



King's Research Portal

DOI:

[10.1093/hmg/ddv472](https://doi.org/10.1093/hmg/ddv472)

Document Version

Peer reviewed version

[Link to publication record in King's Research Portal](#)

Citation for published version (APA):

Felix, J. F., Bradfield, J. P., Monnereau, C., Van Der Valk, R. J. P., Stergiakouli, E., Chesi, A., Gaillard, R., Feenstra, B., Thiering, E., Kreiner-Møller, E., Mahajan, A., Pitk??nen, N., Joro, R., Cavadino, A., Huikari, V., Franks, S., Groen-blokhuis, M. M., Cousminer, D. L., Marsh, J. A., ... Jaddoe, V. W. V. (2015). Genome-wide association analysis identifies three new susceptibility loci for childhood body mass index. *Human Molecular Genetics*, 25, 389-403. Advance online publication. <https://doi.org/10.1093/hmg/ddv472>

Citing this paper

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Research Portal

Take down policy

If you believe that this document breaches copyright please contact librarypure@kcl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

Genome-wide association analysis identifies three new susceptibility loci for childhood body mass index

Janine F. Felix^{1,2,3,4,*}, Jonathan P. Bradfield^{4,5}, Claire Monnereau^{1,2,3,4}, Ralf J.P. van der Valk^{4,6}, Evie Stergiakouli⁷, Alessandra Chesi⁸, Romy Gaillard^{1,2,3}, Bjarke Feenstra⁹, Elisabeth Thiering^{10,11}, Eskil Kreiner-Møller¹², Anubha Mahajan¹³, Niina Pitkänen^{14,15}, Raimo Joro¹⁶, Alana Cavadino^{17,18}, Ville Huikari¹⁹, Steve Franks²⁰, Maria M. Groen-Blokhuis²¹, Diana L. Cousminer²², Julie A. Marsh²³, Terho Lehtimäki^{24,25}, John A. Curtin²⁶, Jesus Vioque^{27,28}, Tarunveer S. Ahluwalia^{29,30,31}, Ronny Myhre³², Thomas S. Price³³, Natalia Vilor-Tejedor^{28,34,35}, Loïc Yengo^{36,37}, Niels Grarup³⁰, Ioanna Ntalla^{38,39}, Wei Ang²³, Mustafa Atalay¹⁶, Hans Bisgaard¹², Alexandra I. Blakemore⁴⁰, Amelie Bonnefond^{36,37}, Lisbeth Carstensen⁹, Bone Mineral Density in Childhood Study (BMDCS) Consortium[†], Early Genetics and Lifecourse Epidemiology (EAGLE) consortium, Johan Eriksson⁴¹, Claudia Flexeder¹⁰, Lude Franke⁴², Frank Geller⁹, Mandy Geserick^{43,44}, Anna-Liisa Hartikainen⁴⁵, Claire M. A. Haworth⁴⁶, Joel N. Hirschhorn^{47,48,49}, Albert Hofman^{1,3}, Jens-Christian Holm^{50,51}, Momoko Horikoshi^{13,52}, Jouke Jan Hottenga²¹, Jinyan Huang⁵³, Haja N. Kadarmideen⁵⁴, Mika Kähönen^{55,56}, Wieland Kiess⁴³, Hanna-Maaria Lakka¹⁶, Timo A. Lakka^{16,57,58}, Alexandra M. Lewin⁵⁹, Liming Liang^{60,61}, Leo-Pekka Lyytikäinen^{24,25}, Baoshan Ma⁶², Per Magnus⁶³, Shana E. McCormack^{8,64,65}, George McMahon⁷, Frank D. Mentch⁵, Christel M. Middeldorp²¹, Clare S. Murray²⁶, Katja Pahkala^{14,66}, Tune H. Pers^{47,48}, Roland Pfäffle^{43,67}, Dirkje S. Postma⁶, Christine Power¹⁸, Angela Simpson²⁶, Verena Sengpiel⁶⁸, Carla M. T. Tiesler^{10,11}, Maties Torrent^{28,69}, André G. Uitterlinden^{1,3,70}, Joyce B. van Meurs^{3,70}, Rebecca Vinding¹², Johannes Waage¹², Jane

Wardle⁷¹, Eleftheria Zeggini⁷², Babette S. Zemel^{65,73}, George V. Dedoussis³⁹, Oluf Pedersen³⁰, Philippe Froguel^{36,74}, Jordi Sunyer^{28,34,35,75}, Robert Plomin⁷⁶, Bo Jacobsson^{32,68}, Torben Hansen³⁰, Juan R Gonzalez^{28,34,35}, Adnan Custovic²⁶, Olli T. Raitakari^{14,77}, Craig E. Pennell²³, Elisabeth Widén²², Dorret I. Boomsma²¹, Gerard H. Koppelman⁷⁸, Sylvain Sebert^{19,79}, Marjo-Riitta Järvelin^{19,59,79,80,81}, Elina Hyppönen^{18,82,83}, Mark I. McCarthy^{13,52,84}, Virpi Lindi¹⁶, Niinikoski Harri⁸⁵, Antje Körner⁴³, Klaus Bønnelykke¹², Joachim Heinrich¹⁰, Mads Melbye^{9,86}, Fernando Rivadeneira^{1,3,70}, Hakon Hakonarson^{5,8,65}, Susan M. Ring^{7,87}, George Davey Smith⁷, Thorkild I.A. Sørensen^{7,30,88}, Nicholas J. Timpson^{7,89}, Struan F.A. Grant^{5,8,64,65,89}, Vincent W.V. Jaddoe^{1,2,3,89}, for the Early Growth Genetics (EGG) Consortium

¹The Generation R Study Group, Erasmus MC, University Medical Center Rotterdam, Rotterdam, the Netherlands

²Department of Pediatrics, Erasmus MC, University Medical Center Rotterdam, Rotterdam, the Netherlands

³Department of Epidemiology, Erasmus MC, University Medical Center Rotterdam, Rotterdam, the Netherlands

⁵Center for Applied Genomics, The Children's Hospital of Philadelphia, Philadelphia, PA, USA

⁶University of Groningen, University Medical Center Groningen, Department of Pulmonology, GRIAC (Groningen Research Institute for Asthma and COPD), Groningen, the Netherlands

⁷MRC Integrative Epidemiology Unit at the University of Bristol, University of Bristol, United Kingdom

⁸Division of Human Genetics, The Children's Hospital of Philadelphia, Philadelphia, PA, USA

⁹Department of Epidemiology Research, Statens Serum Institut, Copenhagen, Denmark

¹⁰Institute of Epidemiology I, Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, Germany

¹¹Division of Metabolic and Nutritional Medicine, Dr. von Hauner Children's Hospital, University of Munich Medical Center, Munich, Germany

¹²Copenhagen Prospective Studies on Asthma in Childhood, Health Sciences, University of Copenhagen, Danish Pediatric Asthma Center, Copenhagen University Hospital, Gentofte, Denmark

¹³Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, United Kingdom

¹⁴Research Centre of Applied and Preventive Cardiovascular Medicine, University of Turku, Finland

¹⁵Institute of Clinical Medicine, Neurology, University of Eastern Finland, Kuopio, Finland

¹⁶Institute of Biomedicine, Physiology, University of Eastern Finland, Kuopio, Finland

¹⁷Centre for Environmental and Preventive Medicine, Wolfson Institute of Preventive Medicine, Barts and the London School of Medicine and Dentistry, Queen Mary University of London, United Kingdom

¹⁸Population, Policy and Practice, UCL Institute of Child Health, University College London, United Kingdom

¹⁹Centre for Life Course Epidemiology, University of Oulu, Finland

²⁰Institute of Reproductive and Developmental Biology, Imperial College London, London, United Kingdom

- ²¹Dept Biol Psychology, VU University , Amsterdam; NCA Neuroscience Campus Amsterdam and EMGO+ Institute for Health and Care Research, Amsterdam, the Netherlands
- ²²Institute for Molecular Medicine, Finland (FIMM), University of Helsinki, Helsinki, Finland
- ²³School of Women's and Infants' Health, The University of Western Australia, Perth, Australia
- ²⁴Department of Clinical Chemistry, Fimlab Laboratories, Tampere, Finland
- ²⁵Department of Clinical Chemistry, University of Tampere School of Medicine, Tampere, Finland
- ²⁶Centre for Respiratory Medicine and Allergy, Institute of Inflammation and Repair, University of Manchester and University Hospital of South Manchester, Manchester Academic Health Sciences Centre, Manchester, United Kingdom
- ²⁷Universidad Miguel Hernandez, Elche-Alicante, Spain
- ²⁸CIBER Epidemiología y Salud Pública (CIBERESP), Spain
- ²⁹COPSAC, Copenhagen Prospective Studies on Asthma in Childhood, Herlev and Gentofte Hospital, University of Copenhagen, Copenhagen, Denmark
- ³⁰Novo Nordisk Foundation Centre for Basic Metabolic Research, Section of Metabolic Genetics, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark
- ³¹Steno Diabetes Center, Gentofte, Denmark
- ³²Department of Genes and Environment, Div of Epidemiology, Norwegian Institute of Public Health, Oslo, Norway
- ³³Department of Systems Pharmacology and Translational Therapeutics, University of Pennsylvania Perelman School of Medicine, USA
- ³⁴Centre for Research in Environmental Epidemiology (CREAL), Barcelona, Spain

- ³⁵Pompeu Fabra University (UPF), Barcelona, Spain
- ³⁶CNRS UMR8199, Pasteur Institute Lille, France
- ³⁷European Genomic Institute for Diabetes (EGID), Lille, France
- ³⁸Department of Health Sciences, University of Leicester, Leicester, United Kingdom
- ³⁹Department of Nutrition and Dietetics, School of Health Science and Education, Harokopio University, Athens, Greece
- ⁴⁰Section of Investigative Medicine, Division of Diabetes, Endocrinology, and Metabolism, Faculty of Medicine, Imperial College, London, United Kingdom
- ⁴¹National Institute for Health and Welfare, Helsinki, Finland
- ⁴²Department of Genetics, University of Groningen, University Medical Centre Groningen, Groningen, the Netherlands
- ⁴³Center of Pediatric Research, Dept. of Women's & Child Health, University of Leipzig, Germany
- ⁴⁴LIFE Child (Leipzig Research Center for Civilization Diseases), University of Leipzig, Germany
- ⁴⁵Institute of Clinical Medicine/Obstetrics and Gynecology, University of Oulu, Oulu, Finland
- ⁴⁶Department of Psychology, University of Warwick, United Kingdom
- ⁴⁷Division of Endocrinology and Center for Basic and Translational Obesity Research, Boston Children's Hospital, Boston, USA
- ⁴⁸Medical and Population Genetics Program, Broad Institute of MIT and Harvard, Cambridge, USA
- ⁴⁹Department of Genetics, Harvard Medical School, Boston, USA

