



King's Research Portal

DOI: 10.1001/jamapsychiatry.2016.0442

Document Version Publisher's PDF, also known as Version of record

Link to publication record in King's Research Portal

Citation for published version (APA):

Merritt, K., Egerton, A., Kempton, M., Taylor, M. J., & McGuire, P. (2016). Nature of Glutamate Alterations in Schizophrenia: A Meta-analysis of Proton Magnetic Resonance Spectroscopy Studies. *JAMA Psychiatry*, 73(7), 665-674. Article 7. https://doi.org/10.1001/jamapsychiatry.2016.0442

Citing this paper

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

•Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research. •You may not further distribute the material or use it for any profit-making activity or commercial gain •You may freely distribute the URL identifying the publication in the Research Portal

Take down policy

If you believe that this document breaches copyright please contact librarypure@kcl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

Original Investigation | META-ANALYSIS

Nature of Glutamate Alterations in Schizophrenia A Meta-analysis of Proton Magnetic Resonance Spectroscopy Studies

Kate Merritt, MSc; Alice Egerton, PhD; Matthew J. Kempton, PhD; Matthew J. Taylor, DPhil; Philip K. McGuire, FMedSci

IMPORTANCE Alterations in glutamatergic neurotransmission may be fundamental to the pathophysiology of schizophrenia, and the glutamatergic system is a target for novel therapeutic interventions in the disorder.

OBJECTIVE To investigate the nature of brain glutamate alterations in schizophrenia by conducting a meta-analysis of glutamate proton magnetic resonance (MRS) spectroscopy studies.

DATA SOURCES The MEDLINE database was searched for studies published from January 1, 1980, to April 1, 2015. Search terms included *magnetic resonance spectroscopy*, *schizophrenia*, *psychosis*, *clinical* or *genetic high risk*, and *schizoaffective*. Inclusion criteria were single voxel 1H-MRS studies reporting glutamate, glutamine or Glx values for a patient or risk group in comparison to a healthy volunteer group.

STUDY SELECTION Fifty-nine studies were identified, which included 1686 patients and 1451 healthy individuals serving as controls.

DATA EXTRACTION AND SYNTHESIS A random-effects, inverse-weighted variance model was used to calculate the pooled effect size. Mean values were extracted and verified independently. Effect sizes were determined for glutamate, glutamine, and Glx in brain regions that had been examined in at least 3 different studies. A secondary analysis grouped studies into those examining patients at different stages of illness (high risk, first-episode psychosis, or chronic schizophrenia). Effects of age, antipsychotic dose, and symptom severity were determined using meta-regression.

RESULTS In schizophrenia, there were significant elevations in glutamate in the basal ganglia (Hedges g = 0.63; 95% CI, 0.15-1.11), glutamine in the thalamus (g = 0.56; 95% CI, 0.02-1.09), and Glx in the basal ganglia (g = 0.39; 95% CI, 0.09-0.70) and medial temporal lobe (g = 0.32; 95% CI, 0.12-0.52). No region showed a reduction in glutamate metabolites in schizophrenia. Secondary analyses revealed that elevated medial frontal Glx levels were evident in individuals at high risk for schizophrenia (g = 0.26; 95% CI, 0.05-0.46) but not in those with first-episode psychosis or chronic schizophrenia, whereas elevated Glx in the medial temporal lobe was seen with chronic schizophrenia (g = 0.40; 95% CI, 0.08-0.71) but not in the high-risk or first-episode groups. Meta-regression found no association with age, symptom severity, or antipsychotic dose.

CONCLUSIONS AND RELEVANCE Schizophrenia is associated with elevations in glutamatergic metabolites across several brain regions. This finding supports the hypothesis that schizophrenia is associated with excess glutamatergic neurotransmission in several limbic areas and further indicates that compounds that reduce glutamatergic transmission may have therapeutic potential.

JAMA Psychiatry. doi:10.1001/jamapsychiatry.2016.0442 Published online June 15, 2016. Editorial
Supplemental content at jamapsychiatry.com

Author Affiliations: Psychosis Studies Department, Institute of Psychiatry, Psychology, and Neuroscience, King's College London, London, England.

