3 (14.2-17.7)<br>23.9 (22.7-26.5)<br>+7.6 (8.4-8.8) | <b>GMP100 (n=16):</b><br>20.1 (16.5-21.9)<br>31.3 (25.6-35.9)<br>+11.2 (9.1-13.9)   |
| <b>Daly et al. (2019)</b>  | DEXA              | Type of protein substitute | FFM not significantly different between groups at baseline. At 36 mo, FFM was significantly greater in the CGMP-AA than for LAA group (p = 0.01)                                      | FFM (kg) at 36 months   | <b>CGMP-AA:</b><br>9.05  | <b>L-AA:</b><br>6.47   |   |
| <b>Evans et al. (2017)</b> | BIA               | BH4, dietary parameters    | %FFM was only measured at baseline. No significant difference in %FFM between groups (p=0.148 All-PKU vs Controls)<br>Neither %EBMR nor P:E ratio contributed significantly to %FFM   | %FFM at baseline (range)  | <b>Diet-PKU:</b><br>84.1 ± 7.4 (64.1-92.1)                                 | <b>BH4-PKU:</b><br>83.5 ± 3.1 (80.8-87.5)                                      | <b>All PKU:</b><br>84 ± 7 (64.0-92.1)<br><b>Controls:</b><br>80.9 ± 4.2 (71.8-86.2) |
| <b>Paci et al. (2018)</b>  | Method not stated | None assessed              | Total body water and FFM (kg) were significantly higher in PKU vs. controls.  | No data shown   |  |  |   |

|                                |   |                    |   |   |   |  |
|--------------------------------|---|--------------------|---|---|---|--|
| <b>Boros et al.</b><br>(2015)  | BIA   | None assessed      | Decreased SMM were found in 2 patients (7%). In 7 patients (26%), SMM was higher than matching normal reference values.   |   |   |  |
| <b>Rocha et al.</b><br>(2013)  | BIA   | None assessed      | No significant differences were found between patients and controls.<br><br>Data as median [25th and 75th percentiles]  | %FFM<br><br>%Muscular mass<br><br>PA                      | PKU:<br>78.0 [71.1–85.7]<br>48.3 [43.0–53.3]<br>5.9 [5.2–6.5] | Controls:<br>77.0 [71.2–83.7]<br>46.7 [43.5–53.2]<br>6.0 [5.5–6.6] |
| <b>Huemer et al.</b><br>(2007) | TOBEC   | Dietary parameters | <u>Cross sectional</u> : No significant difference in FFM between PKU and age- sex-matched controls.<br><br><u>Longitudinal</u> : %FFM remained unchanged over time<br><br>Natural protein intake (g/kg per day) explained 47.7% of the variance of FFM (R2 = 0.477; p < 0.0001). Total protein intake (g/kg per day) and natural or total protein intake (g/day) without reference to body weight explained 8.8%, 7% and 26.1%, respectively of the variance of FFM. | %FFM at baseline<br><br>%FFM at 6 mo<br><br>%FFM at 12 mo | PKU:<br>84.6 ± 16.9<br>83.6 ± 15.8<br>85.5 ± 13.8             | Controls:<br>85.2 ± 9.7  |
| <b>Allen et al.</b><br>(1996)  | Skinfolds<br><br>PGNA for total body nitrogen | Sex                | <u>Cross Sectional Data</u><br><br>No diff in LBM between PKU and controls.<br><br>Sig diff in TBN between PKU and controls (p<0.005)<br><br>TBN lower in females (data not shown)  | LBM (kg) (mean ± SD)<br><br>TBN (g) (mean ± SD)           | PKU:<br>19.7 ± 4.5<br><br>575 ± 200                           | Controls:<br>22.0 ± 4.8<br><br>710 ± 215                           |

|  |  |  |  |  |   |
|--|--|--|--|--|---|
| <p>UAMA were derived from the MUAC and skinfolds</p> | <p>PKU: had 35g less TBN than controls for same LBM. At each age children with PKU had 53g less TBN than controls, representing 6mo lag in PKU vs. controls</p> <p>Control: TBN significantly correlated LBM, weight, height and age (<math>r = 0.97, 0.95, 0.88</math>, respectively, <math>p &lt; 0.001</math>).</p> <p>UAMA similar between PKU and controls (data not shown). UAMA significantly correlated with TBN in both PKU and controls (<math>r_s = 0.84, r_s = 0.89, P &lt; 0.001</math>, respectively).</p> <p><u>Longitudinal Data</u> (n=29 PKU, n=17 controls)</p> <p>Similar increase in TBN and LBM in both groups between the two time periods</p> <p>Annual accretion of nitrogen, NDR and LBMR: Similar between groups.</p> <p>NDR was significantly correlated with the LBMDR in the control but not in PKU.</p> | <p>Annual accretion of nitrogen (g/y)</p> <p>NDR (g/y)</p> <p>Cross-sectional</p> <p>Longitudinal</p> <p>LBMDR (kg/y)</p> <p>Cross-sectional</p> <p>Longitudinal</p> | <p><math>86 \pm 45</math></p> <p>93</p> <p><math>86.1 \pm 45.1</math></p> <p>2.1</p> <p><math>1.8 \pm 0.6</math></p> | <p><math>77 \pm 58</math></p> <p>98</p> <p><math>77 \pm 57.9</math></p> <p>2.3</p> <p><math>2.2 \pm 0.9</math></p> |   |
| <p>Sailer et al (2020)</p>                           | <p>BIA Sex, dietary parameters</p>   | <p>No difference in LBM% in females (<math>p = 0.34</math>), but males with PKU had significantly lower LBM% (<math>p = 0.02</math>) than male controls.</p>         | <p>Lean body mass % males</p>  | <p><b>PKU:</b><br/><math>85.38 \pm 4.9</math></p>  | <p><b>Controls:</b><br/><math>89.09 \pm 5.94</math></p> |

|                                    |   |                       |   |                                    |   |   |
|------------------------------------|---|-----------------------|---|------------------------------------|---|---|
|                                    |   |                       |   | Lean body mass %<br>females        | 77.87 ± 9                                     | 76.50 ± 9.87                                |
| <b>Weng et al.<br/>(2020)</b>      | BIA   | Dietary<br>parameters | Muscle mass %: No significant difference between PKU and controls (p = 0.37), and between PKU patients on phe-free formulas and those taking no PS (p=0.95), although natural protein intake was significantly different.<br><br>Significant positive correlation between total protein intake % of DRIs and muscle mass (r = 0.491, p= 0.020) in PKU.<br><br>% natural protein had no correlation to muscle mass (r = - 0.007, p = 0.974). | Muscle mass %<br><br>Muscle mass % | <b>PKU:</b><br><br><b>PKU + PS:</b>           | <b>Controls:</b><br><br><b>PKU + no PS:</b> |
|                                    |   |                       |   |                                    | 73.39 ± 8.79                                  | 75.51 ± 7.42                                |
|                                    |   |                       |   |                                    | 73.15 ± 8.5                                   | 73.68 ± 9.56                                |
| <b>Sumanszki<br/>et al. (2019)</b> | BIA   | Sex                   | Females: both lumbar spine and femoral BMD correlated positively with LBM (r = 0.53; p< 0.001, r = -0.31; p = 0.042, respectively). Lean mass had a greater effect on BMD than fat mass. Observations not reported in males   |                                    |   |   |
| <b>Evans et al.<br/>(2018)</b>     | BIA<br><br>TBW by<br>deuterium<br>dilution. | None assessed         | No difference between FFM measured by Deut and BIA (p = 0.111)<br><br>Correlation analysis showed that FFM by BIA correlated significantly with FFM by TBWDeut (r = 0.984, p < 0.0001)  | FFM (kg):<br><br>Mean (SD)         | <b>FFM by Deut:</b><br><br><b>FFM by BIA:</b> | <b>FFM by BIA:</b>                          |
|                                    |   |                       |   |                                    | 31.81 (± 12.77)                               | 32.93 (± 13.93)                             |

|  |                           |   |   |                    |                        |                         |
|--|---------------------------|---|---|--------------------|------------------------|-------------------------|
| <b>Stroup et al.</b><br>(2018)                                   | DEXA                      | Sex, genotype   | Males had significantly more lean mass ( $p = 0.0008$ ) and       |                    | <b>Male:</b>           | <b>Female:</b>          |
|  |                           |   | ALM ( $p=0.0002$ ) compared to females. No significant            | Total Lean mass kg | $55.3 \pm 2.8$         | $41.9 \pm 1.4$          |
|  |                           |   | difference in the ALMI (and ALMI z-scores) between                | (mean $\pm$ SE)    |                        |                         |
|  |                           |   | males and females. Mean ALMI for our young male                   | ALM kg             | $24.8 \pm 1.5$         | $18.0 \pm 0.8$          |
| <b>Choukair et al.</b><br>(2017)                                 | pQCT to<br>measure<br>MCA | Sex,<br>classification<br>of PKU,<br>metabolic<br>control | Mean MCA was decreased compared with the ref.                     |                    |                        |                         |
|  |                           |   | population (z-score $-0.98 \pm 1.19$ ; $p < 0.0001$ ); observed   |                    |                        |                         |
|  |                           |   | both in females and males. 30% had MCA $<3$ rd                    |                    |                        |                         |
|  |                           |   | percentile.   |                    |                        |                         |
|  |                           |   | No relationship between MCA (z-scores) and PKU type               |                    |                        |                         |
|  |                           |   | or mean phe concentrations. Bone strength were                    |                    |                        |                         |
|  |                           |   | significantly correlated to MCA. In PKU, the regression           |                    |                        |                         |
|  |                           |   | line slope between SSI and MCA was significantly ( $p <$          |                    |                        |                         |
|  |                           |   | $0.0001$ ) less steep than in the reference population.           |                    |                        |                         |
| <b>Jani et al.</b><br>(2017)                                     | DEXA                      | Dietary<br>parameters,<br>genotype                        | In adults ( $n=17$ ), high intact protein intake was associated   | Median(min,max)    | <b>Males (n = 9):</b>  | <b>Females (n =17):</b> |
|  |                           |   | with high FFMI ( $r_s = 0.75$ , $p = 0.008$ ) and low FMI:FFMI    | <b>Adults</b>      | $52.2 (45.5, 61.8)$    | $38.9 (30.8, 64.3)$     |
|  |                           |   | ( $r_s = -0.59$ , $p = 0.04$ ).                                   | Lean mass (kg)     |                        |                         |
|  |                           |   | In children, protein substitute ( $r_s = 0.38$ , $p = 0.04$ ) and | <b>Children</b>    | <b>Males (n = 25):</b> | <b>Females (n=32):</b>  |
| total protein intake ( $r_s = 0.39$ , $p = 0.04$ ) were directly | Lean mass (kg)            | $25.7 (13.3, 64.4)$                                       | $32.7 (15.5, 50.7)$   |                    |                        |                         |