- ⁵⁰The Children`s Obesity Clinic, Department of Pediatrics, Copenhagen University Hospital Holbæk, The Danish Childhood Obesity Biobank, Denmark
- ⁵¹Institute of Medicine, Copenhagen University, Copenhagen, Denmark
- ⁵²Oxford Centre for Diabetes, Endocrinology and Metabolism, University of Oxford, Oxford, United Kingdom
- ⁵³State Key Laboratory of Medical Genomics, Shanghai Institute of Hematology, Rui Jin Hospital Affiliated with Shanghai Jiao Tong University School of Medicine, Shanghai, China
- ⁵⁴Department of Veterinary Clinical and Animal Sciences, Faculty of Health and Medical Sciences, University of Copenhagen, Frederiksberg C, Denmark
- ⁵⁵Department of Clinical Physiology, Tampere University Hospital, Tampere, Finland
- ⁵⁶Department of Clinical Physiology, University of Tampere School of Medicine, Tampere, Finland
- ⁵⁷Kuopio Research Institute of Exercise Medicine, Kuopio, Finland
- ⁵⁸Department of Clinical Physiology and Nuclear Medicine, Kuopio University Hospital, Kuopio, Finland
- ⁵⁹Department of Epidemiology and Biostatistics, MRC Health Protection Agency (HPE) Centre for Environment and Health, School of Public Health, Imperial College London, United Kingdom
- ⁶⁰Department of Epidemiology, Harvard School of Public Health, Boston, USA
- ⁶¹Department of Biostatistics, Harvard School of Public Health, Boston, USA
- ⁶²College of Information Science and Technology, Dalian Maritime University, Dalian, Liaoning Province, China

- ⁶³Division of Epidemiology, Norwegian Institute of Public Health, Oslo, Norway
- ⁶⁴Division of Endocrinology, Children's Hospital of Philadelphia, Philadelphia, PA, USA
- ⁶⁵Department of Pediatrics, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, USA
- ⁶⁶Paavo Nurmi Centre, Sports & Exercise Medicine Unit, Department of Health and Physical Activity, University of Turku, Finland
- ⁶⁷CrescNet, Medical Faculty, University of Leipzig, Germany
- ⁶⁸Department of Obstetrics and Gynecology, Sahlgrenska Academy, Sahlgrenska University Hospital, Gothenburg, Sweden
- ⁶⁹Area de Salut de Menorca, ib-salut, Menorca, Spain
- ⁷⁰Department of Internal Medicine, Erasmus MC, University Medical Center Rotterdam, Rotterdam, the Netherlands
- ⁷¹Department of Epidemiology and Public Health, University College London, United Kingdom
- ⁷²Wellcome Trust Sanger Institute, The Morgan Building, Wellcome Trust Genome Campus, Hinxton, Cambridgeshire, United Kingdom
- ⁷³Division of Gastroenterology, Hepatology and Nutrition, The Children's Hospital of Philadelphia, Philadelphia, PA, USA
- ⁷⁴Department of Genomics of Common Disease, School of Public Health, Imperial College London, Hammersmith Hospital, London, United Kingdom
- ⁷⁵IMIM (Hospital del Mar Medical Research Institute), Barcelona, Spain
- ⁷⁶King's College London, MRC Social, Genetic and Developmental Psychiatry Centre, Institute of Psychiatry, Psychology & Neuroscience, De Crespigny Park, London, United Kingdom

- ⁷⁷Department of Clinical Physiology and Nuclear Medicine, Turku University Hospital, Turku
Finland
- ⁷⁸University of Groningen, University Medical Center Groningen, Department of Pediatric
Pulmonology and Pediatric Allergology, Beatrix Children's Hospital, GRIAC (Groningen
Research Institute for Asthma and COPD), Groningen, the Netherlands
- ⁷⁹Biocenter Oulu, University of Oulu, Finland
- ⁸⁰Unit of Primary Care, Oulu University Hospital, Oulu, Finland
- ⁸¹Department of Children and Young People and Families, National Institute for Health and
Welfare, Oulu, Finland
- ⁸²School of Population Health and Sansom Institute, University of South Australia, Adelaide,
Australia
- ⁸³South Australian Health and Medical Research Institute, Adelaide, Australia
- ⁸⁴Oxford National Institute for Health Research (NIHR) Biomedical Research Centre, Churchill
Hospital, Oxford, United Kingdom
- ⁸⁵University of Turku, Turku University Hospital, Department of Pediatrics, Turku, Finland
- ⁸⁶Department of Medicine, Stanford University School of Medicine, Stanford, California, USA
- ⁸⁷Avon Longitudinal Study of Parents and Children (ALSPAC), School of Social and
Community Medicine, University of Bristol, Bristol, United Kingdom
- ⁸⁸Institute of Preventive Medicine, Bispebjerg and Frederiksberg Hospital, The Capital Region,
Copenhagen, Denmark
- ⁹⁰Division of General and Community Pediatrics, Cincinnati Children's Hospital Medical Center,
Cincinnati, OH, USA
- ⁹¹Division of Endocrinology, Department of Medicine, Creighton University, Omaha, NB, USA

⁹²Department of Radiology, Children's Hospital Los Angeles, Los Angeles, CA, USA

⁹³Division of Pediatric Endocrinology, Diabetes, and Metabolism, Department of Pediatrics,
Columbia University Medical Center, New York, NY, USA

*Corresponding author: Janine F. Felix, Department of Epidemiology, Room Na-2906, Erasmus
MC, University Medical Center Rotterdam, P.O.Box 2040, 3000 CA Rotterdam, the
Netherlands, Ph. +31 10 70 43 997, Fax +31 10 70 44657, j.felix@erasmusmc.nl

⁴These first authors contributed equally

⁸⁹These last authors contributed equally

Abstract

A large number of genetic loci are associated with adult body mass index. However, the genetics of childhood body mass index are largely unknown. We performed a meta-analysis of genome-wide association studies of childhood body mass index, using sex- and age-adjusted standard deviation scores. We included 35,668 children from 20 studies in the discovery phase and 11,873 children from 13 studies in the replication phase. In total, 15 loci reached genome-wide significance (P -value $< 5 \times 10^{-8}$) in the joint discovery and replication analysis, of which 12 are previously identified loci in or close to *ADCY3*, *GNPDA2*, *TMEM18*, *SEC16B*, *FAIM2*, *FTO*, *TFAP2B*, *TNNI3K*, *MC4R*, *GPR61*, *LMX1B* and *OLFM4* associated with adult body mass index or childhood obesity. We identified three novel loci: rs13253111 near *ELP3*, rs8092503 near *RAB27B*, and rs13387838 near *ADAM23*. Per additional risk allele, body mass index increased 0.04 Standard Deviation Score (SDS) (Standard Error (SE) 0.007), 0.05 SDS (SE 0.008) and 0.14 SDS (SE 0.025), for rs13253111, rs8092503, and rs13387838, respectively. A genetic risk score combining all 15 SNPs showed that each additional average risk allele was associated with a 0.073 SDS (SE 0.011, P -value = 3.12×10^{-10}) increase in childhood body mass index in a population of 1,955 children. This risk score explained 2% of the variance in childhood body mass index. This study highlights the shared genetic background between childhood and adult body mass index and adds three novel loci. These loci likely represent age-related differences in strength of the associations with body mass index.

Introduction

Childhood obesity is an important public health problem with severe consequences, including an increased risk of premature death (1-5). Body mass index (BMI) has a strong genetic component with some reported heritability estimates being over 80% (6-8). Large genome-wide association studies (GWAS) have revealed many genetic loci associated with BMI or adiposity in adults (9-13). However, the genetic loci underlying BMI in children are less well known. The biological background of BMI may differ between children and adults. In addition, it may be that the relative contributions of the same genetic loci differ depending on age, for example due to different gene-environment-interactions or body fat distributions (6, 14, 15). A limited number of loci has been identified to associate with dichotomous definitions of childhood obesity (16-18). Also, the roles of specific known adult loci for BMI, such as *FTO* and *ADCY3*, have been described in children (13, 19). The age-specific effects are illustrated by longitudinal studies on the effects of the well-known adult BMI increasing risk allele of *FTO* with BMI throughout childhood (15). It has been reported that adult BMI increasing risk allele is associated with lower BMI in infancy, an earlier adiposity rebound and a higher BMI from the age of five years onwards (14, 15, 20). To date, studies did not present a large GWAS meta-analysis on the full spectrum of childhood BMI (13, 16-19).

To identify genetic loci influencing childhood BMI, we meta-analyzed 20 GWAS with a total of 35,668 children of European ancestry, combining data for around 2.5 million single nucleotide polymorphisms (SNPs) imputed to the HapMap imputation panel. We used as outcome sex- and age-adjusted standard deviation scores at the oldest age between 2 and 10 years.

Results

Study characteristics are shown in Supplementary Material, Table S1. Childhood BMI was transformed into sex- and age-adjusted standard deviation scores (SDS) (LMS growth; Pan H, Cole TJ, 2012. <http://www.healthforallchildren.co.uk>).

Meta-analysis of genome-wide association studies

Inverse-variance weighted fixed-effects meta-analysis revealed 861 SNPs with genome-wide significant or suggestive P -values ($< 5 \times 10^{-6}$). Two SNPs with high heterogeneity were not followed up (I^2 values of 89.4 and 96.0), leaving 859 SNPs representing 43 loci. A locus was defined as a region of 500 kb to either side of the most significant SNP. The Manhattan and Quantile-Quantile plots of the discovery meta-analysis are shown in Figure 1 and Supplementary Material, Figure S1, respectively. The lambda for the discovery meta-analysis was 1.10. LD score regression analysis showed that this slight inflation was mainly due to polygenicity of the trait, rather than to population stratification, cryptic relatedness or other confounding factors (intercept 1.01). Individual study lambdas are shown in Supplementary Material, Table S2. All 43 loci were taken forward for replication in a sample of 11,873 children from 13 studies. Table 1 and Supplementary Material, Tables S3 and S4 show the results of the discovery, replication, and joint analyses for the 43 genome-wide and suggestive loci.

In total, 15 of these reached genome-wide significance in the joint analysis. Twelve out of these 15 had been reported previously for related phenotypes. SNPs in or close to *ADCY3*, *GNPDA2*, *TMEM18*, *SEC16B*, *FAIM2*, *FTO*, *TFAP2B*, *TNNI3K*, *MC4R*, *GPR61*, *LMX1B* and *OLFM4* are associated with adult BMI or childhood obesity (11, 13, 16). We identified three

novel loci: rs13253111 near *ELP3*, rs8092503 near *RAB27B*, and rs13387838 near *ADAM23*. Per additional risk allele, BMI increased 0.04 Standard Deviation Score (SDS) (Standard Error (SE) 0.007), 0.05 SDS (SE 0.008) and 0.14 SDS (SE 0.025), for rs13253111, rs8092503, and rs13387838, respectively. Figure 2 and Supplementary Material, Figure S2 show the regional plots and the forest plots, respectively, for these loci.