Corresponding Author: Kate Merritt, MSc, Psychosis Studies Department, Institute of Psychiatry, Psychology, and Neuroscience, King's College London, London SE5 8AF, England (kate.merritt@kcl.ac.uk).

everal lines of evidence have implicated alterations of the glutamatergic system in the cause of schizophrenia. The N-methyl-D-aspartate receptor (NMDAR) hypofunction model¹ proposes that schizophrenia is related to dysfunction of NMDARs on parvalbumin-containing y-aminobutyric acid-ergic interneurons, leading to excess glutamate release.² Administration of NMDAR antagonists, such as ketamine, induce a psychotic state in healthy volunteers and exacerbate psychotic symptoms in patients with schizophrenia.³ More recently, an autoimmune disorder associated with autoantibodies to the NMDARs has been associated with psychotic symptoms, and NMDAR autoantibodies may be evident in a small proportion of patients with schizophrenia.⁴ Several genes associated with schizophrenia code for proteins involved in glutamatergic neurotransmission.⁵ The main technique for assessing central glutamate function in man in vivo is proton magnetic resonance spectroscopy (MRS).

Depending on the proton MRS approach and field strength, both glutamate and the glutamate metabolite, glutamine, may be reported separately or in combination (Glx).⁶ Glutamatergic metabolites are usually measured in a predetermined voxel of interest. Concentration estimates reflect both intracellular and extracellular glutamate and glutamate involved in metabolism as well as in neurotransmission.⁷ Over the past 2 decades, several studies have used proton MRS to investigate regional glutamate concentrations in patients with schizophrenia compared with those in healthy volunteers. However, findings have not been consistent across studies, including reports of elevations, 8-26 no differences, 27-51 and reductions 52-63 in the patient group across a variety of brain regions. These differences may relate to regional effects, proton MRS methodologic differences, stage^{24,57-59} or severity⁶⁴ of illness, or treatment effects.^{17,20} The first meta-analysis⁶⁵ of these studies reported decreases in glutamate and increases in glutamine in the medial frontal cortex of patients compared with controls. The total number of publications has more than doubled since the first meta-analysis, which included studies up until 2011. These more recent reports include studies on regions of interest that had previously been examined too infrequently to be included in a meta-analysis. Moreover, the field now includes substantial numbers of studies in individuals at high risk (HR) for schizophrenia and those with first-episode psychosis (FEP), in addition to studies in patients with chronic schizophrenia, permitting separate meta-analyses of these different groups.

The primary aim of this study was to conduct an updated case-control meta-analysis of all published reports of regional glutamatergic measures in those at HR for schizophrenia, with FEP, and with schizophrenia. The second aim was to conduct case-control meta-analyses in clinical subgroups separately (HR, FEP, and chronic schizophrenia [referred to as *schizophrenia* hereinafter]). The third aim was to assess the influences of age, symptom severity, and antipsychotic treatment.

The first hypothesis of the study was that, on the basis of preclinical schizophrenia models showing increases in glutamatergic transmission,⁶⁶ glutamatergic metabolites would be **Question** What is the nature of glutamate alterations in schizophrenia as revealed by studies using proton magnetic resonance spectroscopy?

Findings This meta-analysis evaluated 59 studies reporting on regional glutamate, glutamine, or their combined Glx signals. There were significant elevations in glutamate in the basal ganglia, glutamine in the thalamus, and Glx in the basal ganglia and medial temporal lobe but no associations with age, symptom severity, or antipsychotic medication dose.

Meaning Schizophrenia is associated with elevations in glutamate-related metabolites across several brain regions consistent with the hypothesis that there is excess glutamatergic neurotransmission in this condition.

increased in cases compared with controls. The second hypothesis was that there would be higher glutamatergic metabolite concentrations in FEP and HR individuals compared with those who had schizophrenia, in line with previous studies comparing patient groups.^{23,24,57-59} The third hypothesis was that glutamate and glutamine levels would become lower with antipsychotic treatment^{17,20,21} as well as with age in cases relative to controls,⁶⁵ but that symptom severity would be associated with higher glutamatergic metabolite concentrations.⁶⁴