|                                    |                   |   | associated with FFMI.   | FFMI                     | <b>Total (n=83):</b>  | <b>Adults (n=26):</b>  | <b>Children (n=57):</b> |
|------------------------------------|-------------------|---|---|--------------------------|---|--|-------------------------|
|                                    |                   |   | Genotype was not associated with body composition.  |                          | 14.8 (11.3, 24.2)   | 17.4 (13.4, 24.2)  | 14.2 (11.3, 23.7)       |
| <b>Torriente et al.</b><br>(2017)  | Skinfolds         | None assessed                               | Muscle area: normal 100%  |                          |   |  |                         |
| <b>Mazzola et al.</b><br>(2016)    | BIA               | None assessed                               | No differences in FFM, ECM/BCM ratio, and PA between PKU patients and controls.<br>3 patients with PKU and 3 controls were below the cut-off values for PA  | FFM %                    | <b>PKU:</b><br>80 ± 7   | <b>Controls:</b><br>78 ± 9   |                         |
| <b>Kanufre et al.</b><br>(2015)    | Skinfolds         | None assessed                               | 32 (41.5%) had muscle mass deficit of which 28 (87.5%) were normal weight-for-age and 25 (78%) were adolescents   |                          |   |  |                         |
| <b>Mexia et al.</b><br>(2015)      | Method not stated | None assessed                               | Deficit of FFMI was found in 30.8% of patients  |                          |   |  |                         |
| <b>Doulgeraki et al.</b><br>(2014) | DEXA              | Sex, pubertal status, classification of PKU | No difference in LBM in PKU and controls. No effect for correction for height.<br>MM of mHPA were comparable to controls. No significant difference was detected in body composition parameters between patient with PKU and mHPA.<br>No effect of gender on body composition Pubertal status associated with increased in LBM in adolescents (pubertal) in mHPA (p<0.01) but not in PKU. | LBM z-scores (mean ± SD) | <b>mHPA pre-pubertal:</b><br>-0.65 ± 1.5<br><br><b>PKU pre-pubertal:</b><br>-0.23 ± 1.1 | <b>mHPA pubertal:</b><br>0.7 ± 1.1<br><br><b>PKU pubertal:</b><br>-0.1 ± 1.3 |                         |

PKU: a significant positive correlation between BMD and MM.

|                                 |                              |  |   |  |   |  |
|---------------------------------|------------------------------|--|---|--|---|--|
| <b>Dobbelaere et al. (2003)</b> | Skinfold thickness<br>BIA    | None assessed                            | No difference between PKU and controls.   | FFM (kg) via skinfold<br>FFM (kg) via BIA          | <b>PKU:</b><br>12.4±3.2<br>14.1±1.4         | <b>Controls:</b><br>12.8±2.1<br>12.9±2.3 |
| <b>Nogueira et al. (2021)</b>   | Skinfolds<br>Arm muscle area | Metabolic control, socio-economic status | Association between metabolic control and AMA (linear trend chi square; p=0.042). AMA classified as above average or adequate was associated with worse % of metabolic control.<br><br>No association between AMA with SES. | Metabolic control with ≥70% adequate<br>Phe levels | <b>Low AMA:</b><br>70%                      | <b>High AMA:</b><br>18.5%                |
| <b>Pena et al. (2021)</b>       | BIA                          | Type of protein substitute               | No difference in LM% and PA for all patients taking L-AA compared with CGMP-AA  | LM (%) n=9<br>PA (°) n=9                           | <b>CGMP-AA:</b><br>71.1 ± 13.4<br>6.8 ± 0.6 | <b>L-AA:</b><br>74.5 ± 16.1<br>6.8 ± 0.7 |
| <b>Pinto et al. (2017)</b>      | BIA                          | Type of protein substitute               | All parameters remained unchanged   | LM (%) (n =9)<br>PA (°) (n =9)                     | <b>CGMP-AA:</b><br>71.4±15.0<br>6.7±0.7     | <b>L-AA:</b><br>74.5±16.1<br>6.8 ±0.7    |
| <b>Sumanszki et al. (2020)</b>  | BIA                          | None assessed                            | No difference in body composition parameters in PKU and controls (p = 0.497)  | FFM (%)  | <b>PKU:</b><br>47.5 (42.1–49.3)             | <b>Controls:</b><br>47.3 (45.3–48.5)     |

|                                  |                              |                                    |  |   |  |   |
|----------------------------------|------------------------------|------------------------------------|--|---|--|---|
| <i>Alfheaid et al.</i><br>(2018) | Deuterium dilution technique | None assessed                      | No difference in FFM between PKU vs. controls.   | Data not shown.   |  |   |
| <b>Das et al.</b><br>(2013)      | BIA                          | Dietary parameters                 | LBM/FFM: No results reported.<br>PA: Normal in all dietary groups, no significant difference between groups.<br>BCM increased in vegan + L-AA and reduced in vegan patients compared to PKU-diet patients (non-significant).                           | No data shown   |  |   |
| <i>Nalin et al.</i><br>(2013)    | BIA                          | Metabolic controls                 | % FFM and PA: No difference in PKU vs. controls. A positive correlation between PA and phenylalanine levels in PKU ( $r=0.457$ , $p=0.032$ )   | <b>PKU:</b><br>% FFM<br>PA (°):   | 80.9±7.7<br>6.35   | <b>Controls:</b><br>80.7±7.3<br>6.89  |
| <b>Adamczyk et al.</b><br>(2011) | DEXA                         | Pubertal status, metabolic control | No significant differences between LBM in 2a and 2b (both pubertal).<br>Increased LBM for body height SD scores in adolescents with normal Phe levels.<br>LBM Z-scores: Sign. differences between 2a and 2b. No difference between subgroups 2a and 1. | <b>Group 1:</b><br>prepubertal/<br>normal phe<br>LBM (g)<br>LBM SD scores<br>LBM Z-scores | <b>Group 2a:</b> pubertal/<br>normal phe<br>Data not shown<br>+1.94 ± 3.21<br>+0.51 ± 1.59 | <b>Group 2b:</b><br>pubertal/high phe<br>Data not shown<br>-0.20 ± 1.52<br>-0.69 ± 0.71 |
| <i>Bonifant et al.</i><br>(2010) | TBK                          | None assessed                      | TBK z score was significantly lower ( $p < 0.05$ ) in patients with PKU  |   |  |   |

|                                   |                                   |                    |  |   |   |   |
|-----------------------------------|-----------------------------------|--------------------|--|---|---|---|
| <i>Das et al.</i><br>(2010)       | BIA                               | Dietary parameters | Body composition within target levels  | No data shown                                 |   |   |
| <i>Rocha et al.</i><br>(2010)     | Method not stated                 | BMI                | Patients classified as overweight and obesity had lower FFM% (p<0.001)   | FFM%  | <b>Overweight/obese:</b> 63.7                 | <b>Normal weight:</b> 78.3                  |
| <i>Wilcox et al.</i><br>(2011)    | IVANA for total body protein DEXA | None assessed      | SMM was measured as height-adjusted ALTM; TBP measured as age and sex adjusted nitrogen index (NI) SMM and TBP were normal PKU patients                    | ALTM (kg/m <sup>2</sup> ):<br>Nitrogen Index: | <b>Females:</b><br>5.77 ± 2.35<br>0.98 ± 0.12 | <b>Males:</b><br>8.24 ± 0.45<br>1.13 ± 0.13 |
| <b>Modan-Moses et al.</b> (2007)  | DEXA                              | None assessed      | No data reported for lean body mass or FFM.  | No data shown                                 |   |   |
| <b>Lambruschini et al.</b> (2005) | Skinfolds: Brachial muscular area | BH4                | All values within age- and sex-specific percentiles for a healthy population after 1 yr treatment. No difference in brachial muscular area (BMA).          | BMA (mm <sup>2</sup> ) (range)                | <b>Before BH4:</b><br>984-4505                | <b>After BH4:</b><br>1167-4819              |
| <b>Hillman et al.</b> (1996)      | DEXA                              | None assessed      | No difference between PKU and controls (p = 0.53).   | Lean body weight (kg)                         | <b>PKU:</b><br>27.98 ± 12.43                  | <b>Controls:</b><br>30.71 ± 17.47           |
| <b>Allen et al.</b> (1995)        | Skinfolds                         | Sex                | No difference in FFM between males with PKU and control participants. Females with PKU were younger than controls (P=0.006) and had a lower FFM (p=0.007). | FFM (kg) males<br>FFM (kg) females            | <b>PKU:</b><br>27.2 ± 7.6<br>23.1 ± 8.1       | <b>Controls:</b><br>27.1 ± 7<br>30.9 ± 8.2  |

Note: Authors in italics are abstract only papers

PKU: phenylketonuria; HPA: hyperphenylalaninemia; Phe: phenylalanine; DEXA: Dual-energy X-ray absorptiometry; BIA: Bioelectrical impedance analysis; PGNA: Prompt gamma neutron capture analysis; pQCT: Peripheral quantitative computed tomography; IVANA: Gamma neutron activation analysis; LBM: lean body mass; LM%: lean mass percentage; FFM: fat-free mass; TBW: total body water; FFMI: fat-free mass index; FMI: fat mass index; PA: phase angle; BCM: body cell mass; ECM: extracellular mass; MUAC: Mid-upper arm circumference; UAMA: upper arm muscle area; ALMI: appendicular lean mass index; ALM: appendicular lean mass; NDR: nitrogen deposition rate; LBMDR: lean body mass deposition rate; MCA: muscle cross-sectional area; MM: muscle mass; SMM: skeletal muscle mass; SSI: Strength-Strain Index; TBP: Total body protein; TBK: Total body potassium; BUN: blood urine nitrogen; PAA: plasma amino acids; L-AA: L-amino acids; cGMP-AA: casein glycomacropeptide amino acid; PS: protein substitute

**Table 5: Biochemical measurements of protein status and key findings**

| Author (Year)     | Method used                                     | Modulating factors         | Key findings: Protein status outcomes   | Variable  | Group 1  | Group 2   | Group 3 | Group 4 |
|-------------------|---|----------------------------|---|---|--|---|---------|---------|
| Ney et al. (2016) | PAA, BUN, albumin, prealbumin and total protein | Type of protein substitute | No sig. differences between BUN, total protein and prealbumin for L-AA compared with cGMP-AA. Albumin was sig. higher with cGMP-AA than L-AA ( $p = 0.027$ ). Mean values were within the normal range for both diets. PAA: Except for Phe, all mean conc of AAs were within the normal range for both groups. Thr (mean $\pm$ SE) showed sig. increase with cGMP-AA from $103 \pm 4$ mmol/L to $149 \pm 10$ mmol/L ( $p < 0.001$ ). L-AA diet: sig. change from baseline to 3 weeks for Arg (increased), Leu (increased), Phe (reduced) cGMP-AA diet: sig. change from baseline to 3 weeks for Gly (reduced), Lys (reduced), Met (increased), Thr (increased), Val (reduced) | Blood urea nitrogen, mg/dL<br>Total protein, g/dL<br>Albumin, g/dL<br>Prealbumin, mg/dL | <b>L-AA:</b><br>$11.2 \pm 0.6$<br>$7.35 \pm 0.08$<br>$4.24 \pm 0.04$<br>$27 \pm 1.0$ | <b>cGMP-AA:</b><br>$10.6 \pm 0.6$<br>$7.42 \pm 0.07$<br>$4.35 \pm 0.04$<br>$27 \pm 1.1$ |         |         |