Genetic risk score

We combined the 15 identified genome-wide significant SNPs into a genetic risk score that summed the number of BMI-increasing alleles weighted by their betas from the discovery analysis and rescaled to a range of 0 to 30, which is the maximum number of risk alleles. The risk score was associated with childhood BMI (P -value = 3.12×10^{-10}) in 1,955 children from the PIAMA Study, one of our largest replication cohorts. For each additional average risk allele in the score, childhood BMI increased by 0.073 SDS (SE 0.011) (Figure 3). This risk score explained 2.0% of the variance in childhood BMI.

Associations with adult body mass index and childhood obesity

The genetic correlation between childhood BMI and adult BMI was 0.73. A lookup of the 15 SNPs associated with childhood BMI in a recently published GWAS meta-analysis on adult BMI in more than 300,000 participants, revealed that all SNPs showed evidence for association were nominally significantly associated with adult BMI, with P -values of 0.005, 5.76×10^{-5} and 0.003 for the novel SNPs rs13253111, rs8092503 and rs13387838, respectively. Also, the direction of the effect estimates for all 15 SNPs was the same in children and adults (Supplementary

Material, Table S5) (11). The 15 SNPs found in this study explained 0.94% of the variance in adult BMI in the GIANT consortium (11).

A reverse lookup in our dataset of the 97 known genome-wide significant loci previously reported to be associated with adult BMI showed that 22 out of the 97 loci were significantly associated with childhood BMI index, using a Bonferroni-adjusted P -value cutoff of 5.2×10^{-4} for 97 SNPs. A total of 50 out of the 97 known adult BMI SNPs were nominally associated with childhood BMI (P -value <0.05). The direction of the effect estimates was the same in adults and children for 86 SNPs (P -value binomial sign test $<1.0 \times 10^{-4}$; Supplementary Material, Table S6).

We looked up the association of the three novel loci in a GWAS meta-analysis of childhood obesity. In this study, childhood obesity cases were defined as having a BMI $\geq 95^{\text{th}}$ percentile, whereas childhood normal weight controls were defined as having a BMI $< 50^{\text{th}}$ percentile. This meta-analysis included 22 studies, of which 16 were also included in our current meta-analysis. All three SNPs were associated with childhood obesity (P -values 0.01, 0.005, and 6.0×10^{-4} for rs13253111, rs8092503, and rs13387838, respectively) (16).

Functional analysis

To explore functionality, we first analyzed if the 15 identified SNPs affect messenger RNA expression (eQTLs). We analyzed eQTLs from peripheral blood samples from 5,311 individuals, which revealed two *cis*-eQTLs (false discovery rate (FDR) P -value <0.05) for rs11676272, the top SNP in one of the previously identified loci (*ADCY3*). One of these eQTLs was for *ADCY3*, and one was for *DNAJC27* (21). Also, we found a *cis*-eQTL for *FAM125B* for rs3829849, which is located in *LMX1B* (Supplementary Material, Table S76). eQTL analysis in adipose tissue, a more specific target tissue in relation to BMI, from 856 healthy female twins in the MuTHER

resource in Genevar revealed two significant *cis*-eQTLs (distance to SNP < 1 Mb) for rs11676272, for transcripts of *ADCY3* and *POMC*, with a Bonferroni-corrected *P*-value of < 0.003 (22, 23). The association of rs11676272 with expression of *ADCY3* was also validated in a second eQTL analysis in a smaller set of 206 lymphoblastoid cell lines (24). We did not identify eQTLs related to our three novel loci.

Second, we performed functional analyses with the tool Data-Driven Expression Prioritized Integration for Complex Traits (DEPICT) using all SNPs with a *P*-value < 1×10^{-5} in the discovery analysis (see Materials and methods for details) (25). Gene prioritization analysis did not show prioritized genes, nor did the gene set enrichment analysis reveal evidence for enriched reconstituted gene sets and genes near the associated SNPs were not found to enrich for expression in a panel of 2009 tissue and cell types (FDR < 0.05; Supplementary Material, Tables S87a, b and c).

Discussion

In this GWAS meta-analysis of childhood BMI among more than 47,000 children, we identified 15 genome-wide significant loci, of which three loci, rs13253111 near *ELP3*, rs8092503 near *RAB27B*, and rs13387838 near *ADAM23*, have not been associated with adiposity related phenotypes before.

Large GWAS have revealed many genetic loci associated with BMI or adiposity in adults (9-13). A recent meta-analysis in up to 339,224 individuals identified 97 BMI-associated loci, explaining 2.7% of the adult BMI variation. Pathway analyses showed that the central nervous system may play a large role in obesity susceptibility. The number of identified loci associated with BMI or obesity in childhood is scarce. Of the total of 15 loci associated with childhood BMI

in the current study, 12 have previously been associated with adiposity outcomes in adults or children. All 12 loci are known to be associated with adult BMI (11). Also, eight loci, including those in or near *ADCY3* (annotated to the nearby gene *POMC* in the previous paper), *TMEM18*, *SEC16B*, *FAIM2*, *FTO*, *TNNI3K*, *MC4R* and *OLFM4*, have previously been associated with childhood obesity (16). All three novel loci were nominally associated with the more extreme outcome of childhood obesity in a largely overlapping population of child cohorts (16).

A recent meta-analysis of two studies showed that the known loci *FTO*, *MC4R*, *ADCY3*, *OLFM4* are associated with BMI trajectories in childhood (26). Their findings also suggested that a locus annotated to *FAM120AOS* influences childhood BMI, which could not be replicated in the current study. The lead SNP in this locus, rs944990, had a *P*-value of 1.61×10^{-5} in the current analysis. These findings suggest that the overlap between the genetic background of childhood and adult BMI is relatively large, but not complete.

Rs7550711 represents one of the 12 identified loci known to be associated with BMI or obesity in adults and children. Rs7550711 is a proxy for rs17024258 and rs17024393 (R^2 0.8 with both SNPs), which have previously been associated with adult obesity and BMI, respectively, and annotated with the *GNAT2* gene. However, our proxy resides in *GPR61*, G protein-coupled receptor 61, the biology of which may be more relevant to BMI. Gpr61-deficient mice are obese and have hyperphagia, suggesting the role of Gpr61 in food intake regulation (27). Further studies, including expression studies in relevant human tissues, are needed to establish the causal genes underlying this association.

We identified three loci, rs13253111 near *ELP3*, rs8092503 near *RAB27B*, and rs13387838 near *ADAM23*, which have not been associated with adiposity related phenotypes before in adulthood or childhood. The nearest genes to the novel loci have varying functions. *ELP3*, Elongator

Acetyltransferase Complex, subunit 3, has a potential role in the migration of cortical projection neurons and in paternal demethylation after fertilization in mice (28-30). *RAB27B*, RAS-associated protein *RAB27B*, encodes a membrane-bound protein with a role in secretory vesicle fusion and trafficking. It has been associated with pituitary hormone secretion, regulation of exocytosis of digestive enzyme containing granules from pancreatic acinar cells, and with gastric acid secretion (31-33). Expression of *ADAM23*, A Disintegrin And Metalloproteinase Domain 23, may influence tumor progression and brain development (34, 35). It has also been described to be expressed in mouse adipose tissue and to have a potential role in adipogenesis *in vitro* (36). Two of our novel loci, rs13253111 near *ELP3* and rs13387838 near *ADAM23*, are close to rs4319045 and rs972540, respectively. Both these SNPs were reported as subthreshold results in the GWAS meta-analysis on adult BMI (11). However, the linkage disequilibrium between the SNPs in both pairs is very low ($R^2 \leq 0.1$ for both) suggesting that these SNPs may represent different signals. It is important to note that, although both SNPs reached genome-wide significance in the joint discovery and replication analysis, the P-values in the replication stage were non-significant. This lack of significance may be due to the smaller sample size and lower power. Also, the joint P-values were slightly higher than the discovery P-values. Heterogeneity between the discovery and the replication stages was low to moderate, with I^2 values of 61.1 and 27.8 for rs13253111 and rs13387838, respectively (P-values > 0.01 for both). These two signals need to be interpreted with some caution and further studies with larger sample sizes are needed to fully clarify the role of variants in these regions in the physiology of BMI.

Functional analysis showed *cis*-eQTLs for the lead SNPs in two of the known loci. Rs11676272 was associated with eQTLs in *ADCY3* and *DNAJC27*, also known as *RBJ*. Both these genes have been associated with adult BMI before and the association of rs11676272 with

expression of *ADCY3* has been previously described in childhood BMI (11, 13, 37). Rs3829849 was associated with an eQTL in *FAM125B*, or *MVB12B*, multivesicular body subunit 12B. This gene encodes a component of ESCRT-I (endosomal sorting complex required for transport I), a plasma membrane complex with a role in vesicular trafficking, was recently described to be associated with intra-ocular pressure (38). However, the LD of our SNPs with the peak markers for the *DNAJC27* (R^2 0.11) and the *FAM125B* (R^2 0.03) transcripts was low. Our analysis using DEPICT did not show enriched gene sets. This may reflect the relatively limited sample size in our analysis. Further studies are needed to determine the potential functional impact of all SNPs associated with childhood BMI.

Using LD score regression analysis with our meta-analysis results and the results from the recently published GWAs meta-analysis on adult BMI as input, we found that the genetic correlation between childhood and adult BMI was high (11, 39). The variance in adult BMI explained by the 15 SNPs identified in this study was lower than in children. The novel SNPs reported in this study may represent loci that specifically influence childhood BMI, but not adult BMI. An alternative explanation is that the effect sizes of these loci may be larger in children than in adults, which may explain the discovery in childhood studies but not in adult studies (11). The large overlap between childhood and adult BMI loci suggests that many of these loci may not represent childhood-specific effects, but rather involvement of the same loci with differential effect sizes at different ages. Age-specific effects of genetic variants associated with BMI in children have been described for the *FTO* locus (15). However, longitudinal studies with multiple measurements of BMI are needed to confirm and quantify such varying effects with age. In discussing the genetic overlap between childhood and adult BMI, it needs to be noted that, because of the differences in body proportions and body fat distribution, childhood BMI

may be a different phenotype as compared to adult BMI. Our outcome was the conventional measure of BMI calculated as weight/height². Especially in early childhood, higher orders of magnitude for height may be more appropriate. Results from a previously published GWAS study on childhood BMI in two of the cohorts included in the current meta-analysis suggest that the results for SNPs close to *ADCY3* are different when higher orders of magnitude for height are being used (37). Further studies are needed to identify loci related to more specific and directly assessed measures of adiposity and body fat distribution in young children.

In conclusion, we identified 15 loci associated with childhood BMI, of which three are novel. Our results highlight a considerable shared genetic background between childhood and adult BMI. The novel BMI-related loci may reflect childhood-specific genetic associations or differences in strength of associations between age groups.