Methods

Study Selection

The MEDLINE database was searched to identify journal articles published between January 1, 1980, and April 1, 2015, using the following search terms: *MRS* or *magnetic resonance spectroscopy* and (1) *schizophrenia* or (2) *psychosis* or (3) *UHR* or (4) *ARMS* or (5) *ultra high risk* or (6) *clinical high risk* or (7) *genetic high risk* or (8) *prodrom** or (9) *schizoaffective*. All singlevoxel proton MRS studies reporting glutamate, glutamine, or Glx values for a patient or risk group in comparison with a healthy volunteer group were included in the analysis. In the case of longitudinal studies,^{17,43,52} only the values given for the first time point were included. If the same sample or partially overlapping samples were included in more than 1 report, data from the study with the largest sample were included (References 9, 10, 17, 24, 38, 43, 45, 52, 54, 62).

Meta-analysis

Mean values of proton MRS glutamate, glutamine, or Glx concentrations were extracted by one of us (K.M.) and verified by another (A.E.) independently and categorized into the following brain regions of interest: (1) medial frontal cortex, including studies with voxels in the medial prefrontal cortex and in the anterior cingulate cortex since these voxels often spatially overlap; (2) dorsolateral prefrontal cortex (DLPFC); (3) frontal white matter; (4) thalamus; (5) medial temporal lobe (MTL) (including hippocampus); (6) basal ganglia (including caudate, putamen, and globus pallidus); and (7) cerebellum. Only analyses for which at least 3 independent data sets were available were included. When more than 1 clinical group was reported in a single study, the values were treated as independent data sets and the number of healthy volunteers was adjusted by dividing by the number of clinical groups. When data were reported bilaterally, only those for the left hemisphere were included because the left hemisphere was examined in most studies.

The ability of proton MRS to resolve the overlapping resonances of glutamate and glutamine increases with field strength. Previous estimates⁶ of the degree of contamination of glutamate and glutamine signals at different field strengths using optimized sequences indicated that it would be appropriate to include studies reporting glutamate if the data were acquired at field strengths of 3 T or above and studies reporting glutamine at 4 T or above. A secondary analysis included data acquired at all field strengths.

The proton MRS measures of glutamate, glutamine, or Glx were analyzed separately, which was accounted for by applying a Bonferroni-corrected threshold for statistical significance of P < .017. The effect size statistic Hedges g, which incorporates a correction for bias from small sample sizes, was calculated by subtracting the mean glutamate, glutamine, or Glx values reported in cases by the mean value reported in the control group divided by the pooled SD across groups.⁶⁷ If means or SDs were not reported, authors were contacted for this information. A Hedges g value of 0 indicates no difference between cases and controls, negative values indicate lower glutamatergic metabolite levels in cases than controls, and positive values denote higher glutamatergic metabolite levels in cases than controls.

A random-effects, inverse-weighted variance model⁶⁸ was used to calculate the pooled effect size since the studies were expected to display high heterogeneity as different correction methods and clinical samples were used. Study effect size was weighted according to sample size. Heterogeneity was measured using the *I*² value, with higher percentages denoting higher variation across studies in the meta-analysis. The meta-analysis for each brain region was performed using metaanalytical equations entered into Excel (Microsoft Corp) (http: //www.depressiondatabase.org). These equations are identical to the METAN command in Stata (StataCorp LP), which is commonly used in meta-analyses publications. In terms of validation, the method has been used in parallel with Stata in previous meta-analyses⁶⁹ and produced the same results.

Effect sizes were initially calculated for all patients and controls and then for each clinical group (HR, FEP, and schizophrenia) separately. Separate analysis was also performed of patient groups (FEP and schizophrenia) since most HR individuals will not develop psychosis.

Meta-regression

To explore the relationship between glutamate, glutamine, and Glx effect sizes for each study and selected demographics or clinical variables, random effects meta-regressions were conducted using the metareg command in Stata, version 11.2 2009. The variables investigated were age; Positive and Negative Syndrome Scale (PANSS)⁷⁰ total; positive, negative, and general subscale scores; chlorpromazine-equivalent dose; and dura-

tion of illness. In studies that used the Brief Psychiatric Rating Scale, the scores were converted to PANSS scores.⁷¹ When these measures were not reported, study authors were contacted to request the data. Publication bias was examined using the Egger regression test for regions including at least 10 studies⁷² and meta-regressions of study effect size and year of publication. A leave-one-out jackknife sensitivity analysis was conducted for regions with at least 4 studies in which significant between-group differences were found.