| <b>Giovannini et al. (2014)</b> | Albumin, prealbumin, RBP, PAA | Type of protein substitute | Baseline: PKU had lower albumin, prealbumin, and higher AA ratio than MHP (maximum p = 0.01) and unaffected children (maximum p < 0.001). Albumin was within the reference range for all children. Children with MHP and unaffected children: prealbumin within reference range, n=2 PKU had prealbumin levels below ref range. 29 (52.7%) PKU had prealbumin <20 mg/dL. The AA ratio ranged from 1.7-3.5, 1.5- 3.0, 1.5-2.5 in PKU and MHP and unaffected children, respectively.<br><br>End of study: PKU has albumin and prealbumin within range, except n=1 receiving the conventional substitute (prealbumin 18.3 mg/dL). No overall sig. difference between PKU groups was found for protein status.<br><br>Within-group analysis: sign. increase of prealbumin in children who received the test substitute (p = 0.017). The change in albumin was close to statistical sig. in the test group (p = 0.068). | Mean ± SD             | <b>Prolonged-release substitute (n=27)</b> | <b>Conventional substitute (n=28)</b> |
|---------------------------------|-------------------------------|----------------------------|--|-----------------------|--|---------------------------------------|
|                                 |                               |                            |  | Albumin g/dL          |  |                                       |
|                                 |                               |                            |  | Baseline              | 4.5 ± 0.2                                  | 4.5 ± 0.2                             |
|                                 |                               |                            |  | Albumin g/dL          | 4.6 ± 0.2                                  | 4.5 ± 0.2                             |
|                                 |                               |                            |  | End of study          |  |                                       |
|                                 |                               |                            |  | Transthyretin (mg/dL) | 19.1 ± 6.4                                 | 19.0 ± 6.3                            |
|                                 |                               |                            |  | Baseline              |  |                                       |
|                                 |                               |                            |  | Transthyretin (mg/dL) | 20.7 ± 6.8                                 | 19.2 ± 6.0                            |
|                                 |                               |                            |  | End of study          |  |                                       |
|                                 |                               |                            |  | AA ratio              | 2.6 ± 0.9                                  | 2.6 ± 0.8                             |
|                                 |                               |                            |  | Baseline              |  |                                       |
|                                 |                               |                            |  | AA ratio              | 2.7 ± 0.8                                  | 2.6 ± 0.8                             |
|                                 |                               |                            |  | End of study          |  |                                       |

|                             |                 |                            |  |   |  |   |  |  |  |
|-----------------------------|-----------------|----------------------------|--|---|--|---|--|--|--|
| <b>Prince et al. (1997)</b> | Prealbumin, EAA | None assessed              | Phase 1: Baseline: All EAA (except phe) uniformly low compared with controls. Mean EAA remained low compared to control. No sig diff in serum EAA between control and experimental groups at entry or end.<br><br>Phase 2: Despite reductions in protein substitute intakes, mean serum protein levels were not sig. different and remained in the healthy (non-PKU) age-matched reference range throughout the study. | Mean ( $\pm$ SD),<br>Prealbumin,<br>mg/dL | <b>Baseline:</b><br>17.3 $\pm$ 2         | <b>End of study:</b><br>21.0 $\pm$ 4    | <b>Controls (range):</b><br>17– 42       |  |  |
| <b>Ahring et al. (2018)</b> | BUN, PAA        | Type of protein substitute | AUC (adjusted for baseline) for total AA: No differences between DM1 and DM2 (p = 0.852), or between DM3 and DM4 (p= 0.06). Significant differences for AUC for some individual AA: DM1 and DM2: Lys (p = 0.0287), Asn (p = 0.0210), and Asp (p =0.0047) and DM3 and DM4: citrulline (p = 0.0162).<br><br>BUN: no sig change from baseline to 240 min after meal and DM.   | Peak AA concentration<br><br>ns           | <b>DM1:</b> after 30 min for 90% of AAs. | <b>DM2:</b> after 15 min for 71% of AAs | <b>DM3:</b> after 30 min for 67% of AAs. | <b>DM4:</b> after 15 min for 71% of AA |  |
| <b>Zaki et al. (2016)</b>   | Amino acids,    | Type of protein substitute | Phase I vs Phase II: Individual amino acids were generally lower in phase I, with sig. low level of aspartic acid and citrulline. Levels of all amino acids in phases I and II were  | Mean (SD)<br>(min - max)<br><br>BUN:      | <b>Baseline</b>                          | <b>Phase I</b>                          | <b>Phase II</b>                          |  |  |



|                           |                                       |  |  |  |   |  |                         |
|---------------------------|---------------------------------------|--|--|--|---|--|-------------------------|
|                           | albumin,                              |  | not significantly different.   |  | 20.6 (7.66),                                  | 17.63  | 19.67                   |
|                           | BUN                                   |  | BUN and albumin: no sig. difference between phases of the study.   |  | (6-30)  | (5.18), (8-23)                               | (9.75), (10-37)         |
|                           |                                       |  |  | Albumin:                                   | 4.6 (0.35), (4.2-5.2)                         | 4.5 (0.52), (4.0-5.3)                        | 4.52 (0.31), (4.0- 4.8) |
| <b>Kose et al. (2019)</b> | Prealbumin, albumin and total protein | Dietary adherence, metabolic control and age | Prealbumin was sig. higher in PKU compared to controls (p =0.013). Frequency prealbumin above the ref range was higher in PKU than the controls (p=0.02). Albumin and total protein were not sig. different between PKU and controls. Prealbumin was sig. lower in those with high adherence to diet compared to those with low adherence (p = 0.011). Positive correlation between plasma phe level and prealbumin (r = 0.256, p = 0.003) and between age and prealbumin in PKU (r = 0.556, p < 0.0001) and control groups (r = 0.682, p < 0.0001). | Serum prealbumin, mg/dL (Ref range: 21-41) | <b>PKU (n=112):</b><br>24.1 ± 4.6 (10.5-35.5) | <b>Con (n=36):</b><br>21.9 ± 3.9 (15.9-29.8) |                         |
|                           |                                       |  |  | Prealbumin, (<21 mg/dL), n%                | 21 (21.1)                                     | 14 (38.8)                                    |                         |
|                           |                                       |  |  | Serum prealbumin, mg/dL (Ref range: 21-41) | <b>High dietary adherence:</b> 22.5 ± 4.4     | <b>Low dietary adherence:</b> 24.9 ± 4.6     |                         |

|                                  |                              |                   |   |                                      |   |   |
|----------------------------------|------------------------------|-------------------|---|--------------------------------------|---|---|
| <i>Boros et al.</i><br>(2015)    | Method not reported          | None assessed     | Decreased protein levels in 2 found to have low SMM. Normal protein levels in 7 with SMM above normal ref values.   | No data provided                     |   |   |
| <i>Prochazkova et al.</i> (2012) | Prealbumin                   | None assessed     | No sig. difference in the levels of serum prealbumin among the respective groups.   | No data provided                     |   |   |
| <i>Allen et al.</i> (1996)       | PAA                          |                   | No difference in PAA between PKU and controls, except for phe.  | No data provided                     |   |   |
| <i>Viau et al.</i> (2021)        | EAA, prealbumin and albumin  | Blood phe levels  | No deficiencies were identified in EAA, prealbumin, or albumin. No sig differences in protein intake or prealbumin in patients with blood Phe <30 $\mu\text{mol/L}$ vs. $\geq 30$ $\mu\text{mol/L}$ . | Prealbumin, mg/dL                    | <b>Phe &lt; 30 <math>\mu\text{mol/L}</math></b><br><b>(n=11):</b><br>27.0 $\pm$ 1.7 | <b>Phe <math>\geq</math> 30 <math>\mu\text{mol/L}</math></b><br><b>(n=7):</b><br>30.6 $\pm$ 1.1 |
| <i>Sumanszki et al.</i> (2019)   | Prealbumin                   | Sex               | Pre-albumin levels were higher in males compared to females with PKU (p = 0.008)  | Prealbumin, mg/dL<br>(mean $\pm$ SD) | <b>Females:</b><br>28 $\pm$ 4.1   | <b>Males:</b> 30.5 $\pm$ 3.8  |
| <i>Andrade et al.</i> (2017)     | Prealbumin and total protein | Metabolic control | Prealbumin: 15% were below the normal range. Patients with values >97th or <3rd percentile: 1/39>P97; 6/39<P3<br>Plasma phe sig. correlated with prealbumin (r=0.479, P=.002)                         | Prealbumin, mg/dL (ref range 20–40)  | <b>PKU:</b><br>22.6 [11.2–53.1]   |   |

|                                |                              |  |   |  |                                     |                                  |
|--------------------------------|------------------------------|--|---|--|-------------------------------------|----------------------------------|
|                                |                              |  |   | Total protein, g/dL                      | 7.2 [6.0–7.8]                       |                                  |
|                                |                              |  |   | (ref range: 6–8)                         |                                     |                                  |
| <b>Crujeiras et al. (2015)</b> | Total protein and prealbumin | Age, dietary adherence, time of diagnosis, classification of PKU and BH4 | Total protein was in the normal range in almost all patients. Prealbumin was <21mg/dL in 34.6% of patients, (74% from PKU) with 94.4% of them <18y, 96.3% having an adequate adherence and 12.96% were on BH4 treatment. Prealbumin was sig. lower in mHPA vs. PKU (p = 0.024) Significant positive correlation between age and prealbumin (p < 0.001) and total protein (p = 0.002). Total protein (p = 0.0072) and prealbumin (p <0.001) were found sig. lower in the patients with high adherence vs. low adherence to diet. More significantly altered in low adherence and >18 years compared to other participants. Time of diagnosis: no differences found | Prealbumin, mg/dL (ref range: 21–41)     | <b>mHPA:</b> 21.2                   | <b>PKU:</b> 24.5                 |
|                                |                              |  |   | Prealbumin, mg/dL                        | <b>Aged ≤18 years:</b> 21.9         | <b>Aged &gt;18 years:</b> 28.5   |
|                                |                              |  |   | Total protein, g/dL (ref range: 6.3-8.5) | 7.05                                | 7.35                             |
|                                |                              |  |   | Prealbumin, mg/dL                        | <b>High dietary adherence:</b> 22.6 | <b>Low dietary adherence:</b> 29 |

|                                 |                              |                            |   |                                    |   |   |
|---------------------------------|------------------------------|----------------------------|---|------------------------------------|---|---|
|                                 |                              |                            |   | Total<br>protein, g/dL             | 7.1   | 7.3   |
|                                 |                              |                            |   | Prealbumin,<br>mg/dL               | <b>Low<br/>adherence<br/>+ &gt; 18<br/>years:<br/>28.91</b> | <b>Other<br/>participant<br/>s:<br/>22.81</b>   |
|                                 |                              |                            |   | Total<br>protein, g/dL             | 7.41  | 7.09  |
| <b>Doulgeraki et al. (2014)</b> | Total protein, albumin       | None assessed              | All biochemical measures were within normal limits. | No data provided                   |   |   |
| <b>Pena et al. (2021)</b>       | Albumin, prealbumin, and BUN | Type of protein substitute | No difference in the biochemical parameters.        | BUN (mg/dL):<br>Prealbumin (mg/dL) | <b>L-AA:</b><br>1.68 ± 0.63<br>240 (224–278);               | <b>cGMP-AA:</b><br>1.90 ± 0.55<br>272 (200–293) |
| <b>Pinto et al. (2017)</b>      |                              |                            | All parameters remained unchanged.                  | Albumin (g/dl)                     | <b>L-AA:</b><br>4.69± 0.33                                  | <b>cGMP-AA:</b><br>4.69±0.21                    |

|                                    | Albumin<br>and<br>prealbumin                    | Type of<br>protein<br>substitute |   | Prealbumin<br>(mg/dl)  | 249± 28   | 245±53   |
|------------------------------------|---|----------------------------------|---|--|---|--|
| <b>van Vliet et al.<br/>(2019)</b> | Albumin,<br>total<br>protein, and<br>prealbumin | BH4,<br>dietary<br>parameters    | No significant differences between groups with relevant<br>biochemical measures.<br><br>No correlations were found between natural protein intake<br>and relevant biochemical markers | Min-max<br>(median)<br><br>Albumin,<br>g/L<br><br>Patients with<br>deficiency:<br><br>Patients with<br>excess:<br><br>Total<br>Protein, g/L<br><br>Patients with<br>deficiency:<br><br>Patients with<br>excess:<br><br>Pre albumin,<br>g/L | <b>PKU-<br/>nBH4:</b><br><br>43-53 (48)<br><br>0<br><br>4(24%)<br><br>58-84 (73)<br><br>1 (2%)<br><br>1 (2%)<br><br>0.15–0.51<br>(0.30) | <b>PKU-BH4:</b><br><br>44-53 (49)<br><br>0<br><br>3 (23%)<br><br>66-81 (74)<br><br>0<br><br>3 (10%)<br><br>0.17–0.45<br>(0.30) |