Materials and methods

Study populations

Characteristics of each discovery and replication study population can be found in Supplementary Material, Table S1 and Supplementary Methods. The discovery analysis included 20 studies with an age range from 3 to 10 years: the Avon Longitudinal Study of Parents and Children (ALSPAC, 6887 children), the Children's Hospital of Philadelphia (CHOP, 2456 children), the Copenhagen Studies on Asthma in Childhood 2000 birth cohort (COPSAC2000, 309 children), the Danish National Birth Cohort (DNBC, 1020 children), the Generation R Study (GenerationR, 2226 children), the GOYA Study (GOYA, 199 children), the Helsinki Birth Cohort Study (HBCS, 1674 children), the Infancia y Medio Ambiente Project (INMA, 756 children), the Leipzig study (Leipzig, 555 children), the Lifestyle – Immune System – Allergy

Study plus German Infant Study on the influence of Nutrition Intervention (LISA+GINI, 1147 children), the Manchester Asthma and Allergy Study (MAAS, 801 children), the Norwegian Mother and Child Cohort Study (MoBa, 126 children), the Northern Finland Birth Cohort 1966 (NFBC 1966, 3948 children), the Northern Finland Birth Cohort 1986 (NFBC 1986, 4000 children), the Netherlands Twin Register (NTR, 1810 children), the Physical Activity and Nutrition in Children Study (PANIC, 423 children), the Western Australian Pregnancy Cohort (Raine) Study (Raine, 1458 children), the Special Turku coronary Risk factor Intervention Project (STRIP, 569 children), the Young Finns Study (YFS, 1134 children), the British 1958 Birth Cohort Study, with two subcohorts which were entered into the meta-analysis separately (1958BC-T1DGC, 1974 children, and 1958BC-WTCCC2, 2196 children).

We included 13 replication studies. Eleven of these were cohort studies: 574 children from the Copenhagen Studies on Asthma in Childhood 2010 birth cohort (COPSAC2010), 676 additional children from the DNBC, 386 additional children from LISA+GINI, 3152 children from the TEDS Study, 1955 children from the Prevention and Incidence of Asthma and Mite Allergy birth cohort study (PIAMA), 1665 children from the BREATHE Study, 447 children from the Bone Mineral Density in Childhood Study (BMDCS), 200 children from the TEENs of Attica: Genes and Environment (TEENAGE) study, additional imputed data on 857 children from the Leipzig Study, 480 additional children from PANIC, and additional imputed data for 569 children from STRIP. We also included two obesity case-control studies in the replication: the Danish Childhood Obesity Biobank (306 cases, 158 controls) and the French Young Study (304 cases, 144 controls). In the BREATHE Study, information was available about six SNPs only (rs8046312, rs12429545, rs13130484, rs3845265, rs543874, rs8084077).

All included children were of European ethnic origin. Sex- and age-adjusted standard deviation scores were created for BMI at the latest time point (oldest age, if multiple measurements existed) between 2 and 10 years using the same software across all studies (LMS growth; Pan H, Cole TJ, 2012. Available from: <http://www.healthforallchildren.co.uk>). Syndromic cases of obesity and children of non-European ethnic origin were excluded. In the case of twin pairs, only one twin was included, either randomly or based on genotyping or imputation quality.

Statistical approach

Cohort-specific genome-wide association analyses were first run in the discovery cohorts, using high-density Illumina or Affymetrix SNP arrays, followed by imputation to the HapMap CEU release 22 imputation panel. The MAAS study imputed to the combined 1000 Genomes (1000G) Pilot + HapMap 3 (release June 2010/Feb 2009) panel. Before imputation, studies applied study-specific quality filters on samples and SNP call rate, minor allele frequency and Hardy-Weinberg disequilibrium (see Supplementary Material, Table S1 for details). Leipzig (discovery sample), NFBC1986, STRIP (discovery sample), and PANIC (discovery sample) contributed unimputed data from the MetaboChip. Linear regression models assuming an additive genetic model were run in each study, to assess the association of each SNP with SDS-BMI, adjusting for principal components if this was deemed needed in the individual studies. As SDS-BMI is age- and sex-specific, no further adjustments were made. Before the meta-analysis, we applied quality filters to each study, filtering out SNPs with a minor allele frequency below 1% and SNPs with poor imputation quality (MACH $r2_hat \leq 0.3$, IMPUTE $proper_info \leq 0.4$ or $info \leq 0.4$). For studies contributing unimputed metabochip data to the discovery analysis, we excluded SNPs

with a SNP call rate < 0.95 or with a Hardy Weinberg Equilibrium P -value of ≤ 0.00001 . We performed fixed effects inverse-variance weighted meta-analysis of all discovery samples using Metal (40). Genomic control was applied to every study before the meta-analysis. Individual study lambdas ranged from 0.985 to 1.077 (Supplementary Material, Table S2). The lambda of the discovery meta-analysis was 1.10. After the meta-analysis, SNPs for which information was available in only one study were removed.

The final dataset consisted of 2,499,691 autosomal SNPs. The most significant SNP for each of 43 genome-wide significant or suggestive loci (P -value $< 5 \times 10^{-6}$) was taken forward for replication in 13 replication cohorts. A locus was defined as a region 500 kb to either side of the most significant SNP. All replication cohorts had *in silico* data available. One of them only had non-imputed data (BREATHE), two (TEENAGE and TEDS) had data imputed to HapMap release 22, one cohort (PANIC) used exome chip data and the other nine performed imputation to 1000G. The replication samples of the STRIP and Leipzig studies only contributed 20 and 21 imputed SNPs, respectively, as the unimputed SNPs were part of the discovery analysis). Fixed effects inverse-variance meta-analysis was performed for these 43 SNPs combining the discovery samples and all replication samples, giving a joint analysis beta, standard error and P -value (Table 1 and Supplementary Material, Table S2).

Sensitivity analyses

Allele frequency differences between the discovery and the replication samples were small and stayed within a range of seven percentage points for all SNPs, except for rs1573972, which had a minor allele frequency of 9% in the discovery analysis and 28% in the replication analysis. This was likely due to the inclusion of one study (MAAS) that had imputed to the combined

HapMap+1000G panel, whereas all other studies with imputed data had imputed to HapMap. To increase homogeneity, we performed several sensitivity analyses. First, we reran the discovery meta-analysis excluding the MAAS study. This analysis did not materially change our findings, with one additional SNP (rs10055577) reaching the subthreshold level of significance (P -value = 1.10×10^{-6}) and five SNPs (rs4870949, rs1838856, rs633143, rs10866069, and rs1573972) losing significance. None of these five SNPs had replicated in the primary analysis. Second, we reran the replication and joint meta-analysis including only those cohorts that imputed to 1000G. Results of this analysis were very similar to the primary analysis, with two additional replicated SNPs, rs17309930 near *BDNF* and rs13107325 in *SLC39A8*. Both of these are known loci for adult BMI (11, 13). Third, we reran the replication including only the HapMap-imputed and unimputed studies (TEDS, TEENAGE, and BREATHE). The results were very similar to those using all studies, with rs4870949 and rs2590942 now passing the significance threshold and rs8092503 and rs3829849 now just above it (results not shown). Rs1573972 was not replicated in any of the analyses. As results of the third and fourth sensitivity analyses were very similar to those including all replication cohorts, we used the latter as our main analysis for reasons of power.

Genetic risk score and percentage of variance explained

A weighted risk score was computed as the sum of the number of SDS-BMI-increasing alleles (dosage) weighted by the effect sizes from the discovery meta-analysis. Then, the score was rescaled to range from zero to the maximum number of SDS-BMI increasing alleles (30 alleles for 15 SNPs) and rounded to the nearest integer. The association of the risk score with SDS-BMI was assessed in one of the largest replication cohorts (PIAMA, N=1955) by running a linear

regression model. The variance in SDS-BMI explained by the risk score was estimated by the unadjusted R^2 of this model. The percentage of variance in adult BMI explained by the 15 SNPs was calculated using the published data from the recently published large meta-analysis of GWAs studies on adult BMI (11). For each SNP, the variance explained was calculated as: $2 * (\text{adult effect size}^2) * \text{MAF} * (1 - \text{MAF})$ and these variances were then summed to give the total percentage of variance in adult BMI explained by the 15 SNPs (11, 41).

LD score regression

LD score regression was used with the standard settings (39). Changing the minor allele frequency filter from 0 to 0.05 did not change the results. Therefore, we report the results of the unfiltered analysis only.

eQTL analysis

eQTL analysis was conducted using most significant SNP from each of the 15 genome-wide significant loci from the joint analysis. There was no linkage disequilibrium between these SNPs. First, we assessed whether the top SNPs or their proxies, identified on the basis of $R^2 > 0.7$, were associated with gene expression in whole-blood cells in a sample of 5,311 individuals (21). Expression in this dataset was assessed using Illumina Whole-Genome Expression BeadChips (HumanHT-12). eQTLs were deemed *cis* when the distance between the SNP chromosomal position and the probe midpoint was less than 250 kb. eQTLs were mapped using Spearman's rank correlation, using imputation dosage values as genotypes. A n FDR P -value of < 0.05 was considered significant. Second, the 15 SNPs were introduced to the online eQTL database Genevar (www.sanger.ac.uk/resources/software/genevar) to explore their associations with

expression transcripts of genes in proximity (<1Mb distance) to the SNP in adipose tissue from 856 healthy female twins of the MuTHER resource (22, 23). We used Bonferroni correction for the significance threshold (P -value < 0.003).

Data-driven Expression Prioritized Integration for Complex Traits (DEPICT)(42)

DEPICT was run using SNPs with P -value < 10^{-5} as yielding 56 independent DEPICT loci comprising 100 genes. DEPICT was run using default settings, that is using 500 permutations for bias adjustment, 20 replications for false discovery rate estimation, normalized expression data from 77,840 Affymetrix microarrays for gene set reconstitution (see (43) for details), 14,461 reconstituted gene sets for gene set enrichment analysis, and testing 209 tissue/cell types assembled from 37,427 Affymetrix U133 Plus 2.0 Array samples for enrichment in tissue/cell type expression (42).

Acknowledgements

ALSPAC Study

The MRC IEU is supported by the Medical Research Council and the University of Bristol (grant code MC_UU_12013/1-9). The authors are extremely grateful to all the families who took part in the ALSPAC study, the midwives for their help in recruiting them, and the whole ALSPAC team, which includes interviewers, computer and laboratory technicians, clerical workers, research scientists, volunteers, managers, receptionists, and nurses. The UK Medical Research Council and the Wellcome Trust (Grant ref: 102215/2/13/2) and the University of Bristol provide core support for ALSPAC. ALSPAC GWAS data was generated by Sample Logistics and Genotyping Facilities at the Wellcome Trust Sanger Institute and LabCorp (Laboratory

Corporation of America) supported by 23andMe. The MRC IEU is supported by the Medical Research Council and the University of Bristol (grant code MC_UU_12013/1-9).