Results

The literature search identified 59 studies, with a total of 1686 cases and 1451 controls (PRISMA flow diagram presented in eFigure 1 in the Supplement). The sample sizes ranged from 5 to 84 for cases and 4 to 81 for healthy volunteers (eTable 1 in the Supplement). Two studies^{73,74} reporting multivoxel data were excluded.

Fourteen studies (References 18, 25, 26, 29, 31, 37, 49, 51-53, 56, 57, 62, 75) examined participants at HR for psychosis. Eighteen studies (References 9, 12, 17-19, 23, 24, 28, 30, 32, 33, 45, 48, 57-59, 63, 76) examined patients experiencing a first episode of psychosis (FEP), all with an onset of illness within the last $2\frac{1}{2}$ years. Thirty-six studies (References 8, 10, 11, 13-16, 20-24, 27, 31, 34-36, 38-44, 46, 47, 50, 53-55, 57-61, 77) examined patients with established (chronic) schizophrenia (eResults in the Supplement provide detailed patient information).

Meta-analysis

In the medial frontal cortex, there were no significant findings for glutamate (HR group, 3; FEP group, 3; and schizophrenia group, 11), Glx (HR group, 8; FEP group, 3; and schizophrenia group, 13) or glutamine (HR group, 0; FEP group, 2; and schizophrenia group, 3) (**Table** and eFigure 2 in the **Supplement**). Analysis of each clinical group separately revealed higher Glx concentrations in HR individuals (g = 0.26; 95% CI, 0.05-0.46; P = .01). There were no significant betweengroup differences in glutamate or glutamine levels.

In the frontal white matter, there were no significant effects overall for glutamate (HR group, 1; FEP group, 1; schizophrenia group, 1; and FEP + schizophrenia group, 1) or Glx (HR group, 0; FEP group, 2; and schizophrenia group, 7) in cases compared with controls, and only 1 study⁷⁶ examined glutamine (Table). The schizophrenia group showed elevated Glx levels compared with controls (g = 0.42; 95% CI, 0.18-0.66; P = .001).

In the DLPFC, there were no significant effects for Glx in cases (HR group, 2; FEP group, 2; and schizophrenia group, 8) or in the schizophrenia group. There were insufficient data above 1.5 T for glutamate and at 4 T for glutamine in the DLPFC.

In the basal ganglia, both glutamate (HR group, 1; FEP group, 2; and schizophrenia group, 1; g = 0.63; 95% CI, 0.15-1.11; P = .01) and Glx concentrations (HR group, 4; FEP group, 3; and schizophrenia group, 2; g = 0.39; 95% CI, 0.09-0.70; P = .01) (**Figure 1** and **Figure 2**) were higher in cases than in controls. Subgroup analysis of Glx found elevations in the FEP group (g = 0.66; 95% CI, 0.28-1.03; P < .001) but not in the HR