Patients with 6 (11%); 1 (4%)  
 deficiency:  
 Patients with 4 (8%) 3 (11%)  
 excess:

|                            |            |   |  |   |   |  |                   |
|----------------------------|------------|---|--|---|---|--|-------------------|
| <b>Rocha et al. (2010)</b> | Prealbumin | Age, classification of PKU, amount of protein substitute, metabolic control | Prealbumin z-score was $-0.5248$ (1.09), significantly below z-score = 0 ( $p < 0.001$ ). 9 patients (13%) had a prealbumin z-score $< 5$ th percentile ( $-1.64$ ), and one $> 95$ th percentile ( $+1.64$ ). All patients with z-score of prealbumin $< 5$ th percentile were $< 15$ years. No association between classification of PKU and prealbumin z-score $< 5$ th percentile ( $p = 0.237$ ).<br>No sig correlation between prealbumin z-score and the amount of protein substitute prescribed ( $R^2 = 0.01$ ; $p = 0.38$ ). Prealbumin z-score and blood phe were not sig. correlated ( $R^2 = 0.003$ ; $p = 0.65$ ).<br>Prealbumin $< 20$ mg/dl: 38 patients (55%)<br>Association between prealbumin $< 20$ mg/dl and age $< 15$ years ( $p < 0.0001$ ). Prealbumin sig. higher in the group of older patients vs. group of patients $< 15$ years ( $p < 0.001$ ). | Classification of PKU with prealbumin z-scores $< 5$ th percentile<br>Prealbumin, mg/dL | <b>Classical PKU</b><br>5%<br><b>Aged <math>&lt; 15</math> years:</b><br>$18.38 \pm 4.12$ | <b>mild PKU</b><br>16%<br><b>15 years and over</b><br>$25.53 \pm 4.38$ | <b>HPA</b><br>21% |
|----------------------------|------------|---|--|---|---|--|-------------------|

|                                  |                     |                                 |   |   |  |   |
|----------------------------------|---------------------|---------------------------------|---|---|--|---|
| <b>Gokmen-Ozel et al. (2009)</b> | Albumin             | Type of protein substitute, age | >18 years taking LPS: sig. increase in albumin ( $p < 0.05$ ), although within the reference ranges<br>7–18 years taking LPS: median albumin sig. improved ( $p < 0.01$ ), median values were within the reference range. | Median (range)<br>Albumin (g/L):<br>Baseline:<br>On LPS:<br>Median diff between PPS and LPS | <b>Aged <math>\leq 18</math> years:</b><br>43 (35–48) (n=21)<br>45 (42–49) (n=20)<br>2.5 (-3.0 to 9.5) | <b>Aged &gt;18 years:</b><br>43.5 (38.5–48.0) (n=9)<br>47 (41–49) (n=11)<br>2.8 (-2 to 7) |
| <b>Kose et al. (2016)</b>        | Albumin, prealbumin | None assessed                   | Albumin was not sig. different between PKU vs. controls.<br>Prealbumin was significantly higher in PKU vs. controls.  | No data provided  |  |   |
| <b>Mazzola et al. (2015)</b>     | BCAA                | Exercise                        | In rest and fasted state, patients showed lower levels of BCAA in comparison to controls ( $p = 0.001$ ). Levels of BCAA were not modified with exercise in PKU and controls.   | BCAA ( $\mu\text{mol/L}$ )  | <b>PKU:</b> 332 $\pm$ 50   | <b>Controls:</b> 456 $\pm$ 86   |
| <b>Desloovere et al. (2014)</b>  | Prealbumin PAA      | Age, protein substitute,        | Prealbumin levels were $< 20$ mg/dL in 60 % of participants<br>Prealbumin correlated with age. No sig, correlation was  | No data provided  |  |   |





|                                 |  |                               |   |   |                                |                                 |                                 |                                 |  |
|---------------------------------|--|-------------------------------|---|---|--------------------------------|---------------------------------|---------------------------------|---------------------------------|--|
| <i>Douglas et al.</i><br>(2013) | Creatinine,<br>BUN,<br>albumin,<br>total protein | BH4,<br>dietary<br>parameters | Classified as Definitive (DR) or Provisional (PR)<br>Responders to BH4. Non-responders (NR) discontinued drug.<br>Protein markers did not differ between groups or with increased intact protein intake in Responders. Baseline creatinine was sig. lower in PKU vs controls, but within normal ranges. BUN declined in DR and albumin, globulin, and total serum protein declined in NR over time (p <0.05). | Baseline<br>urine<br>creatinine(m<br>g/dL)  | <b>PKU:</b><br>74±40           | <b>Controls:</b><br>141±61      |                                 |                                 |  |
| <i>Das et al.</i><br>(2010)     | Urea   | Not<br>reported               | Urea concentrations were lower than normal in all patients.   | No data<br>provided   |                                |                                 |                                 |                                 |  |
| <i>Singh et al.</i><br>(2010)   | Prealbumin,<br>total protein<br>and albumin      | BH4                           | Albumin and total protein: within reference range for all patients and remained stable<br>Prealbumin: lower end of ref range at baseline and increased during the first 12 mo of follow-up (p<0.001), then remained stable.   | <b>Baseline:</b><br>Prealbumin<br>(mg/dl):<br>(ref range:<br>19.0–38.0)<br>19.8 (3.3)<br>Albumin<br>(g/dl): | <b>6 months:</b><br>21.0 (2.8) | <b>12 months:</b><br>21.7 (3.1) | <b>18 months:</b><br>23.0 (7.1) | <b>24 months:</b><br>23.3 (3.4) |  |

(ref range:  
3.5–5.0) 4.7  
(0.1)  
Total protein 6.8 (0.2) 6.9 (0.3) 6.9 (0.2) 6.9 (0.3)  
(g/dl):  
(ref range:  
6.3–7.9) 6.9  
(0.3)

|  |  |                            |   |                  |
|--|--|----------------------------|---|------------------|
| <b><i>Giovannini et al. (2009)</i></b> | Albumin, prealbumin, RBP, amino acid ratio | Type of protein substitute | Treated HPA vs. mild untreated HPA and controls: lower albumin, p = 0.012, prealbumin, p = 0.005, RBP, p = 0.001, and amino acid ratio, p < 0.0001, respectively.<br>Three days after introducing the new protein substitute prealbumin improved (p = 0.02).                          | No data provided |
| <b><i>Rocha et al. (2009)</i></b>      | Prealbumin                                 | Age                        | 8 (13%) revealed prealbumin z scores <5th percentile<br>Significant linear correlation between prealbumin and haemoglobin conc (adjusted for age) (R <sub>2</sub> =0.446; p=0.017)<br>Prealbumin z-score average was significantly lower in the group with low haemoglobin (p=0.051). | No data provided |

|                                  |  |                            |   |  |   |  |
|----------------------------------|--|----------------------------|---|--|---|--|
| <b>van Calcar et al. (2009)</b>  | PAA, prealbumin, albumin, total protein, BUN | Type of protein substitute | BUN was sig. lower with GMP diet on both day 7 and day 8 than with the AA diet on day 4. No sig. differences among albumin, prealbumin, or total protein on the last day of the AA diet (day 4) compared with the GMP diet (day 8). Total PAA was sign. greater, and BUN was sig. lower, with the GMP diet compared with the AA diet when measured 2.5 h after eating breakfast. GMP vs AA diet led to 2.25- to 2.47-fold increase in postprandial conc of isoleucine and threonine within 24 h of ingesting the GMP diet (consistent with the high concentrations of these AAs in GMP). No further sig increases in isoleucine and threonine after days 5 and 7, respectively. | Blood urea nitrogen (mmol/L)<br>Total protein (g/L)<br>Albumin (g/L)<br>Prealbumin (g/L) | <b>AA diet:</b><br>4.2 ± 0.3<br>68 ± 1.4<br>44 ± 0.9<br>317 ± 7.5 | <b>GMP diet:</b><br>3.4 ± 0.2<br>67 ± 1.4<br>44 ± 0.8<br>310 ± 7.3 |
| <b>Modan-Moses et al. (2007)</b> | Total protein and albumin                    | Dietary adherence          | Total protein and albumin were normal in all patients and did not differ between diet-adherent and non-adherent.  | No data provided   |   |  |
| <b>van Rijn et al. (2007)</b>    | Amino acids,                                 | None assessed              | Both groups were comparable in baseline albumin and total protein.  |  | <b>PKU:</b><br>43 ± 2   | <b>Controls:</b><br>45 ± 2   |

|            |   |  |         |         |
|------------|---|--|---------|---------|
| albumin,   | Sig differences in PAA concentrations between the two groups in the pre-prandial period for phe and cystine.  | Albumin  |         |         |
| total      |   | (g/L)  |         |         |
| protein,   | Sig higher valine, isoleucine, leucine, phe, and lysine in PKU vs. controls at the end of the meal period.  | Total protein  | 70 ± 3  | 71 ± 3  |
| whole-body |   | (g/L)  |         |         |
| protein    | Whole-body protein metabolism:<br>The Ra of valine did not differ between groups before and after meals. Sig higher oxidation rate during the prandial vs pre-prandial period in PKU group (p <0.01).<br>Prandial period, sig. difference in the Ra values of dietary valine into the peripheral circulation in PKU vs controls (p=0.02). Meal decreased whole-body protein breakdown in both groups to a similar extent. Net protein balance: no sig. difference between groups during pre-prandial or prandial phase. | Valine (Ox)  | 24 ± 6  | 28 ± 4  |
| metabolism |   | Pre-prandial   |         |         |
|            |   | Prandial   | 35 ± 2  | 33 ± 7  |
|            |   | R <sub>a</sub> (dietary valine into peripheral circulation), | 51 ± 8  | 42 ± 4  |
|            |   | μmol valine/kg/h   |         |         |
|            |   | Net protein balance,   | -17 ± 6 | -21 ± 4 |
|            |   | μmol valine/kg/h   |         |         |
|            |   | Pre-prandial   |         |         |
|            |   | Prandial   | 23 ± 8  | 16 ± 9  |

|                                   |                                |                            |   |  |  |  |
|-----------------------------------|--------------------------------|----------------------------|---|--|--|--|
| <i>Giovannini et al. (2006)</i>   | PAA, total protein and albumin | Type of protein substitute | Baseline: AA profiles and blood levels of protein were comparable between groups. Plasma proteins and albumin levels sig. increased from T0 to T1 in Group 1. Changes in plasma protein and albumin in Group 2 were not significant. From T0 to T1 the amino acid profiles showed an increased in methionine, lysine and arginine concentrations (all p=0.02). At T1 there was a sig. difference in tyrosine in Group 1 vs Group 2. | Mean ± SD<br>Plasma proteins, mg/dL<br>T0<br>T1<br><br>Albumin, mg/dL<br>T0<br>T1<br><br>Tyrosine (µmol/L)<br>T1 | <b>Group 1</b><br><b>(100% N</b><br><b>daily</b><br><b>needs)</b><br><br>6.9 ± 0.3<br><br>7.3 ± 0.2<br><br>4.3 ± 0.1<br><br>4.5 ± 0.2<br><br>80 ± 18 | <b>Group 2</b><br><b>(80% N</b><br><b>daily</b><br><b>needs)</b><br><br>No data<br>provided<br><br>No data<br>provided<br><br>No data<br>provided<br><br>47 ± 17 |
| <b>Lambruschini et al. (2005)</b> | Albumin                        | BH4                        | No sig. difference in albumin before and after BH4 treatment.   | Albumin, g/L   | <b>Before</b><br><b>BH4:</b><br>46 ± 5   | <b>After BH4:</b><br>45 ± 3  |