1958BC-T1DGC and 1958CB-WTCCC

DNA collection was funded by MRC grant G0000934 and cell-line creation by Wellcome Trust grant 068545/Z/02. This research used resources provided by the Type 1 Diabetes Genetics Consortium, a collaborative clinical study sponsored by the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK), National Institute of Allergy and Infectious Diseases, National Human Genome Research Institute, National Institute of Child Health and Human Development, and Juvenile Diabetes Research Foundation International (JDRF) and supported by U01 DK062418. This study makes use of data generated by the Wellcome Trust Case-Control Consortium. A full list of investigators who contributed to generation of the data is available from the Wellcome Trust Case-Control Consortium website. Funding for the project was provided by the Wellcome Trust under the award 076113. The work was supported by the Department of Health Policy Research Programme through the Public Health Research Consortium (PHRC). Information about the wider programme of the PHRC is available from <http://phrc.lshtm.ac.uk>. Great Ormond Street Hospital/University College London, Institute of Child Health and Oxford Biomedical Research Centre, University of Oxford receive a proportion of funding from the Department of Health's National Institute for Health Research (NIHR) ('Biomedical Research Centres' funding). This paper presents independent research and the views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR, or the Department of Health. Mark I. McCarthy acknowledges support from the EU Framework 7 ENGAGE (HEALTH-F4-2007-201413) and The Wellcome Trust (098381 and 090367). Mark I. McCarthy is a Wellcome Trust Senior Investigator and a NIHR Senior Investigator.

BREATHE

The research leading to these results has received funding from the European Research Council under the ERC Grant Agreement number 268479 – the BREATHE project. We are acknowledged with all the families participating into the study for their altruism and particularly to the schools Antoni Brusi, Baloo, Betània – Patmos, Centre d'estudis Montseny, Collegi Shalom, Costa i Llobera, El sagrer, Els Llorers, Escola Pia de Sarrià, Escola Pia Balmes, Escola concertada Ramon Llull, Escola Nostra Sra. de Lourdes, Escola Tècnica Professional del Clot, Ferran i Clua, Francesc Macià, Frederic Mistral, Infant Jesús, Joan Maragall, Jovellanos, La Llacuna del Poblenou, Lloret, Menéndez Pidal, Nuestra Señora del Rosario, Miralletes, Ramon Llull, Rius i Tauler, Pau Vila, Pere Vila, Pi d'en Xandri, Projecte, Prosperitat, Sant Ramon Nonat - Sagrat Cor, Santa Anna, Sant Gregori, Sagrat Cor Diputació, Tres Pins, Tomàs Moro, Torrent d'en Melis, Virolai.

BMDCS

The study was funded by R01 HD58886 and R01 HD076321 and the Eunice Kennedy Shriver National Institute of Child Health and Human Development (NICHD) contracts (N01-HD-1-3228, -3329, -3330, -3331, -3332, -3333) and the CTSA program Grant 8 UL1 TR000077.

CHOP

We thank the network of primary care clinicians, their patients and families for their contribution to this project and clinical research facilitated through the Pediatric Research Consortium (PeRC) at The Children's Hospital of Philadelphia. Rosetta Chiavacci, Elvira Dabaghyan, Hope Thomas, Kisha Harden, Andrew Hill, Kenya Fain, Crystal Johnson-Honesty, Cynthia Drummond, Shanell Harrison and Sarah Wildrick, Cecilia Kim, Edward Frackelton, George Otieno, Kelly Thomas,

Cuiping Hou, Kelly Thomas and Maria L. Garris provided expert assistance with genotyping or data collection and management. We would also like to thank Smari Kristinsson, Larus Arni Hermannsson and Asbjörn Krisbjörnsson of Raförinn ehf for their extensive software design and contribution. This research was financially supported by an Institute Development Award from the Children's Hospital of Philadelphia, a Research Development Award from the Cotswold Foundation and NIH grant R01 HD056465.

COPSAC2000 and COPSAC2010

The authors gratefully express their gratitude to the children and families of the COPSAC cohort studies for all their support and commitment, and we acknowledge and appreciate the unique efforts of the COPSAC research team. COPSAC is funded by private and public research funds. The Lundbeck Foundation; the Pharmacy Foundation of 1991; Augustinus Foundation; the Danish Medical Research Council; and The Danish Pediatric Asthma Centre provided the core support for the study. For full list please see copsac.com.

DNBC

Support for the Danish National Birth Cohort was obtained from the Danish National Research Foundation, the Danish Pharmacists' Fund, the Egmont Foundation, the March of Dimes Birth Defects Foundation, the Augustinus Foundation and the Health Fund of the Danish Health Insurance Societies. The generation of GWAS genotype data for the DNBC samples was carried out within the GENEVA consortium with funding provided through the NIH Genes, Environment and Health Initiative (GEI) (U01HG004423). Assistance with phenotype harmonization and genotype cleaning, as well as with general study coordination, was provided by the GENEVA Coordinating Center (U01HG004446). Genotyping was performed at Johns

Hopkins University Center for Inherited Disease Research, with support from the NIH GEI (U01HG004438). Bjarke Feenstra is supported by an Oak Foundation Fellowship.

French Young

The authors thank the subjects and families who participated in this study. This work was supported by grants from the Agence Nationale de la Recherche, the Conseil Régional Nord-Pas de Calais as part of Fonds Européen de Développement Economique et Regional, Genome Quebec as part of Genome Canada and the UK MRC.

Generation R

The Generation R Study is conducted by the Erasmus Medical Center in close collaboration with the School of Law and Faculty of Social Sciences of the Erasmus University Rotterdam, the Municipal Health Service Rotterdam area, Rotterdam, the Rotterdam Homecare Foundation, Rotterdam and the Stichting Trombosedienst & Artsenlaboratorium Rijnmond (STAR-MDC), Rotterdam. We gratefully acknowledge the contribution of children and parents, general practitioners, hospitals, midwives and pharmacies in Rotterdam. The study protocol was approved by the Medical Ethical Committee of the Erasmus Medical Centre, Rotterdam. Written informed consent was obtained from all participants. The general design of Generation R Study is made possible by financial support from the Erasmus Medical Center, Rotterdam, the Erasmus University Rotterdam, the Netherlands Organization for Health Research and Development (ZonMw), the Netherlands Organisation for Scientific Research (NWO), the Ministry of Health, Welfare and Sport and the Ministry of Youth and Families. Vincent W. Jaddoe received an additional grant from the Netherlands Organization for Health Research and Development (VIDI 016.136.361) and a European Research Council Consolidator Grant (ERC-2014-CoG-648916). The generation and management of GWAS genotype data for the Generation R Study were done

at the Genetic Laboratory of the Department of Internal Medicine, Erasmus MC, the Netherlands. We would like to thank Karol Estrada, Dr. Tobias A. Knoch, Anis Abuseiris, Luc V. de Zeeuw, and Rob de Graaf, for their help in creating GRIMP, BigGRID, MediGRID, and Services@MediGRID/D-Grid, (funded by the German Bundesministerium fuer Forschung und Technology; grants 01 AK 803 A-H, 01 IG 07015 G) for access to their grid computing resources. We thank Mila Jhamai, Manoushka Ganesh, Pascal Arp, Marijn Verkerk, Lizbeth Herrera and Marjolein Peters for their help in creating, managing and QC of the GWAS database. Also, we thank Karol Estrada for their support in creation and analysis of imputed data.

GOYA

This study was conducted as part of the activities of the Gene-diet Interactions in Obesity project (GENDINOBS, www.gendinob.dk) and the MRC centre for Causal Analyses in Translational Epidemiology (MRC CAiTE). Tarunveer S. Ahluwalia was supported by funding from GENDINOBS project (www.gendinob.dk).

HBCS

The Helsinki Birth Cohort Study (HBCS/HBCS 1934-44) thanks Professor David Barker and Tom Forsen. Major financial support was received from the Academy of Finland (project grants 120315, 129287, 218029 and 267561). The DNA extraction, sample quality control, biobank up-keep and aliquotting were performed at the National Institute for Health and Welfare, Helsinki, Finland.

Danish Childhood Obesity Biobank

The Danish Childhood Obesity Biobank would like to thank all children, youths, and their families for participation in the studies and thus providing opportunities to detect novel insights in regards to childhood obesity. This study was supported by grants from the Program

Committee for Individuals, Disease and Society of the Danish Council for Strategic Research (grant numbers: 11-115917, TARGET (<http://target.ku.dk>) and 11-116714, BIOCHILD (<http://biochild.ku.dk/>)). The Novo Nordisk Foundation Center for Basic Metabolic Research, an independent Research Center at the University of Copenhagen partially funded by an unrestricted donation from the Novo Nordisk Foundation. Haja N Kadarmideen is supported by a EU-FP7 Marie Curie Actions - Career Integration Grant (CIG-293511).

INMA

This study was funded by grants from Instituto de Salud Carlos III (CB06/02/0041, G03/176, FIS PI041436, PI081151, PI041705, PI061756, PI091958, and PS09/00432, FIS-FEDER 03/1615, 04/1509, 04/1112, 04/1931, 05/1079, 05/1052, 06/1213, 07/0314, 09/02647, 11/01007, 11/02591, 11/02038, 13/1944, 13/2032 and CP11/0178), Spanish Ministry of Science and Innovation (SAF2008-00357), European Commission (ENGAGE project and grant agreement HEALTH-F4-2007-201413, HEALTH.2010.2.4.5-1, FP7-ENV-2011 cod 282957), Fundació La Marató de TV3, Generalitat de Catalunya-CIRIT 1999SGR 00241 and Conselleria de Sanitat Generalitat Valenciana. Part of the DNA extractions and genotyping was performed at the Spanish National Genotyping Centre (CEGEN-Barcelona). The authors are grateful to Silvia Fochs, Anna Sànchez, Maribel López, Nuria Pey, Muriel Ferrer, Amparo Quiles, Sandra Pérez, Gemma León, Elena Romero, Maria Andreu, Nati Galiana, Maria Dolores Climent, Amparo Cases and Cristina Capo for their assistance in contacting the families and administering the questionnaires. The authors would particularly like to thank all the participants for their generous collaboration. A full roster of the INMA Project Investigators can be found at http://www.proyectoinma.org/presentacion-inma/listado-investigadores/en_listado-investigadores.html.

Leipzig

The Leipzig Childhood Obesity cohort is supported by grants from Integrated Research and Treatment Centre (IFB) Adiposity Diseases FKZ: 01EO1001, from the German Research Foundation for the Clinical Research Center “Obesity Mechanisms” CRC1052/1 C05 and the European Community’s Seventh Framework Programme (FP7/2007-2013) project Beta-JUDO under grant agreement n° 279153. We are grateful to all the patients and families for contributing to the study. We highly appreciate the support of the Obesity Team and Auxo Team of the Leipzig University Children’s Hospital for management of the patients and to the Pediatric Research Center Lab Team for support with DNA banking.

LISA+GINI

The authors thank all families for participation in the studies; the obstetric units for allowing recruitment and the LISA and GINI study teams for excellent work. Generation of GWA data in the LISApplus and GINIplus studies in Munich were covered by Helmholtz Zentrum Munich, Helmholtz Centre for Environmental Research. Personal and financial support by the Munich Center of Health Sciences (MCHEALTH) as part of the Ludwig-Maximilians University Munich LMU innovative is gratefully acknowledged. In addition, this work was supported by the Kompetenznetz Adipositas (Competence Network Obesity) funded by the Federal Ministry of Education and Research (FKZ: 01GI1121A).