Brain Region by Metabolite	Group	Studies	Cases	Healthy Controls	Effect Size	Heterogeneity		
					95% CI	P Value	1100000000000000000000000000000000000	P Value
Medial frontal cortex								
Glutamate	All cases (3 HR, 3 FEP, 11 SZ)	17	411	381	-0.14 (-0.34 to 0.06)	.17	39.7	.05
	HR	3	102	80	-0.09 (-0.68 to 0.50)	.77	61.3	.08
	FEP (0 medicated, 3 unmedicated)	3	34	28	-0.09 (-0.46 to 0.29)	.64	12.8	.32
	SZ (11 medicated, 0 unmedicated)	11	242	233	-0.16 (-0.44 to 0.12)	.24	18.7	.04
Glutamine	All cases (0 HR, 2 FEP, 3 SZ)	5	84	78	0.19 (-0.38 to 0.76)	.52	67.7	.02
	SZ (3 medicated, 0 unmedicated)	3	50	50	-0.19 (-0.78 to 0.40)	.53	51.7	.13
Glx	All cases (8 HR, 3 FEP, 13 SZ)	24	487	440	0.12 (-0.02 to 0.27)	.10	13.1	.28
	HR	8	203	183	0.26 (0.05 to 0.46)	.01 ^a	0.0	.51
	FEP (1 medicated, 2 unmedicated)	3	49	49	0.03 (-0.37 to 0.42)	.90	0.0	.37
	SZ (12 medicated, 1 unmedicated)	13	235	208	0.02 (-0.21 to 0.24)	.89	20.5	.24
DLPFC								
Glutamate	All cases (0 HR, 0 FEP, 1 SZ)	1	NA	NA	NA	NA	NA	NA
Glutamine	All cases (0 HR, 0 FEP, 0 SZ)	0	NA	NA	NA	NA	NA	NA
Glx	All cases (2 HR, 2 FEP, 8 SZ)	12	262	184	-0.27 (-0.63 to 0.09)	.15	68.6	<.001
	SZ (7 medicated, 1 unmedicated)	8	172	132	-0.32 (-0.85 to 0.21)	.23	77.6	<.001
Frontal WM								
Glutamate	All cases (1 HR, 1 FEP, 1 SZ, 1 FEP + SZ)	4	57	48	-0.18 (-0.64 to 0.28)	.44	25.6	.26
Glutamine	All cases (0 HR, 1 FEP, 0 SZ)	1	NA	NA	NA	NA	NA	NA
Glx	All cases (0 HR, 2 FEP, 7 SZ)	9	261	135	0.24 (-0.06 to 0.54)	.11	46.2	.06
	SZ (4 medicated, 3 unmedicated)	7	200	110	0.42 (0.18 to 0.66)	.001 ^a	0.0	.58
MTL								
Glutamate	All cases (3 HR, 0 FEP, 3 SZ)	6	83	91	-0.24 (-0.71 to 0.24)	.33	55.3	.05
	HR	3	47	49	-0.34 (-0.86 to 0.17)	.19	30.7	.24
	SZ (2 medicated, 1 mixed)	3	36	43	-0.08 (-1.02 to 0.86)	.87	75.0	.02
Glutamine	All cases (0 HR, 0 FEP, 0 SZ)	0	NA	NA	NA	NA	NA	NA
Glx	All cases (5 HR, 5 FEP, 8 SZ)	18	441	350	0.32 (0.12 to 0.52)	.002ª	42.0	.03
	HR	5	112	78	0.36 (-0.14 to 0.86)	.16	56.7	.06
	FEP (4 medicated, 1 unmedicated)	5	132	94	0.12 (-0.16 to 0.40)	.39	2.2	.39
	SZ (5 medicated, 3 unmedicated)	8	197	179	0.40 (0.08 to 0.71)	.01 ^a	51.3	.04
Basal ganglia								
Glutamate	All cases (1 HR, 2 FEP, 1 SZ)	4	89	83	0.63 (0.15 to 1.11)	.01 ^a	57.2	.07
Glutamine	All cases (0 HR, 0 FEP, 0 SZ)	0	NA	NA	NA	NA	NA	NA
Glx	All cases (4 HR, 3 FEP, 2 SZ)	9	216	182	0.39 (0.09 to 0.70)	.01 ^a	51.5	.04
	HR	4	116	102	0.23 (-0.34 to 0.80)	.43	73.7	.01
	FEP (1 medicated, 2 unmedicated)	3	59	56	0.66 (0.28 to 1.03)	<.001 ^a	0.0	.99
Thalamus								
Glutamate	All cases (1 HR, 2 FEP, 1 SZ)	4	125	103	-0.20 (-0.47 to 0.08)	.16	4.5	.37
Glutamine	All cases (0 HR, 2 FEP, 1 SZ)	3	50	48	0.56 (0.02 to 1.09)	.04	40.5	.19
Glx	All cases (3 HR, 2 FEP, 2 SZ)	7	240	159	0.07 (-0.17 to 0.32)	.56	24.1	.24
	HR	3	120	101	0.16 (-0.39 to 0.71)	.57	71.7	.03
Cerebellum								
Glutamate	All cases (1 HR, 2 FEP, 0 SZ)	3	59	58	0.38 (-0.17 to 0.93)	.17	54.2	.11
Glutamine	All cases (0 HR, 0 FEP, 0 SZ)	0	NA	NA	NA	NA	NA	NA
Glx	All cases (1 HR, 2 FEP, 0 SZ)	3	59	58	0.22 (-0.25 to 0.68)	.36	37.5	.20