**Arnold et al. (2002)**      Prealbumin      Height, age, metabolic control

Mean prealbumin conc: 20.5 mg/dL

Prealbumin conc was positively correlated with both height and age: children with higher prealbumin were taller ( $r = 0.38, P < .02$ ) and older ( $r = 0.65, P < .001$ ).

Prealbumin was also positively correlated to plasma phe levels ( $r = 0.38, P < .03$ ). Both lower phe levels and younger age were found in the low prealbumin group.

Multiple regression analysis: After controlling for age, BMI, and mean phenylalanine level, children with prealbumin  $<20$  mg/dL had a mean height decrease of 44.9 percentiles.

| <b>Arnold et al. (2001)</b> | Prealbumin, albumin, total protein and PAA | Age, metabolic control | Albumin and total protein levels: normal in all 41 patients, and did not differ sig. from the normal ranges. Prealbumin was sig. lower in younger children ( $p = 0.03$ ). Sig. positive correlation between prealbumin and phe level ( $r = 0.38, p = 0.02$ ). Prealbumin deficiency ( $<15$ mg/dL) $n = 2$ , both poorly compliant with protein substitutes. Marginal prealbumin ( $<20$ mg/dL) $n = 12$ . All age ranges | Prealbumin, mg/dL               | <b>Aged &lt;6 years:</b> | <b>Aged &gt;6 years:</b> |
|-----------------------------|--|------------------------|---|---------------------------------|--------------------------|--------------------------|
|                             |  |                        |   | 19.5                            | 22.1                     |                          |
|                             |  |                        |   | Deficiency in at least one EAA: | <b>PKU (n):</b> 9        | <b>Controls (n):</b> 1   |

and were among the most compliant patients.

The PKU children with low prealbumin were more likely to have an EAA deficiency (p = 0.05).

| <b>Mönch et al. (1996)</b> | 24h urinary nitrogen excretion, 13C-enrichment of expired CO2 | Dose of protein substitute | Study 3: Reduced nitrogen excretion with PS taken in 3 vs. 2 doses.<br><br>Study 4: Higher and later maximum 13C-enrichment of expired CO2 and high oxidation with PS taken as one large dose vs only one-third of PS taken. Increased nitrogen excretion when taking one large portion compared to only one-third of the total daily amount of PS. | Study 3: 24-h urinary nitrogen excretion (g/24 h)<br><br>Study 4: Maximum 13C-enrichment of expired CO2 13C-leucine (Ox) after 5 h<br><br>24-h urinary nitrogen | <b>PS in two portions:</b><br><br>6.3-12.4<br><br><b>One large dose:</b> 12 ‰ after 3 h<br><br>19.5%<br><br>6.9 | <b>PS in three portions:</b><br><br>4.7-10.8<br><br><b>1/3 of dose taken:</b> 7‰ after 2 h<br><br>9.5%<br><br>4.3 |
|----------------------------|---|----------------------------|---|---|---|---|
|                            |   |                            |   |   |   |   |

|                                 |   |                  |  | excretion<br>(g/24 h)  |                                 |                                 |                                 |                                      |
|---------------------------------|---|------------------|--|--|---------------------------------|---------------------------------|---------------------------------|--------------------------------------|
| <b>Acosta et al.<br/>(1999)</b> | PAA,<br>albumin,<br>BUN, RBP,<br>prealbumin | None<br>assessed | All mean plasma indices of protein status were in normal reference ranges. Amino Acids: Mean conc of all amino acids except Cys, Gly and Phe were in the reference ranges. Amino acids below the lower limit of the ref: Arg (12%), Cys (71%), Ile (12%), Lys (14%) and Thr (11%). Dietary intakes of Arg (month 3, r = 0.36, p = 0.05), Met (month 4, r = 0.43, p = 0.05), Phe (months 1, 2 and 5, r = 0.65, p = 0.01; r = 0.42, p = 0.05; r = 0.33, p = 0.07), Trp (month 1, r = 0.51, p = 0.05), Tyr (month 4, r = 0.37, p = 0.05) and Val (month 3, r = 0.51, p = 0.01) were positively correlated with the respective PAA conc. | Mean (SD)<br>[n=]<br>Albumin,<br>g/dL<br>(ref range:<br>3.0-4.6)<br><b>Baseline</b><br>3.7 (0.1)<br>[22]<br>RBP, mg/dL<br><b>Baseline:</b><br>NE<br>Prealbumin,<br>mg/dL<br>(ref range:<br>6.7-21) | <b>1mo</b><br>3.6 (0.1)<br>[24] | <b>3mo</b><br>4.1 (0.1)<br>[23] | <b>6mo</b><br>4.1 (0.1)<br>[26] | <b>During<br/>study</b><br>3.9 (0.1) |
|                                 |   |                  |  |  | NE<br>[22]                      | 3.38 (0.2)<br>[22]              | 3.74 (0.2)<br>[21]              | 3.56 (0.2)                           |
|                                 |   |                  |  |  | NE<br>[22]                      | 17.6 (0.8)<br>[22]              | 17.9 (0.9)<br>[22]              | 17.8 (0.8)                           |



---

**Baseline:**

NE

Urea 12.3 (0.8) 12.0 (0.6) 11.9 (0.5) 12.1 (0.6)

nitrogen, [23] [24] [24]

mg/dL

(ref range: 5-

17)

**Baseline:**

12.9 (1.2)

[23]

---

|  |                                      |                       |   |  |   |  |
|--|--------------------------------------|-----------------------|---|--|---|--|
| <b>Hillman et al.</b><br><b>(1996)</b> | Albumin,<br>creatinine               | Dietary<br>parameters | Serum albumin and creatinine were similar in children with PKU and controls. No sig. correlations between total protein intake, protein intake/kg, or serum albumin after age correcting. | <b>PKU:</b><br>Albumin,<br>g/dL<br>Serum<br>creatinine | 4.9 ± 0.5<br>(n=11)<br>0.97± 0.19<br>(n=11) | <b>Controls:</b><br>5.1 ±.58<br>(n=35)<br>0.82±.0.10<br>(n=18) |
| <b>Graffin et al.</b><br><b>(1995)</b> | Total<br>protein,<br>albumin,<br>BUN | None<br>assessed      | Children had normal levels of total protein; but abnormal values for albumin (high) and BUN (low)   | No data<br>shown                                       |   |  |

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|                               |                                    |  |  |                                 |                                |                                    |
|-------------------------------|------------------------------------|--|--|---------------------------------|--------------------------------|------------------------------------|
| <b>Thompson et al. (1990)</b> | PAA, whole-body protein metabolism | Plasma phe levels, classification of PKU | Rates of protein synthesis in PKU were similar to or above control values, as were rates of protein catabolism. Net protein loss during fasting tended to be lower in PKU than in controls (not statistically different). Protein turnover values were similar in HPA to PKU and controls. Protein synthesis did not change sig. with plasma phenylalanine concentration. Amino Acids: Mean conc of many AAs were in the lower normal range in PKU on normal diets without PS. Those with protein-restricted diets and PS, the mean conc of all AAs other than phe were similar to controls. |                                 |                                |                                    |
| <b>Nord et al. (1988)</b>     | Total protein, albumin and BUN     | Dietary adherence, diagnosis             | Total protein, albumin and BUN were within normal limits and no sig diff between groups.   | No data provided                |                                |                                    |
| <b>Shenton et al. (1983)</b>  | Prealbumin and albumin             | None assessed                            | No sig difference in albumin between groups. Significant difference in prealbumin between groups (p<0.01)  | Mean (SD) (range) Albumin (g/l) | <b>PKU:</b> 44.4 (3.3) (40-53) | <b>Control:</b> 44.6 (3.1) (37-50) |

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|            |           |           |
|------------|-----------|-----------|
| Prealbumin | 168 (33)  | 216 (47)  |
| (mg/L)     | (105-219) | (128-332) |

---

|                    |     |            |  |
|--------------------|-----|------------|--|
| <b>Pena et al.</b> | BUN | Type of    | Meta-analysis for BUN reported no sig. differences |
| <b>(2018)</b>      |     | protein    | between GMP-AAs and AAs (MD = -0.22 mg/dL (-1.49,  |
|                    |     | substitute | 1.04); I2 = 0%; p = 0.73)                          |

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Note: Authors in italics are abstract only papers

PKU: phenylketonuria; HPA: hyperphenylalaninemia; Phe: phenylalanine; SMM: skeletal muscle mass; BUN: blood urine nitrogen; PAA: plasma amino acids; AA: amino acids; L-AA: L-amino acids; RBP: retinol-binding protein; cGMP-AA: casein glycomacropeptide amino acid; PS: protein substitute; PPS: Pretrial Protein Substitute; LPS: Liquid Protein Substitute; AUC: area under the curve; NE: Not evaluated