MAAS

We would like to thank the children and their parents for their continued support and enthusiasm. We greatly appreciate the commitment they have given to the project. We would also like to acknowledge the hard work and dedication of the study team (post-doctoral scientists, research fellows, nurses, physiologists, technicians and clerical staff). MAAS was supported by the

Asthma UK Grants No 301 (1995-1998), No 362 (1998-2001), No 01/012 (2001-2004), No 04/014 (2004-2007) and The Moulton Charitable Foundation (2004-current); age 11 years clinical follow-up is funded by the Medical Research Council (MRC) Grant G0601361.

MoBa

This work was supported by grants from the Norwegian Research Council (FUGE 183220/S10, FRIMEDKLI-05 ES236011), Swedish Medical Society (SLS 2008-21198), Jane and Dan Olsson Foundations and Swedish government grants to researchers in the public health service (ALFGBG-2863, ALFGBG-11522), and the European Community's Seventh Framework Programme (FP7/2007-2013), ENGAGE Consortium, grant agreement HEALTH-F4-2007-201413. The Norwegian Mother and Child Cohort Study was also supported by the Norwegian Ministry of Health and the Ministry of Education and Research, NIH/NIEHS (contract no N01-ES-75558), NIH/NINDS (grant no.1 U01 NS 047537-01 and grant no.2 U01 NS 047537-06A1), and the Norwegian Research Council/FUGE (grant no. 151918/S10). We are grateful to all the participating families in Norway who take part in this ongoing cohort study. Researchers interested in using MoBa data must obtain approval from the Scientific Management Committee of MoBa and from the Regional Committee for Medical and Health Research Ethics for access to data and biological material. Researchers are required to follow the terms of an Assistance Agreement containing a number of clauses designed to ensure protection of privacy and compliance with relevant laws.

NFBC 1966 and 1986

NFBC1966 and 1986 has received financial support from e.g. the Academy of Finland, University Hospital Oulu, Biocenter, University of Oulu, Sigrid Juselius Foundation, Finland, the European Commission (EURO-BLCS, Framework 5 award QLG1-CT-2000-01643, ENGAGE

project and grant agreement HEALTH-F4-2007-201413, EurHEALTHAgeing (277849), European Regional Developmental Fund), NHLBI grant 5R01HL087679-02 through the STAMPEED program (1RL1MH083268-01), NIH/NIMH (5R01MH63706:02), USA, Stanley Foundation, USA, the Medical Research Council, UK and the Wellcome Trust, UK. Our most sincere thanks go to all cohort members and their parents. The Section of Investigative Medicine, Imperial College London is funded by grants from the MRC, BBSRC, NIHR, an Integrative Mammalian Biology (IMB) Capacity Building Award, an FP7- HEALTH- 2009- 241592 EuroCHIP grant and is supported by the NIHR Imperial Biomedical Research Centre Funding Scheme. AIFB is funded by the MRC, EPSRC and Diabetes UK.

NTR

We thank all participating twin families. NTR is supported by ARRA RC2 2MH08995; the European Research Council (Genetics of Mental Illness, ERC-230374); Spinozapremie (NWO/SPI 56-464-14192) and Twin-family database for behavior genetics and genomics studies (NWO 480-04-004).

PANIC

We thank the children and their families who participated in The PANIC Study. The study protocol was approved by the Research Ethics Committee of the Hospital District of Northern Savo. All children and their parents gave their informed written consent. The PANIC Study has been financially supported by grants from the Ministry of Social Affairs and Health of Finland, the Ministry of Education and Culture of Finland, the University of Eastern Finland, the Finnish Innovation Fund Sitra, the Social Insurance Institution of Finland, the Finnish Cultural Foundation, the Juho Vainio Foundation, the Foundation for Paediatric Research, the Paulo Foundation, the Paavo Nurmi Foundation, the Diabetes Research Foundation, Kuopio University

Hospital (EVO-funding number 5031343) and the Research Committee of the Kuopio University Hospital Catchment Area for the State Research Funding.

PIAMA

The PIAMA birth cohort study is a collaboration of the Institute for Risk Assessment Sciences, University Utrecht (B. Brunekreef), Julius Center for Health Sciences and Primary Care, University Medical Center Utrecht (H.A. Smit), Centre for Prevention and Health Services Research, National Institute for Public Health and the Environment, Bilthoven (A.H. Wijga), Department of Pediatrics, Division of Respiratory Medicine, Erasmus MC -Sophia, Rotterdam (J.C. de Jongste), Pulmonology (D.S. Postma) and Pediatric Pulmonology and Pediatric Allergology (G.H. Koppelman) of the University Medical Center Groningen and the Department of Immunopathology, Sanquin Research, Amsterdam (R.C. Aalberse), the Netherlands. The study team gratefully acknowledges the participants in the PIAMA birth cohort study, and all coworkers who helped conducting the medical examinations, field work and data management. The PIAMA study was funded by grants from the Dutch Asthma Foundation (grant 3.4.01.26, 3.2.06.022, 3.4.09.081 and 3.2.10.085CO), the ZON-MW Netherlands Organization for Health Research and Development (grant 912-03-031), the Stichting Astmabestrijding and the Ministry of the Environment. Genome-wide genotyping was funded by the European Commission as part of GABRIEL (A multidisciplinary study to identify the genetic and environmental causes of asthma in the European Community) contract number 018996 under the Integrated Program LSH-2004-1.2.5-1 Post genomic approaches to understand the molecular basis of asthma aiming at a preventive or therapeutic control and a Grant from BBMRI-NL (CP 29). Gerard H. Koppelman received grants from the Dutch Lung Foundation and BBMRI-NL.

RAINE

The authors are grateful to the Raine Study participants and their families, and to the Raine Study research staff for cohort coordination and data collection. The authors gratefully acknowledge the NH&MRC for their long term contribution to funding the study over the last 20 years and also the following Institutions for providing funding for Core Management of the Raine Study: The University of Western Australia (UWA), Raine Medical Research Foundation, UWA Faculty of Medicine, Dentistry and Health Sciences, Telethon Kids Institute and Women and Infants Research Foundation and Curtin University. The authors gratefully acknowledge the assistance of the Western Australian DNA Bank (National Health and Medical Research Council of Australia National Enabling Facility). This study was supported by the National Health and Medical Research Council of Australia (grant numbers 572613, 403981 and 003209) and the Canadian Institutes of Health Research (grant number MOP-82893).

STRIP

The STRIP study was financially supported by Academy of Finland (grants 206374, 251360 and 26080328); Juho Vainio Foundation; Finnish Cardiac Research Foundation; Finnish Cultural Foundation; Finnish Ministry of Education and Culture; Sigrid Juselius Foundation; Yrjö Jahnsson Foundation; C.G. Sundell Foundation; Special Governmental Grants for Health Sciences Research, Turku University Hospital; Foundation for Pediatric Research; and Turku University Foundation.

TEDS

We gratefully acknowledge the on-going contribution of the families in the Twins Early Development Study (TEDS). TEDS is supported by a program grant from the UK Medical Research Council (G0901245), with genotyping provided by the Wellcome Trust Case Control Consortium 2 project (085475/B/08/Z and 085475/Z/08/Z). R. Plomin is supported by

a research professorship from the UK Medical Research Council (G19/2) and a European Research Council Advanced Investigator Award (295366).

TEENAGE

We would like to thank all study participants and their families as well as all volunteers for their contribution in this study. We thank the following staff from the Sample Management and Genotyping Facilities at the Wellcome Trust Sanger Institute for sample preparation, quality control and genotyping: Dave Jones, Doug Simpkin, Emma Gray, Hannah Blackburn, Sarah Edkins. The TEENAGE study has been co-financed by the European Union (European Social Fund—ESF) and Greek national funds through the Operational Program “Education and Lifelong Learning” of the National Strategic Reference Framework (NSRF)—Research Funding Program: Heracleitus II. Investing in knowledge society through the European Social Fund. This work was funded by the Wellcome Trust (098051).

YFS

Young Finns Study was financially supported by the Academy of Finland (134309 (Eye), 126925, 121584, 124282, 129378 (Salve), 117787 (Gendi), and 41071 (Skidi)); the Social Insurance Institution of Finland, Kuopio, Tampere; Turku University Hospital Medical Funds (grant 9M048 and 9N035 to Terho Lehtimäki); Juho Vainio Foundation; Paavo Nurmi Foundation; Finnish Foundation of Cardiovascular Research and Finnish Cultural Foundation; Tampere Tuberculosis Foundation; and Emil Aaltonen Foundation (to Terho Lehtimäki).

Further acknowledgements

Baoshan Ma is supported by the National Natural Science Foundation of China (NO.61471078), and the China Postdoctoral Science Foundation (No.2014M551084). Tune H. Pers is supported by the Alfred Benzon Foundation.

Conflict of Interest Statement

The authors declare no competing financial interests.

References

1. Baker, J.L., Olsen, L.W. and Sorensen, T.I. (2007) Childhood body-mass index and the risk of coronary heart disease in adulthood. *N. Engl. J. Med.*, **357**, 2329-2337.
2. Franks, P.W., Hanson, R.L., Knowler, W.C., Sievers, M.L., Bennett, P.H. and Looker, H.C. (2010) Childhood obesity, other cardiovascular risk factors, and premature death. *N. Engl. J. Med.*, **362**, 485-493.
3. Ng, M. and Fleming, T. and Robinson, M. and Thomson, B. and Graetz, N. and Margono, C. and Mullany, E.C. and Biryukov, S. and Abbafati, C. and Abera, S.F. *et al.* (2014) Global, regional, and national prevalence of overweight and obesity in children and adults during 1980-2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet*, **384**, 766-781.
4. Reilly, J.J., Methven, E., McDowell, Z.C., Hacking, B., Alexander, D., Stewart, L. and Kelnar, C.J. (2003) Health consequences of obesity. *Arch. Dis. Child.*, **88**, 748-752.
5. Silventoinen, K., Pietilainen, K.H., Tynelius, P., Sorensen, T.I., Kaprio, J. and Rasmussen, F. (2007) Genetic and environmental factors in relative weight from birth to age 18: the Swedish young male twins study. *Int. J. Obes. (Lond)*, **31**, 615-621.
6. Elks, C.E., den Hoed, M., Zhao, J.H., Sharp, S.J., Wareham, N.J., Loos, R.J. and Ong, K.K. (2012) Variability in the heritability of body mass index: a systematic review and meta-regression. *Front. Endocrinol. (Lausanne)*, **3**, 29.