Table. Meta-analysis Results Summary for All Patient Groups in All Brain Regions

Abbreviations: DLPFC, dorsolateral prefrontal cortex; FEP, first-episode psychosis; Glx, combined glutamate and glutamine signal; HR, high risk; MTL, medial temporal lobe; NA, not applicable; SZ, chronic schizophrenia; WM, white matter.

^a Results that survived multiple comparisons for each region as glutamate, glutamine, and Glx were investigated.

E4 JAMA Psychiatry Published online June 15, 2016

٨٢٥٦	Hedges a (95% CI)	12 %	Lower in Higher in
Medial frontal cortex		1,70	
Glutamate	-0.14 (-0.34 to 0.06)	40	
Glutamine	0.19 (-0.38 to 0.76)	68	
Glx	0.12 (-0.02 to 0.27)	13	-
DLPFC			-
Glx	-0.27 (-0.63 to 0.09)	69	
Frontal WM			—
Glutamate	-0.18 (-0.64 to 0.28)	26	
Glx	0.24 (-0.06 to 0.54)	46	
MTL			
Glutamate	-0.24 (-0.71 to 0.24)	55	
Glx	0.32 (0.12 to 0.52)	42	
Basal ganglia			
Glutamate	0.63 (0.15 to 1.11)	57	
Glx	0.39 (0.09 to 0.70)	52	
Thalamus			
Glutamate	-0.20 (-0.47 to 0.08)	5	
Glutamine	0.56 (0.02 to 1.09)	41	
Glx	0.07 (-0.17 to 0.32)	24	— — —
Cerebellum			
Glutamate	0.38 (-0.17 to 0.93)	54	
Glx	0.22 (-0.25 to 0.68)	38	
			-1.0 -0.5 0 0.5 1.0 1
			Pooled Effect Size (95% CI)

Figure 1. Summary Effect Sizes for Glutamatergic Differences Between Cases and Controls in Each Brain Region Examined

Negative Hedges g values denote lower glutamatergic metabolite concentrations in cases than controls; positive values denote higher glutamatergic metabolite concentrations in cases than controls. The size of the data markers is proportional to the total number of individuals. DLPFC indicates dorsolateral prefrontal cortex; Glx, combined glutamate and glutamine signal; MTL, medial temporal lobe; and WM, white matter.

group; there were insufficient studies in patients with schizophrenia to determine the results.

In the MTL, Glx was increased in cases compared with controls (HR group, 5; FEP group, 5; and schizophrenia group, 8; g = 0.32; 95% CI, 0.12-0.52; P = .002) (Figures 1 and 2) but not glutamate (HR group, 3; FEP group, 0; and schizophrenia group, 3). Subgroup analysis found significantly higher Glx only in the schizophrenia group (g = 0.40; 95% CI, 0.08-0.71; P = .01) (Figure 2). There were no between-group differences in glutamate levels. The same results were found after excluding patients with 22q11 deletion.¹⁶ There were insufficient data at 4 T for glutamine.

In the thalamus, glutamine concentrations were higher in cases than controls (FEP group, 2; schizophrenia group, 1; g = 0.56; 95% CI, 0.02-1.09; P = .04) (Figure 2). There were no between-group differences in glutamate (HR group, 1; FEP group, 2; and schizophrenia group, 1) or Glx (HR group, 3; FEP group, 2; and schizophrenia group, 2). No significant effects were present for glutamate or Glx in the cerebellum (HR group, 1; FEP group, 2; and schizophrenia group, 0) (Table).