**Table 6: Functional measurements of protein status and key findings**

| Author (Year)                  | Method used   | Modulating factors                                 | Key findings: Protein status outcomes  | Variable   | Group 1                         | Group 2  |
|--------------------------------|---|--|--|--|---------------------------------|--|
| <b>Choukair et al. (2017)</b>  | Maximal isometric grip force (hand dynamometry)             | Sex, MCA, classification of PKU, metabolic control | Mean grip force z-score was significantly decreased compared with the reference population ( $-0.64 \pm 1.26$ ; $p = 0.0004$ ); observed both in female and male patients. 38% a grip force <3rd percentile. A significant linear correlation between MCA and grip force was found for PKU patients ( $r = 0.827$ , $p < 0.0001$ ) and reference population ( $r = 0.66$ , $p < 0.001$ ). No relationship between grip force (z-scores) and PKU type or mean phe.  |  |                                 |  |
| <b>Sumanszki et al. (2020)</b> | Maximum physical stress test: evaluated VO <sub>2</sub> max | None assessed                                      | Duration of aerobic or anaerobic exercise: No significant difference between groups (PKU vs controls, $p = 0.883$ and $p = 0.247$ , respectively). VO <sub>2</sub> max: Significantly lower in the PKU group vs. controls ( $p = 0.004$ ); relative VO <sub>2</sub> max (adjusted for body weight) was similar between groups. Cumulative workload (watts): Significantly higher in the control compared with the PKU group ( $p=0.002$ ). Handgrip test: Used as a stress stimuli intervention and no outcome data provided | VO <sub>2</sub> max (ml/min)<br><br>Relative VO <sub>2</sub> max (ml/kg/min) | <b>PKU:</b><br>3080 (2813–3768) | <b>Controls:</b><br>3970 (3758–4108)<br><br>48.5 (46.5–59) |
| <b>Mazzola et al. (2015)</b>   |   | Metabolic control                                  | Patients showed similar aerobic capacity and workload peak in the VO <sub>2</sub> peak test in comparison to controls. PKU patients and controls showed similar values of  | VO <sub>2</sub> peak (mL/kg/min)   | <b>PKU:</b><br>28 ± 8           | <b>Controls:</b><br>31 ± 6                                 |

|               |  |                            |          |          |
|---------------|--|----------------------------|----------|----------|
| VO2peak       | prescribed and actual VO2 during exercise. Poorly controlled patients showed the lowest percentage of actual VO2 during exercise in relation to the prescribed value (not sig) | Workload peak (W)          | 203 ± 31 | 216 ± 49 |
| Workload peak |  | Prescribed VO2 (mL/kg/min) | 21 ± 6   | 22 ± 4   |
|               |  | Actual VO2 (mL/kg/min)     | 18 ± 5   | 22 ± 4   |

## Supplemental material: Protein intake data

| Author (Year)                      | Protein requirements            | Total protein intake   | Natural protein intake   | Protein substitute intake  | Additional comments   |
|------------------------------------|---------------------------------|--|--|--|---|
| <b>ANTHROPOMETRIC MEASURES</b>     |                                 |  |  |  |   |
| <b>Daly et al.</b><br>(2021)       | Not reported                    | Not reported   | Median amount of prescribed natural protein: 5.5 g protein/day (range 3–30 g) or 275 mg/day of phenylalanine (range 150–1500 mg) | Median daily dose of protein substitute: 60 g/day (range 40–80 g)                    | cGMP-AA and L-AA groups. All were adherent with protein substitute            |
| <b>Alfheaid et al.</b><br>(2018)   | Not reported                    | Not reported   | Not reported   | Not reported   | Unclear what patients' normal dietary patterns are.                           |
| <b>Dobbelaere et al.</b><br>(2003) | RDAs (Comite de Nutrition 2001) | Protein intake varied from 1.2 to 2.1 (mean 1.67±0.23) g/kg per day, representing 109-191% (146%±25%) of RDA         | Phe intake ranged from 204-768mg/day (based on 4DDD)   | Not reported   |   |
| <b>Huemer et al.</b><br>(2007)     | RDA + 20-40% (DACH 2000)        | Mean total protein intake: 1.2 ± 0.3 g/kg per day (median 1.1, range 0.8–2.4) in PKU patients.<br>Mean total protein | Mean natural protein intake: 0.3 g/kg/day (SD 0.2, median 0.24, range 0.1–1.1)   | Mean Phe-free L-AA mixture intake: 0.9 g/kg/day (SD 0.2, median 0.84, range 0.6–1.4) | Mean total protein intake in PKU patients was 124% (range 77–193%) of the RDA |

|                                |   |  |   |  |  |
|--------------------------------|---|--|---|--|--|
|                                |   | intake: 33.7 ± 10.3 g/day<br>(median 32.5, range 11.3–32.5).   |   |  |  |
| <b>Sailer et al.</b><br>(2020) | Not reported                              | g/kg:<br>Female: PKU 1.3 ± 0.56<br>vs Con 1.9 ± 0.96<br>Male: PKU 1.44 ± 0.48 vs<br>Con 2 ± 0.77<br><br>No diff in the total grams<br>of protein/ kg in PKU<br>compared to controls. | g/kg:<br>PKU<br>Female: 0.36 ± 0.31<br>Male: 0.41 ± 0.32<br>(no sig. difference)    | g/kg:<br>PKU<br>Female: 0.95 ± 0.65<br>Male: 1.03 ± 0.37<br>(no sig. difference) | % total energy of protein among<br>PKU male subjects was lower than<br>male control subjects (10.4 ± 2.1%<br>vs. 14.5 ± 3.7; p=0.003), but %<br>energy from protein was similar<br>between PKU female participants<br>and controls (10.1 ± 2.6% vs. 11.0 ±<br>5.2%; p = 0.63).<br><br>Protein to energy ratio was<br>significantly lower among PKU male<br>subjects compared to controls (2.51<br>± 0.58 vs. 3.62 ± 0.92; p = .0003).<br>This difference was not seen in<br>females. |
| <b>Allen et al.</b><br>(1995)  | Not reported                              | Not reported   | Not reported  | Not reported   |  |
| <b>Evans et al.</b><br>(2017)  | FAO/WHO/UNU<br>recommended<br>safe levels | Total protein: g/kg/d<br>(median ± SD (range))<br>D-PKU: 2.05 ± 0.60   | Natural protein: g/kg/d (median ±<br>SD (range))<br>D-PKU: 0.50 ± 0.18 (0.18–0.80); | AAF g/kg/d: median ± SD<br>(range)<br>D-PKU: 1.54 ± 0.50 (0.80–                  | Reported that the median total-<br>protein intake exceeded the   |

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|                                  |              | (1.00–3.50); BH4-PKU:<br>1.90 ± 0.16 (1.70–2.10);<br>All-PKU: 2.00 ± 0.56<br>(1.00–3.50)  | BH4-PKU: 1.10 ± 0.60 (0.55–2.00);<br>All-PKU: 0.50 ± 0.37 (0.17–2.00)  | 2.70); BH4-PKU: 1.00 ± 0.61<br>(0.00–1.30); All-PKU: 1.43 ±<br>0.59 (0.00–2.70)   | FAO/WHO/UNU recommended<br>safe levels (data not shown).   |
| <b>Nogueira et al.</b><br>(2021) | Not reported | Not reported  | Not reported   | not reported  |  |
| <b>Evans et al.</b><br>(2018)    | Not reported | Not reported  | Not reported   | Not reported  | Reported participants used phe-free<br>AA formula  |
| <b>Adamczyk et al.</b><br>(2011) | Not reported | Not reported  | Not reported   | Not reported  | L-AA PKU supplements   |
| <b>Mazzola et al.</b><br>(2016)  | Not reported | Not reported  | Not reported   | Not reported  | No access to protein-enriched low-<br>phe food   |
| <b>Rocha et al.</b><br>(2013)    | Not reported | Mean (SD): 1.92 (0.57)<br>g/kg/day for patients<br>aged <19 years (n=63)<br>and 1.43 (0.35) g/kg/day<br>for patients aged ≥19<br>years (n=26) | Mean (SD): 0.76 (0.46) g/kg/day for<br>patients aged <19 years (n=63);<br>0.57 (0.39) g/kg/day for patients<br>aged ≥19 years (n=26) | Mean (SD): 1.39 (0.44)<br>g/kg/day for patients aged<br><19 years (n=63); 1.03<br>(0.45) g/kg/day for patients<br>aged ≥19 years (n=26) | Total protein (g/kg/d)<br>HPA and mild PKU 1.92 ± 0.61<br>Classical PKU 1.88 ± 0.34<br>Natural Protein (g/kg/d): HPA and<br>mild PKU 0.82 ± 0.49 Classical PKU<br>0.49 ± 0.14 g/kg/d |



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|                                |              |   |                      |   | AA Mix (g/kg/d):<br>HPA and mild PKU 1.33 ± 0.45<br>Classical PKU 1.67 ± 0.3                          |
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| <b>Weng et al.</b><br>(2020)   | DRI          | 1.265 ± 0.592 g/kg/d<br>(%DRIs 105.448 ± 33.41) | 0.874 ± 0.602 g/kg/d | 12 (55%) PKU pts phe-free formula; 10 non-phe-free formula (getting 100% of DRI from natural protein)   | 71% (70.90±31.24) of protein consumed by PKU patients was from medical food and 29% from natural food |
| <b>Stroup et al.</b><br>(2018) | Not reported | Not reported                                    | Not reported         | g PE from AA-MF: Male: 67 ± 6; Female: 52 ± 4<br>(p(sex)=0.057; p(gt)=0.09)<br><br>g PE from AA-MF/kg: Male 0.89 ± 0.09 Female: 0.77 ± 0.08 (p(sex)=0.46; p(gt)=0.21)<br><br>g PE from AA-MF/kg lean mass:<br>Males: 1.20 ± 0.08 females: 1.24 ± 0.10 (p(sex)= 0.54; p(gt)= 0.14) | Discussed the negative impact on BMD with increase protein substitute                                 |

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| <b>Jani et al.</b><br>(2017)  | 120-140% RDA | Total Protein (g/day;<br>g/kg/day):<br>Total (n=83): 59.3 (10.1,<br>119.7); 1.09 (0.19, 2.20)<br>Adults (n=25): 69.9<br>(42.1, 119.6); 0.92 (0.86,<br>0.76)<br>Children (n=58): 53.1<br>(10.1, 119.7); 1.12 (0.63,<br>0.97)<br><br>Median total protein<br>intake (g/d) was highest<br>among males (57.5,<br>range: 46.8, 76.2)<br>compared to PKU<br>patients | Intact protein (g/day); g/kg/day:<br>Total: 13.4 (3.6, 79.4); 0.25 (0.07,<br>1.46)<br>Adults: 25.1 (8.2, 79.4); 0.33 (0.17,<br>0.50);<br>children: 11.3 (3.6, 74.7); 0.24<br>(0.23, 0.61).<br><br><b>Adults:</b> higher median intakes of<br>intact protein (25.1 vs. 9.9 g/d, U =<br>73.5, p < 0.001) compared to<br>prescribed intakes.<br><br><b>Children:</b> actual median intact<br>protein intake was higher than<br>prescribed<br>(6.0 vs. 11.3 g/d, U=2549.5, p <<br>0.001). | MF protein (g/day;<br>g/kg/day):<br>Total (n=73): 45.0 (5.0,<br>90.0); 0.83 (0.31, 0.57)) g/d;<br><br>Adults (n=19): 53.8 (25.0,<br>90.0); 0.71(0.51, 0.57) g/d;<br><br>Children (n=54): 43.2 (5.0,<br>79.6); 0.91 (0.31, 0.65) g/d<br><br>Adults: consuming lower<br>actual median intakes of MF<br>(53.8 vs. 60.0 g/d, U = 133.5,<br>p = 0.03) compared to<br>prescribed intakes | Majority of participants consumed<br>higher intact proteins (n= 76, 92.7%<br>vs n = 6, 7.3%) than as prescribed. A<br>similar proportion consumed<br>medical food protein as per<br>prescription (n = 35, 51.5%) or<br>lower than as prescribed (n=33,<br>48.5%). |
| <b>Paci et al.</b><br>(2018)  | Not reported | Not reported   | Not reported  | Not reported   | Dietary data focus on glycaemic<br>index and glycaemic load   |
| <b>Mexia et al.</b><br>(2015) | Not reported | Not reported   | Not reported  | Not reported   |   |