7. Maes, H.H., Neale, M.C. and Eaves, L.J. (1997) Genetic and environmental factors in relative body weight and human adiposity. *Behav. Genet.*, **27**, 325-351.
8. Silventoinen, K., Rokholm, B., Kaprio, J. and Sorensen, T.I. (2010) The genetic and environmental influences on childhood obesity: a systematic review of twin and adoption studies. *Int. J. Obes. (Lond)*, **34**, 29-40.
9. Berndt, S.I. and Gustafsson, S. and Magi, R. and Ganna, A. and Wheeler, E. and Feitosa, M.F. and Justice, A.E. and Monda, K.L. and Croteau-Chonka, D.C. and Day, F.R. *et al.* (2013) Genome-wide meta-analysis identifies 11 new loci for anthropometric traits and provides insights into genetic architecture. *Nat. Genet.*, **45**, 501-512.
10. Heid, I.M. and Jackson, A.U. and Randall, J.C. and Winkler, T.W. and Qi, L. and Steinthorsdottir, V. and Thorleifsson, G. and Zillikens, M.C. and Speliotes, E.K. and Magi, R. *et al.* (2010) Meta-analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution. *Nat. Genet.*, **42**, 949-960.
11. Locke, A.E. and Kahali, B. and Berndt, S.I. and Justice, A.E. and Pers, T.H. and Day, F.R. and Powell, C. and Vedantam, S. and Buchkovich, M.L. and Yang, J. *et al.* (2015) Genetic studies of body mass index yield new insights for obesity biology. *Nature*, **518**, 197-206.
12. Shungin, D. and Winkler, T.W. and Croteau-Chonka, D.C. and Ferreira, T. and Locke, A.E. and Magi, R. and Strawbridge, R.J. and Pers, T.H. and Fischer, K. and Justice, A.E. *et al.* (2015) New genetic loci link adipose and insulin biology to body fat distribution. *Nature*, **518**, 187-196.

13. Speliotes, E.K. and Willer, C.J. and Berndt, S.I. and Monda, K.L. and Thorleifsson, G. and Jackson, A.U. and Lango Allen, H. and Lindgren, C.M. and Luan, J. and Magi, R. *et al.* (2010) Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nat. Genet.*, **42**, 937-948.
14. Haworth, C.M., Carnell, S., Meaburn, E.L., Davis, O.S., Plomin, R. and Wardle, J. (2008) Increasing heritability of BMI and stronger associations with the FTO gene over childhood. *Obesity (Silver Spring)*, **16**, 2663-2668.
15. Sovio, U., Mook-Kanamori, D.O., Warrington, N.M., Lawrence, R., Briollais, L., Palmer, C.N., Cecil, J., Sandling, J.K., Syvanen, A.C., Kaakinen, M. *et al.* (2011) Association between common variation at the FTO locus and changes in body mass index from infancy to late childhood: the complex nature of genetic association through growth and development. *PLoS Genet.*, **7**, e1001307.
16. Bradfield, J.P., Taal, H.R., Timpson, N.J., Scherag, A., Lecoeur, C., Warrington, N.M., Hypponen, E., Holst, C., Valcarcel, B., Thiering, E. *et al.* (2012) A genome-wide association meta-analysis identifies new childhood obesity loci. *Nat. Genet.*, **44**, 526-531.
17. Comuzzie, A.G., Cole, S.A., Laston, S.L., Voruganti, V.S., Haack, K., Gibbs, R.A. and Butte, N.F. (2012) Novel genetic loci identified for the pathophysiology of childhood obesity in the Hispanic population. *PLoS One*, **7**, e51954.
18. Scherag, A., Dina, C., Hinney, A., Vatin, V., Scherag, S., Vogel, C.I., Muller, T.D., Grallert, H., Wichmann, H.E., Balkau, B. *et al.* (2010) Two new Loci for body-weight regulation identified in a joint analysis of genome-wide association studies for early-onset extreme obesity in French and German study groups. *PLoS Genet.*, **6**, e1000916.

19. Namjou, B., Keddache, M., Marsolo, K., Wagner, M., Lingren, T., Cobb, B., Perry, C., Kennebeck, S., Holm, I.A., Li, R. *et al.* (2013) EMR-linked GWAS study: investigation of variation landscape of loci for body mass index in children. *Front. Genet.*, **4**, 268.
20. Jess, T., Zimmermann, E., Kring, S.I., Berentzen, T., Holst, C., Toubro, S., Astrup, A., Hansen, T., Pedersen, O. and Sorensen, T.I. (2008) Impact on weight dynamics and general growth of the common FTO rs9939609: a longitudinal Danish cohort study. *Int. J. Obes. (Lond)*, **32**, 1388-1394.
21. Westra, H.J., Peters, M.J., Esko, T., Yaghootkar, H., Schurmann, C., Kettunen, J., Christiansen, M.W., Fairfax, B.P., Schramm, K., Powell, J.E. *et al.* (2013) Systematic identification of trans eQTLs as putative drivers of known disease associations. *Nat. Genet.*, **45**, 1238-1243.
22. Grundberg, E., Small, K.S., Hedman, A.K., Nica, A.C., Buil, A., Keildson, S., Bell, J.T., Yang, T.P., Meduri, E., Barrett, A. *et al.* (2012) Mapping cis- and trans-regulatory effects across multiple tissues in twins. *Nat. Genet.*, **44**, 1084-1089.
23. Yang, T.P., Beazley, C., Montgomery, S.B., Dimas, A.S., Gutierrez-Arcelus, M., Stranger, B.E., Deloukas, P. and Dermitzakis, E.T. (2010) Genevar: a database and Java application for the analysis and visualization of SNP-gene associations in eQTL studies. *Bioinformatics*, **26**, 2474-2476.
24. Liang, L., Morar, N., Dixon, A.L., Lathrop, G.M., Abecasis, G.R., Moffatt, M.F. and Cookson, W.O. (2013) A cross-platform analysis of 14,177 expression quantitative trait loci derived from lymphoblastoid cell lines. *Genome Res.*, **23**, 716-726.
25. Geller, F., Feenstra, B., Carstensen, L., Pers, T.H., van Rooij, I.A., Korberg, I.B., Choudhry, S., Karjalainen, J.M., Schnack, T.H., Hollegaard, M.V. *et al.* (2014) Genome-

- wide association analyses identify variants in developmental genes associated with hypospadias. *Nat. Genet.*, **46**, 957-963.
26. Warrington, N.M., Howe, L.D., Paternoster, L., Kaakinen, M., Herrala, S., Huikari, V., Wu, Y.Y., Kemp, J.P., Timpson, N.J., St Pourcain, B. *et al.* (2015) A genome-wide association study of body mass index across early life and childhood. *Int. J. Epidemiol.*, **44**, 700-712.
27. Nambu, H., Fukushima, M., Hikichi, H., Inoue, T., Nagano, N., Tahara, Y., Nambu, T., Ito, J., Ogawa, Y., Ozaki, S. *et al.* (2011) Characterization of metabolic phenotypes of mice lacking GPR61, an orphan G-protein coupled receptor. *Life Sci.*, **89**, 765-772.
28. Creppe, C., Malinouskaya, L., Volvert, M.L., Gillard, M., Close, P., Malaise, O., Laguesse, S., Cornez, I., Rahmouni, S., Ormenese, S. *et al.* (2009) Elongator controls the migration and differentiation of cortical neurons through acetylation of alpha-tubulin. *Cell*, **136**, 551-564.
29. Okada, Y., Yamagata, K., Hong, K., Wakayama, T. and Zhang, Y. (2010) A role for the elongator complex in zygotic paternal genome demethylation. *Nature*, **463**, 554-558.
30. Simpson, C.L., Lemmens, R., Miskiewicz, K., Broom, W.J., Hansen, V.K., van Vught, P.W., Landers, J.E., Sapp, P., Van Den Bosch, L., Knight, J. *et al.* (2009) Variants of the elongator protein 3 (ELP3) gene are associated with motor neuron degeneration. *Hum. Mol. Genet.*, **18**, 472-481.
31. Chen, X., Li, C., Izumi, T., Ernst, S.A., Andrews, P.C. and Williams, J.A. (2004) Rab27b localizes to zymogen granules and regulates pancreatic acinar exocytosis. *Biochem. Biophys. Res. Commun.*, **323**, 1157-1162.

32. Suda, J., Zhu, L., Okamoto, C.T. and Karvar, S. (2011) Rab27b localizes to the tubulovesicle membranes of gastric parietal cells and regulates acid secretion. *Gastroenterology*, **140**, 868-878.
33. Zhao, S., Torii, S., Yokota-Hashimoto, H., Takeuchi, T. and Izumi, T. (2002) Involvement of Rab27b in the regulated secretion of pituitary hormones. *Endocrinology*, **143**, 1817-1824.
34. Hu, C., Lv, H., Pan, G., Cao, H., Deng, Z., Hu, C., Wen, J. and Zhou, J. (2011) The expression of ADAM23 and its correlation with promoter methylation in non-small-cell lung carcinoma. *Int. J. Exp. Pathol.*, **92**, 333-339.
35. Sun, Y.P., Deng, K.J., Wang, F., Zhang, J., Huang, X., Qiao, S. and Zhao, S. (2004) Two novel isoforms of Adam23 expressed in the developmental process of mouse and human brains. *Gene*, **325**, 171-178.
36. Kim, H.A., Park, W.J., Jeong, H.S., Lee, H.E., Lee, S.H., Kwon, N.S., Baek, K.J., Kim, D.S. and Yun, H.Y. (2012) Leucine-rich glioma inactivated 3 regulates adipogenesis through ADAM23. *Biochim. Biophys. Acta*, **1821**, 914-922.
37. Stergiakouli, E., Gaillard, R., Tavaré, J.M., Balthasar, N., Loos, R.J., Taal, H.R., Evans, D.M., Rivadeneira, F., St Pourcain, B., Uitterlinden, A.G. *et al.* (2014) Genome-wide association study of height-adjusted BMI in childhood identifies functional variant in ADCY3. *Obesity (Silver Spring)*, **22**, 2252-2259.
38. Nag, A., Venturini, C., Small, K.S., International Glaucoma Genetics, C., Young, T.L., Viswanathan, A.C., Mackey, D.A., Hysi, P.G. and Hammond, C. (2014) A genome-wide association study of intra-ocular pressure suggests a novel association in the gene FAM125B in the TwinsUK cohort. *Hum. Mol. Genet.*, **23**, 3343-3348.

39. Bulik-Sullivan, B.K., Loh, P.R., Finucane, H.K., Ripke, S., Yang, J., Schizophrenia Working Group of the Psychiatric Genomics, C., Patterson, N., Daly, M.J., Price, A.L. and Neale, B.M. (2015) LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat. Genet.*, **47**, 291-295.
40. Willer, C.J., Li, Y. and Abecasis, G.R. (2010) METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics*, **26**, 2190-2191.
41. Park, J.H., Wacholder, S., Gail, M.H., Peters, U., Jacobs, K.B., Chanock, S.J. and Chatterjee, N. (2010) Estimation of effect size distribution from genome-wide association studies and implications for future discoveries. *Nat. Genet.*, **42**, 570-575.
42. Pers, T.H., Karjalainen, J.M., Chan, Y., Westra, H.J., Wood, A.R., Yang, J., Lui, J.C., Vedantam, S., Gustafsson, S., Esko, T. *et al.* (2015) Biological interpretation of genome-wide association studies using predicted gene functions. *Nat. Commun.*, **6**, 5890.
43. Fehrmann, R.S., Karjalainen, J.M., Krajewska, M., Westra, H.J., Maloney, D., Simeonov, A., Pers, T.H., Hirschhorn, J.N., Jansen, R.C., Schultes, E.A. *et al.* (2015) Gene expression analysis identifies global gene dosage sensitivity in cancer. *Nat. Genet.*, **47**, 115-125.