Analysis Limited to Patient Groups

When analysis was limited to patients by excluding the HR group, no additional significant findings were apparent in any region (eTable 2 in the Supplement). Elevated Glx levels in MTL (g = 0.31; 95% CI, 0.09-0.53; P = .007) and basal ganglia (g = 0.57; 95% CI, 0.26-0.88; P < .001), as well as elevated glutamine levels in the thalamus (g = 0.56; 95% CI, 0.02-1.10; P = .04), remained significant; however, glutamate in the basal ganglia (P = .08) was no longer significant.

Meta-analysis Including Studies at Low Field Strength Meta-analyses were repeated including studies that measured glutamate or glutamine at low field strengths (<3 T for glutamate^{12,23,28,32} or <4 T for glutamine (References 10, 12, 13, 16, 23, 32, 35, 55, 62, 77)). Inclusion of these studies did not change the finding of no difference in glutamate levels overall in the medial frontal region, although glutamine levels were significantly elevated in cases (HR group, 1; FEP group, 3; and schizophrenia group, 7; *g* = 0.35; 95% CI, 0.02-0.67; *P* = .04; *I*² = 63%) and in the FEP group (*g* = 0.84; 95% CI, 0.38-1.30;

 $P < .001; I^2 = 0\%$).

In the DLPFC, there were no significant effects for glutamate (HR group, 0; FEP group, 3; and schizophrenia group, 3) or glutamine (HR group, 0; FEP group, 2; schizophrenia group, 4) in cases compared with controls. The schizophrenia group showed elevated glutamine levels in the DLPFC (g = 0.46; 95% CI, 0.06-0.86; P = .02; $I^2 = 25\%$), which was no longer significant when 1 study¹⁶ in patients with a 22q11 deletion was excluded. There were no between-group differences in glutamate levels.

In the MTL, glutamine levels were increased in cases compared with controls (HR group, 1; FEP group, 1; and schizophrenia group, 2; g = 0.41; 95% CI, 0.02-0.80; P = .04; $I^2 = 0$ %). There remained no differences in glutamate for cases or separate clinical groups (HR group, 3; FEP group, 1; and schizophrenia group, 4). The same results were found after excluding patients with a 22q11 deletion.¹⁶

All studies of glutamate in the frontal white matter, basal ganglia, thalamus, and cerebellum were above 1.5 T. All studies of glutamine in the thalamus were at 4 T.

Figure 2. Study Effect Sizes in Brain Regions Showing Significant Glutamatergic Differences Between Cases and Controls

Source	Hedges g (95% CI)			Lower in	Higher in natients	
Basal ganglia: Glx	(patients	patients	
Block et al, ³¹ 2000	0.06					Type study
de la Fuente-Sandoval et al, ¹⁸ 2011	0.40			_	-	FEP
Keshavan et al, ³⁷ 2009	-0.34					Chronic
Tandon et al, ²⁵ 2013	0.88					schizophrenia
de la Fuente-Sandoval et al, ¹⁸ 2011	0.66					
de la Fuente-Sandoval et al, ¹⁷ 2013	0.63					
Goto et al, ¹⁹ 2012	0.69					
Block et al, ³¹ 2000	0.31					
Yamasue et al, ⁵⁰ 2003	0.49			-		
Summary: g = 0.39 (95% CI, 0.09-0.70); I ² = 52%					\diamond	
		2		1 1	0 1	
		-5	-2	Effect Siz	e (95% CI)	2 5
Source	Hedges g (95% CI)			Lower in patients	Higher in patients	
Basal ganglia: glutamate						
de la Fuente-Sandoval et al, ¹⁸ 2011	0.86					
de la Fuente-Sandoval et al, ¹⁸ 2011	0.97				-	
de la Fuente-Sandoval et al, ¹⁷ 2013	0.80					
Tayoshi et al, ⁵⁵ 2009	0.00			_	-	
Summary: g = 0.63 (95% CI, 0.15-1.11); I ² = 57%					\diamond	
		-3	-2	-1 Effect Siz	0 1 e (95% CI)	2 3
	Hedges g			Lower in	Higher in	
Source	(95% CI)			patients	patients	
Medial temporal lobe: Glx						
Capizzano et al, ⁷⁵ 2011	-0.42				<u> </u>	
Capizzano et al, ⁷⁵ 2011	0.22				-	
Stone et al, ⁶² 2009	0.13					
Wood et al, ⁴⁹ 2010	1.52					
Wood et al, ⁴⁹ 2010	0.63					
Galińska et al, ³⁰ 2009	0.14					
Hasan et al, ³³ 2014	-0.11			-	-	
Szulc et al, ²⁴ 2004	0.82					
Wood et al, ⁴⁸ 2008	0.16					
Wood et al, ⁴⁸ 2008	0.38			_		
Chang et al, ¹⁴ 2007	0.92					
da Silva Alves et al, ¹⁶ 2011	1.23					_
Hutcheson et al, ³⁴ 2012	0.00					
Kegeles et al, ³⁶ 2000	0.17					
Kraguljac et al, ³⁸ 2012	-0.10			-	-	
Kraguljac et al, ²¹ 2013	0.61					
Szulc et al, ²⁴ 2004	0.43				-	
Szulc et al, ⁴³ 2011	0.37			-		
Summary: g = 0.32 (95% CI, 0.12-0.52); I ² = 42%					\diamond	
		-3	-2	-1 Effect Siz	0 1 e (95% CI)	2 3
Source	Hedges g (95% CI)			Lower in patients	Higher in patients	
Thalamus: glutamine						
Bustillo et al, ⁷⁶ 2010	-0.08					
Théberge et al, ⁹ 2002	0.70				—	
Théberge et al, ⁵⁴ 2003	0.89					
Summary: g = 0.56 (95% CI, 0.02-1.09); l ² = 41%		_			\bigcirc	
		-3	-2	-1 Effect Siz	0 1 e (95% CI)	2 3