|                                   |   |              |              |   |   |
|-----------------------------------|---|--------------|--------------|---|---|
| <b>Torriente et al.</b><br>(2017) | Not reported                                  | Not reported | Not reported | Not reported  |   |
| <b>Daly et al.</b><br>(2019)      | Not reported                                  | Not reported | Not reported | Not reported  |   |
| <b>Kanufre et al.</b><br>(2015)   | Not reported                                  | Not reported | Not reported | Not reported  |   |
| <b>Bonifant et al.</b><br>(2010)  | Not reported                                  | Not reported | Not reported | Not reported  |   |
| <b>Rocha et al.</b><br>(2010)     | Not reported                                  | Not reported | Not reported | Not reported  | Medians of natural protein and protein substitute intake were not significantly different in patients with overweight/obesity compared to the others. |
| <b>Wilcox et al.</b><br>(2011)    | Not reported                                  | Not reported | Not reported | Not reported  |   |
| <b>Nalin et al.</b><br>(2013)     | Not reported                                  | Not reported | Not reported | Not reported  |   |
| <b>BIOCHEMICAL MEASURES</b>       |   |              |              |   |   |
| <b>Prince et al.</b><br>(1997)    | (Phase 2) RDA minus exchanges and prescribing | Not reported | Not reported | AA Intakes (g protein/kg)<br>Prescribed by clinician<br>Entry (yr. 1): 1.15 ±0.5<br>End (yr. 5): 0.67 ± 0.2 | Mean ages associated with these intakes were 6.9 years at entry and 11.1 years at end, which results in the mean 'Received Intakes' of AA =           |

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|                                 |              |  |   | <p>P&lt;0.001</p> <p>Reported by participant</p> <p>Entry (yr. 1): 1.31 ± 0.8</p> <p>End (yr. 5): 0.68 ± 0.2</p> <p>P&lt;0.001</p> <p>Received by participant</p> <p>Entry (yr. 1): 0.91 ± 0.4</p> <p>End (yr. 5): 0.4 ± 0.3</p> | <p>75% RDA protein and 40% RDA protein, respectively. Observed no significant reductions in protein status, with 'Received Intakes' 50–100% below prescribed intakes recommended by Medical Research Council (1993) for the mean age of subjects.</p> |
| <b>Singh et al. (2010)</b>      | Not reported | Total protein intake remained at approximately 1.0±0.08 g/kg per day (43.7±4.2 g/day) throughout the 24 months of the study. | At 3mo, phenylalanine increased from a baseline average of 11.9± 4.1 mg/kg to 39.9 ± 11.5 mg/kg (p=0.001), and phenylalanine intake from food increased from 15.9 ± 5.3 mg/kg to 34.2 ± 13.8 mg/kg (p=0.007). | By 3mo of BH4 therapy, n=3 were consuming a reduced MF prescription (50%, 20%, and 38%, respectively). n=3 no longer required MF. n=1 who initially discontinued MF consumption experienced a growth spurt, MF was reintroduced  | Stage 2: MF was reintroduced as needed to stabilize plasma phe <360 µmol/L and to keep serum transthyretin within normal limits   |
| <b>Giovannini et al. (2014)</b> | Italian RDA  | Children who received the test or conventional substitute: Baseline protein intake was 1.9 (0.8) vs 2.0 (0.9)                | Baseline mean (SD) PHE intake was 403 (213) vs 392 (227) mg/day in children who received the test or conventional substitute, respectively. End of the trial the  | 41 (68.3%) children had 3 doses/day (test substitute 21/30; conventional substitute 20/30) and 19 (31.7%) had 4 doses/day  | Unaffected children: protein intakes were 150% higher than the Italian RDA  |

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|                               |  | g/kg/day; end of trial was<br>1.8 (1.0) vs 2.0 (0.9)<br>g/kg/day   | corresponding values were 392<br>(207) vs 400 (208) mg/day. | (test substitute 9/30;<br>conventional substitute<br>10/30).                      | PKU: mean protein intake was<br>around 120% of RDA  |
| <b>Zaki et al.</b><br>(2016)  | All patients<br>received the<br>same total protein<br>intake to match<br>recommended<br>protein<br>requirement for<br>their age and<br>weight* | Not reported   | Not reported  | Phase I: 50 % GMP and 50%<br>AAF and Phase II: 100% AAF<br>(amounts not provided) | Required to be compliant to<br>treatment for at least two months<br>prior to start of study.  |
| <b>Mönch et al.</b><br>(1996) | Not reported   | Not reported   | Not reported  | Not reported  | Took their usual prescription<br>divided in 2 and 3 doses   |
| <b>Thompson et al.</b> (1990) | Not reported   | Mean Protein (g/kg/d)<br>PKU: 1.06 ± 0.34<br>Range:<br>Free diet: 0.76-1.64<br>g/kg/day<br>Diet+AA: 0.5-1.36<br>g/kg/day<br><br>HPA: 1.4 & 1.1 (n=2) | Unclear of current dietary protein<br>intake                | Not reported  | n = 5 were on free diet; n = 5 were<br>on PKU diet + AA<br><br>No alteration in dietary intake of<br>any subject in the 3 mo. prior to the<br>study |

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|                                 |              | Control: 1.25 g/kg/d<br>(range 0.9-1.5)   |   |   |  |
| <b>van Calcar et al.</b> (2009) | DRI          | Not reported  | Phe allowance ranged from 5.8 mg/kg (subject 10) to 26.7 mg/kg (subject 2)  | Not reported  | Two dietary treatments of 4 d each: the AA diet (days 1–4) and the GMP diet (days 5–8).  |
| <b>Ney et al.</b> (2016)        | Not reported | Protein g/d:<br>AA-MF: 80 ± 3<br>GMP-MF: 79 ± 4<br><br>Protein g/kg/d:<br>AA-MF: 1.15 ± 0.05<br>GMP-MF: 1.14 ± 0.06 | Classical: mean ± SE 0.34 ± 0.04 g protein from natural food/kg/day; 15 ± 2 mg Phe/kg/d;<br><br>Variant: 0.50 ± 0.07 g protein from natural food/kg/day, 22 ± 3 mg Phe/kg/day | Mean ± SE prescribed dose was 0.85 ± 0.03 g PEs from AA-MF/kg/d.<br><br>AA-MFs or GMP-MFs provided 66–68% of total protein intake or 0.74–0.76 g protein/kg/d | Reduce intake of natural foods that contain Phe to offset the Phe intake in GMP-MFs and maintain constant Phe intake.<br><br>Medical food logs: intake was higher for GMP-MFs during both stages of the study and significantly higher during visits 3 and 4 than it was with AA-MFs (3.74 servings GMP-MFs/d compared with 2.43 servings AA-MFs/d; P = 0.001) |
| <b>Ahring et al.</b> (2018)     | Not reported | In each test meal, the total content of protein was equivalent to 25% of 1 g/kg/d                                   | Not reported  | Given test protein substitute to provide 25% of requirements  | Intervention included four different drink mixtures:<br>DM1: 100% cGMP<br>DM2: 100% L-AA (equivalent AA profile to DM1)<br>DM3: cGMP + L-AA (to ensure nutritionally complete)   |

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|                                  |              |   |  |   | DM4: L-AA (equivalent AA profile as DM3, but without phe)   |
| <b>van Rijn et al.</b><br>(2007) | RDA + 20%    | Protein intake (g /kg/d)<br>(mean, SD)<br>PKU: 1.1 ± 0.1<br>Controls: 1.2 ± 0.1   | Tolerances of dietary Phe: based on daily intake of natural protein at 5 y of age were: 21± 9 and 11 ±4 mg Phe/kg/d at the time of the test. | Not reported  |   |
| <b>Giovannini et al.</b> (2009)  | Italian RDA  | Reports nutrient intake consistent with the Italian RDA   | Not reported   | Not reported  |   |
| <b>Giovannini et al.</b> (2006)  | Not reported | Not reported  | Not reported   | Not reported  | Compliance with the new protein substitute mixture was 100% |
| <b>Douglas et al.</b><br>(2013)  | Not reported | Definitive responders (DR): decrease in total protein intake (Baseline: 1.4±0.7, 1 year:1.0±0.7; p<0.001) due to 75% decline in medical food (MF) intake.<br>Total protein intake declined significantly in NR and PR without change in Phe tolerance | DR increased intact protein (g/kg (Baseline:0.58±0.4, 1 year:0.75±0.3)   | Intake not reported, in DR group there was a 75% decline in medical food intake |   |

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|                                  |              | or prescribed MF,<br>indicating nonadherence  |  |   |  |
| <b>Acosta et al.</b><br>(1999)   | RDA (1980)   | Protein (g/day) 17.3 ±<br>0.6<br>Protein (g/kg/day) 2.7 ±<br>0.1  | Not reported   | Medical Food (g/day) 79 ± 4   | 17% of Phenex-fed infants had<br>protein intakes below 100% of 1980<br>RDA.<br>Mean intake of EAA/kg were greater<br>than recommended. |
| <b>Shenton et al.</b><br>(1983)  | Not reported | Not reported  | Not reported   | Not reported  |  |
| <b>Schulpis et al.</b><br>(2013) | Not reported | Total protein (g)<br>Group A: 72 ± 20<br>Group B: 70 ± 18<br>Controls: 73 ± 17<br>P= NS   | Natural protein (g)<br>Group A: 40 ± 20<br>Group B: 9 ± 1.2<br>Controls: 73 ± 17<br>A vs C P<0.001<br>B vs C p<0.001<br>A vs B p<0.001 | Phe free formula dose<br>depended on age, weight and<br>residual activity of Phe<br>hydroxylase as related to<br>their molecular analysis | Group A - 'loose diet' and Group B<br>strictly adhered to diet   |
| <b>Arnold et al.</b><br>(2001)   | Not reported | Children ages 1-4 were<br>prescribed approx. 30 g<br>protein/day. Children<br>aged 4-7: 35 grams<br>protein; Children aged 7-<br>11: 40 grams protein;<br>Females ≥12 years: 50 | Not reported   | Not reported  |  |



|                                   |   |   |  |  |                                  |
|-----------------------------------|---|---|--|--|----------------------------------|
|                                   |   | grams protein; Males ≥12<br>years: 55 grams protein   |  |  |                                  |
| <b>Arnold et al.</b><br>(2002)    | Not reported  | Not reported  | Not reported   | Medical foods prescribed<br>accordingly: ages 2-4 years,<br>30 g/d; ages 4-7 years, 35<br>g/d; ages 7-11 years, 40 g/d;<br>ages ≥12 years, 50 g/d for<br>female and 55 g/d for male. |                                  |
| <b>Nord et al.</b><br>(1988)      | Not reported  | Not reported  | Not reported   | Not reported   |                                  |
| <b>Rocha et al.</b><br>(2010)     | MRC on PKU,<br>adopted by the<br>Portuguese<br>guidelines | Not reported  | Not reported   | Not reported   |                                  |
| <b>Kose et al.</b><br>(2019)      | Not reported  | Not reported  | Not reported   | Not reported   | L-AA used as protein substitutes |
| <b>Andrade et al.</b><br>(2017)   | RDA (0.8–1.3<br>g/kg/day)                                 | Total proteins, g/kg/day<br>PKU 1.0 [0.2–2.0]<br>117% RDA   | Natural proteins, g/kg/day<br>PKU 0.5 [0.2–2.0]<br>48% RDA | Not reported   |                                  |
| <b>van Vliet et al.</b><br>(2019) | Not reported  | Significantly higher<br>natural protein intake for<br>PKU-BH4 patients vs.<br>PKU-nBH4 (p < 0.001)) | Not reported   | Not reported   |                                  |