Legends to figures

Figure 1 Manhattan plot of results of the discovery meta-analysis of 20 studies

Chromosomes are shown on the x-axis, the $-\log_{10}$ of the P -value on the y-axis. The grey dotted line represents the genome-wide significance cutoff of 5×10^{-8} . Genes shown in black are the known loci that were significantly associated with childhood BMI in the joint discovery and replication analysis. Genes shown in grey were significant in the discovery, but not in the joint discovery and replication analysis. * indicates novel loci that were significantly associated with childhood BMI in the joint discovery and replication analysis. See also Table 1.

Figure 2 Regional plots of the three novel loci for childhood BMI

On the x-axis the position of SNPs on the chromosome is shown. On the left y-axis is the $-\log_{10}$ of the P -values from the discovery analysis, on the right y-axis is the estimated recombination rate (from HapMap), shown by the light blue line in the figure. The named SNP is the most significant SNP in the locus from the discovery meta-analysis. . The linkage disequilibrium of all SNPs with the most significant SNP is shown by the symbols, with dark grey diamonds indicating an R^2 of ≥ 0.8 , inversed dark grey triangles indicating an R^2 of 0.6-0.8, dark grey triangles indicating an R^2 of 0.4-0.6, dark grey circles indicating an R^2 of 0.2-0.4, and light grey circles indicating an R^2 of 0-0.2. Genes (from HapMap release 22) are plotted below the x-axis.

Figure 3 Association of the weighted risk score with BMI

Along the x axis, categories of the weighted risk score are presented, the mean standard deviation score (SDS)-BMI per group is shown on the right y axis, with the line representing the

regression of the mean SDS-BMI values on the categories of the weighted risk score. The left y axis represents the number of children in each risk score category, shown in the histogram. The *P*-value is derived from the analysis of the continuous risk score. Analysis was performed in the PIAMA Study (N =1955).

Tables

Table 1 Results of the discovery, replication and joint analyses for 43 loci with P -values $< 5 \times 10^{-6}$ in the discovery phase

SNP	CHR	Position	Nearest gene	EA/Non-EA	EAF ^a	Beta ^a	SE ^a	P -value discovery	P -value replication	P -value joint
rs13130484 ^b	4	44870448	<i>GNPDA2</i>	T/C	0.44	0.067	0.007	8.94×10^{-11}	4.29×10^{-18}	1.58×10^{-23}
rs11676272 ^b	2	24995042	<i>ADCY3</i>	G/A	0.46	0.068	0.007	8.55×10^{-23}	0.020	7.12×10^{-23}
rs4854349 ^b	2	637861	<i>TMEM18</i>	C/T	0.83	0.090	0.009	6.00×10^{-21}	0.005	5.41×10^{-22}
rs543874 ^b	1	176156103	<i>SEC16B</i>	G/A	0.20	0.077	0.009	2.38×10^{-17}	8.77×10^{-4}	2.20×10^{-19}
rs7132908 ^b	12	48549415	<i>FAIM2</i>	A/G	0.39	0.066	0.008	4.99×10^{-19}	0.043	1.57×10^{-18}
rs1421085 ^b	16	52358455	<i>FTO</i>	C/T	0.41	0.059	0.007	3.20×10^{-19}	0.654	4.53×10^{-16}
rs12429545 ^c	13	53000207	<i>OLFM4</i>	A/G	0.13	0.076	0.010	3.66×10^{-11}	1.01×10^{-4}	2.08×10^{-14}
rs987237 ^b	6	50911009	<i>TFAP2B</i>	G/A	0.19	0.062	0.009	3.81×10^{-13}	0.224	1.80×10^{-12}
rs12041852 ^b	1	74776088	<i>TNNI3K</i>	G/A	0.46	0.046	0.007	1.77×10^{-10}	0.142	2.28×10^{-10}
rs6567160 ^b	18	55980115	<i>MC4R</i>	C/T	0.23	0.050	0.008	4.06×10^{-12}	0.996	1.21×10^{-9}
rs13253111	8	28117893	<i>ELP3</i>	A/G	0.57	0.042	0.007	4.13×10^{-9}	0.114	4.89×10^{-9}
rs8092503	18	50630485	<i>RAB27B</i>	G/A	0.27	0.045	0.008	8.55×10^{-8}	0.034	8.17×10^{-9}
rs3829849 ^b	9	128430621	<i>LMX1B</i>	T/C	0.36	0.041	0.007	1.46×10^{-6}	0.001	8.81×10^{-9}
rs13387838	2	206989692	<i>ADAM23</i>	A/G	0.04	0.139	0.025	2.40×10^{-8}	0.306	2.84×10^{-8}
rs7550711 ^d	1	109884409	<i>GPR61</i>	T/C	0.04	0.105	0.019	1.50×10^{-8}	0.401	4.52×10^{-8}
rs17309930 ^b	11	27705069	<i>BDNF</i>	A/C	0.21	0.045	0.009	2.47×10^{-8}	0.540	1.41×10^{-7}
rs2590942 ^b	1	72657869	<i>NEGR1</i>	T/G	0.82	0.047	0.009	3.88×10^{-9}	0.966	1.91×10^{-7}
rs13107325 ^b	4	103407732	<i>SLC39A8</i>	T/C	0.07	0.081	0.016	1.19×10^{-8}	0.970	3.79×10^{-7}
rs10151686 ^b	14	29536217	<i>PRKDI</i>	A/G	0.04	0.096	0.019	1.50×10^{-6}	0.109	6.99×10^{-7}
rs25832	5	66211438	<i>LOC375449</i>	A/G	0.71	0.039	0.008	2.41×10^{-6}	0.177	1.62×10^{-6}

rs7869969	9	95257268	<i>FAM120A</i>	G/A	0.33	0.036	0.008	4.43 x 10 ⁻⁶	0.425	1.68 x 10 ⁻⁶
rs11079830 ^c	17	44037629	<i>HOXB6</i>	A/G	0.58	0.034	0.007	1.43 x 10 ⁻⁶	0.254	1.98 x 10 ⁻⁶
rs4569924	5	153520218	<i>GALNT10</i>	T/C	0.43	0.032	0.007	4.06 x 10 ⁻⁶	0.823	3.48 x 10 ⁻⁶
rs8046312 ^b	16	19886835	<i>GPRI39</i>	A/C	0.81	0.042	0.009	4.06 x 10 ⁻⁶	0.185	3.97 x 10 ⁻⁶
rs1838856	2	113822060	<i>PAX8</i>	A/C	0.46	0.034	0.008	1.85 x 10 ⁻⁶	0.588	1.47 x 10 ⁻⁵
rs633143	1	179716108	<i>CACNA1E</i>	T/C	0.14	0.044	0.011	3.40 x 10 ⁻⁶	0.648	2.44 x 10 ⁻⁵
rs4923207	11	24713901	<i>LUZP2</i>	T/G	0.79	0.039	0.010	1.60 x 10 ⁻⁶	0.834	3.52 x 10 ⁻⁵
rs6971577	7	140350204	<i>MRPS33</i>	C/G	0.78	0.036	0.009	1.17 x 10 ⁻⁶	0.690	6.80 x 10 ⁻⁵
rs10866069	3	64366964	<i>ADAMTS9</i>	T/C	0.17	0.041	0.011	3.05 x 10 ⁻⁶	0.687	8.43 x 10 ⁻⁵
rs12457682	18	7216505	<i>LAMA1</i>	C/A	0.23	0.035	0.009	3.81 x 10 ⁻⁶	0.942	9.01 x 10 ⁻⁵
rs11165675 ^b	1	96812556	<i>PTBP2</i>	A/G	0.27	0.031	0.008	2.93 x 10 ⁻⁶	0.520	1.01 x 10 ⁻⁴
rs12096993	1	217931859	<i>SLC30A10</i>	T/C	0.27	0.031	0.008	1.38 x 10 ⁻⁶	0.583	1.02 x 10 ⁻⁴
rs760931	1	1637388	<i>CDC2L1</i>	C/G	0.93	0.103	0.027	3.15 x 10 ⁻⁶	0.160	1.27 x 10 ⁻⁴
rs2968990	4	131098524	<i>C4orf33</i>	C/T	0.37	0.028	0.007	1.67 x 10 ⁻⁶	0.487	1.42 x 10 ⁻⁴
rs1247117	10	120418792	<i>C10orf46</i>	G/A	0.11	0.040	0.011	3.47 x 10 ⁻⁶	0.339	2.62 x 10 ⁻⁴
rs6580706	12	47959818	<i>TUBA1C</i>	C/G	0.34	0.031	0.009	1.83 x 10 ⁻⁶	0.047	3.68 x 10 ⁻⁴
rs8092620	18	41433991	<i>SLC14A2</i>	G/T	0.48	0.024	0.007	3.31 x 10 ⁻⁶	0.199	7.32 x 10 ⁻⁴
rs188584	3	62675007	<i>CADPS</i>	C/A	0.77	0.028	0.008	3.23 x 10 ⁻⁶	0.139	0.001
rs4870949	8	126704776	<i>TRIB1</i>	T/C	0.07	0.164	0.054	3.13 x 10 ⁻⁶	0.061	0.002
rs1573972	4	171559399	<i>AADAT</i>	C/T	0.19	0.030	0.013	3.78 x 10 ⁻⁶	0.232	0.020
rs214821	20	2258291	<i>TGM3</i>	T/C	0.02	0.479	0.208	3.38 x 10 ⁻⁶	0.575	0.021
rs8084077	18	49532928	<i>DCC</i>	T/C	0.73	0.014	0.008	3.47 x 10 ⁻⁶	2.72 x 10 ⁻⁵	0.062
rs3845265	18	63690108	<i>DSEL</i>	G/A	0.71	0.013	0.008	8.86 x 10 ⁻⁷	1.44 x 10 ⁻⁶	0.087

^a From joint analysis

^b Locus previously reported in (11)

^c Locus previously reported in (11, 16)

^d Locus previously reported in (9, 11)

CHR: Chromosome; EA: Effect Allele; EAF: Effect Allele Frequency; SE: Standard Error

Abbreviations

1000G – 1000 Genomes

BMI – Body mass index

CHR - Chromosome

DEPICT – Data-Driven Expression Prioritized Integration for Complex Traits

EA – Effect allele

EAF – Effect allele frequency

eQTL – Expression quantitative trait locus

FDR – False discovery rate

GWAS – Genome-wide association study

LD – Linkage disequilibrium

MAF – Minor allele frequency

HWE – Hardy-Weinberg Equilibrium

SDS – Standard deviation score

SE – Standard error

SNP – Single Nucleotide Polymorphism