|                                  |                                       |   |   |  |  |
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| <b>Crujeiras et al.</b> (2015)   | RDA (recommendation was 1.3-1.5x RDA) | Not reported  | Not reported  | Not reported   |  |
| <b>Viau et al.</b> (2021)        | DRI 0.8 g protein/kg/d                | Mean protein intake: 73.2 ± 17.6 g/d (range: 46.9–125.4 g/d) and 1.0 ± 0.3 g/kg/d (range: 0.5–1.8 g/kg/d). Majority (16/18) of participants' intake met or exceeded the DRI, two male participants consumed less at 0.5 and 0.6 g/kg/d. | Intact Protein, g/day<br>Phe < 30 µmol/L (n=11): 72.2 ± 11.4<br>Phe ≥ 30 µmol/L (n=7): 72.5 ± 25.8<br>P=0.97<br><br>Intact Protein, g/kg<br>Phe < 30 µmol/L (n=11): 1.0 ± 0.2<br>Phe ≥ 30 µmol/L (n=7): 1.0 ± 0.4<br>P=0.84 | None   | FFQ: participants ate a median of 92.8% (IQR 70.0 – 111.1%) of the recommended daily servings of protein foods (e.g., meat, poultry, seafood, eggs, soy, nuts, seeds and legumes) and 55.4% (IQR 32.1–85.7%) of dairy foods. On average, animal proteins comprised 62 ± 10% of total protein intake. |
| <b>Pena et al.</b> (2018)        | Not reported                          | Not reported  | Not reported  | Not reported   |  |
| <b>Gokmen-Ozel et al.</b> (2009) | Not reported                          | Not reported  | Median phenylalanine exchanges were 6 x 50mg daily (range 3–15). n=6 adults were not following measured phenylalanine exchanges but avoided high protein foods  | Median protein substitute dose was 60 g PE daily (range 45–75 g day) both before and during the study. |  |

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|--|--------------|--|---|--|--|
| <b>Graffin et al.</b><br>(1995)              | Not reported | Mean protein intake: 62<br>± 15 % of their<br>recommended levels | Not reported  | Not reported   |  |
| <b>Prochazkova et al.</b> (2012)             | Not reported | Not reported   | Not reported  | Not reported   |  |
| <b>Rocha et al.</b><br>(2009)                | Not reported | Not reported   | Not reported  | Not reported   |  |
| <b>Desloovere et al.</b> (2014)              | Not reported | Not reported   | Not reported  | Not reported   |  |
| <b>Kose et al.</b><br>(2016)                 | Not reported | Not reported   | Not reported  | Not reported   |  |
| <b>ANTHROPOMETRIC + BIOCHEMICAL MEASURES</b> |              |  |   |  |  |
| <b>Lambruschini et al.</b><br>(2005)         | Not reported | Not reported   | Phe-restricted diet initially. Phe tolerance increased significantly from 356 ± 172 mg/day (mean ± SD; range: 201–600) to 1546 ± 192 mg/day (range: 1240–1801) (Wilcoxon test; p = 0.004) | Stopped in 11/14 patients with BH4 treatment   | All participants initially were treated with phe-restricted diet and protein substitutes, prior to BH4 treatment. Phe-restricted diet unknown duration |
| <b>Pena et al.</b><br>(2021)                 | Not reported | Not reported   | Natural protein intake (g/kg/day) Baseline: 0.41 (0.26–0.62) When on cGMP-AA 0.34 (0.21–0.69)   | Amount of protein equivalent from protein substitute remained unchanged [(0.86 ± 0.24 g/kg/day | At the last ANSE, CGMP-AA contributed a mean of 66 ± 31% (range 23 to 100) to the total protein substitute intake.                                     |

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|                             |   |  |              | vs $0.74 \pm 0.23$ g/kg/day; p = 0.126) and ( $50.8 \pm 16.3$ g/day vs $44.6 \pm 12.8$ g/day; p = 0.118)].   |   |
| <b>Das et al.</b><br>(2013) | DACH-<br>recommendations<br>(German-<br>Austrian-Swiss<br>dietary<br>association) (DGE<br>2012) | PKU: protein intakes<br>were below DACH-<br>recommendations<br>Protein (g/kg/d)<br>Baseline<br>Protein reduced + AAM:<br>1.1<br>Vegan + AAM: 0.9<br>Vegan – AAM: 0.5<br>Normal Food: 0.6<br><br>Follow-Up<br>Protein reduced + AAM:<br>1.0<br>Vegan + AAM: 1.0<br>Vegan – AAM: 0.7<br>Normal Food: 1.0 | Not reported | Not reported<br><br>All patients not taking AAM<br>at the beginning of the study<br>agreed to supplement their<br>original diet with an AAM<br>subsequently. | Participants were grouped into the<br>following:<br>1. Normal food (“normal food”) -<br>36%<br>2. Vegan without amino acid<br>mixture (“vegan”) -14%<br>3. Vegan with amino acid mixture<br>(“vegan + AAM”) - 8%<br>4. Protein reduced with amino acid<br>mixture which is the<br>recommended form of nutrition<br>(“PKU-diet”) - 42% |

|                                     |                      |   |  |  |  |
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| <b>Pinto et al.</b><br>(2017)       | Not reported         | Not reported  | Natural protein intake (g/kg/day)<br>AA diet: $0.47 \pm 0.27$ ; GMP diet:<br>$0.59 \pm 0.49$ (p = 0.241)           | Protein substitute (g/kg/day)<br>AA diet: 0.85 (0.73–1.08);<br>GMP diet: 0.75 (0.61–0.99)<br>(p = 0.182) | Mean GMP contribution to the total<br>protein substitute intake was 57%<br>(27 to 100%) providing an<br>additional<br>$34 \pm 12$ mg of PHE per day. |
| <b>Allen et al.</b><br>(1996)       | FAO/WHO/UNU,<br>1985 | Protein intake was<br>similar for the PKU and<br>controls (median):<br>PKU: 2.11 g/kg<br>equivalent to 209% of<br>recommended<br>Controls: 1.9 g/kg<br>equivalent to 193% of<br>recommended<br>FAO/WHO/UNU 1985 | Median phe intake was significantly<br>lower<br>in the PKU group (23 mg/kg) than<br>in<br>the controls (92 mg/kg)  | Not reported   |  |
| <b>Doulgeraki et al.</b><br>(2014)  | Not reported         | Not reported  | Not reported   | Not reported   | PKU - L-AA supplements   |
| <b>Hillman et al.</b><br>(1996)     | RDA                  | Mean intake: $46.1 \pm 12.1$<br>g/d ( $1.5 \pm 0.6$ g/kg)   | Not reported   | Not reported   | Protein intakes were above the RDA   |
| <b>Modan-Moses et al.</b><br>(2007) | RDA                  | Protein intake (g/day):<br>All: $72.7 \pm 34.9$ ; Diet-<br>adherent: $86.8 \pm 30$ ;  | PHE intake (mg/day). All: $1394 \pm$<br>982; Diet-adherent: $1097 \pm 1063$ ;<br>non-adherent: $1859 \pm 624$ (NS) | Not reported   | All diet-adherent patients achieved<br>protein intake above RDA, only 3  |

|  |              |   |              |              |   |
|--|--------------|---|--------------|--------------|---|
|  |              | non-adherent: 45.3 ±<br>22.2 (p = 0.011)<br>Protein intake was<br>significantly higher in the<br>diet-adherent patients |              |              | non-adherent patients met<br>the RDA for protein intake   |
| <b><i>Boros et al.</i></b><br>(2015)     | Not reported | Not reported  | Not reported | Not reported |   |
| <b><i>Das et al.</i></b><br>(2010)       | Not reported | Not reported  | Not reported | Not reported | 41% of PKU-patients followed<br>recommended protein restriction<br>supplemented with AM. 14% said<br>they follow a less restricted 'vegan'<br>diet supplemented with AM. 45%<br>claimed to have normal eating<br>habits without AM. |
| <b><i>Sumanszki et al.</i></b><br>(2019) | Not reported | Not reported  | Not reported | Not reported |   |
| <b>ANTHROPOM</b>                         |              |   |              |              |   |
| <b>ETRIC +</b>                           |              |   |              |              |   |
| <b>FUNCTIONAL</b>                        |              |   |              |              |   |
| <b>MEASURES</b>                          |              |   |              |              |   |

|                                   |              |              |              |              |   |
|-----------------------------------|--------------|--------------|--------------|--------------|---|
| <b>Choukair et al.</b><br>(2017)  | Not reported | Not reported | Not reported | Not reported | 33 (4 adolescents and 29 adults) were on a PKU diet + protein substitute<br>16 (2 adolescents and 14 adults) did not follow a diet or protein substitute<br>3 adults had protein substitute only;<br>1 adolescent and 3 adults followed a PKU diet exclusively. |
| <b>Sumanszki et al.</b><br>(2020) | Not reported | Not reported | Not reported | Not reported |   |

**FUNCTIONAL + BIOCHEMICAL MEASURES**

|                                 |              |              |              |              |  |
|---------------------------------|--------------|--------------|--------------|--------------|--|
| <b>Mazzola et al.</b><br>(2015) | Not reported | Not reported | Not reported | Not reported |  |
|---------------------------------|--------------|--------------|--------------|--------------|--|

Note: Authors in italics are abstract only papers

\* V. R. Young and S. Borgonha, "Nitrogen and amino acid requirements: the Massachusetts Institute of Technology amino acid requirement pattern," The Journal of Nutrition, vol. 130, no.7, pp. 1841S–1849S, 2000

## Supplemental material: Search strategy

Database: MEDLINE (Ovid)

(exp Phenylketonurias/ OR Phenylketonurias.mp. OR PKU.mp. OR Hyperphenylalaninemia.mp. OR exp Phenylalanine Hydroxylase/ OR "Phenylalanine Hydroxylase".mp. OR "Phenylalanine Hydroxylase deficiency".mp.) **AND** (exp Nutritional Status/ OR "nutritional status".mp. OR "protein status".mp. OR exp Muscle Proteins/ OR "muscle proteins".mp. OR "protein metabolism".mp. OR "muscle protein metabolism".mp. OR exp Body Composition/ OR "body composition".mp. OR exp Muscle Strength/ OR "muscle strength".mp. OR "muscle function".mp. OR exp Prealbumin/ OR prealbumin.mp. OR Transthyretin.mp. OR exp Albumins/ OR albumin.mp. OR "3-methylhistidine concentrations".mp. OR exp Retinol-Binding Proteins/ OR "retinol-binding protein".mp. OR "urea production".mp. OR exp Nitrogen/ OR nitrogen.mp. OR exp Creatinine/ OR creatinine.mp. OR "VO2max" OR exp Physical Exertion/ OR "physical exertion".mp. OR exp Exercise Test/ OR "exercise test".mp.) **[Limit to English language]**