Each data marker represents a study, and the size of the data marker is proportional to the total number of individuals in that study. The summary effect size for each brain region is denoted by a blue diamond. FEP indicates first-episode psychosis; Glx, combined glutamate and glutamine signal; and HR, high risk.

E6 JAMA Psychiatry Published online June 15, 2016

Heterogeneity

Significant heterogeneity was found in numerous groups for all regions (Table), which justifies the use of a randomeffects model to combine the effect sizes. Heterogeneity may result from methodologic differences (eTable 3 in the Supplement). Meta-regression analysis investigated possible sources of heterogeneity.

Meta-regression

The clinical and demographic measures that were available from each study for meta-regression are presented in eTable 1 in the Supplement. In all brain regions, there were no significant correlations between the study effect sizes for glutamate, glutamine, or Glx and the mean PANSS subscale scores, chlorpromazine-equivalent dose, and duration of psychotic illness of patients with FEP or schizophrenia.

In all brain regions, there was no association between the mean age of the patients and the effect size for glutamate, glutamine, or Glx.

Small-Study Bias

Small-study bias, which could reflect publication bias, was evident for reports of Glx in the medial frontal region (Egger test, P = .01) and Glx in the basal ganglia (P = .04). The year of publication was not significantly associated with metabolite reports in any brain region.

Sensitivity Analysis

Leave-1-out sensitivity analysis showed that significant results were generally robust. Significant differences did not remain in 2 of 3 tests for cases of glutamine in the thalamus, indicating an unreliable result.

Discussion

The number of publications reporting proton MRS measures of brain glutamate, glutamine, or Glx in schizophrenia has more than doubled since the last meta-analysis.⁶⁵ In addition to analyzing data from a large number of new studies, we were able to include findings from brain regions precluded from previous meta-analysis in the present study.

We found significant differences in glutamatergic metabolites across several cortical and subcortical regions in cases compared with controls. Although the nature of the findings varied depending on the patient subgroup and brain region, all of the significant findings reflected elevations in glutamatergic metabolites in patients and HR individuals. This finding is consistent with data from animal models of schizophrenia that propose an increase in glutamatergic activity resulting from NMDAR hypofunction.⁶⁶ The finding is also consistent with human studies of NMDAR hypofunction that show increases in both glutamate and glutamine concentrations in the cortex following ketamine administration to healthy volunteers.

The HR, FEP, and schizophrenia groups had higher glutamate and Glx levels in the basal ganglia, higher glutamine levels in the thalamus, and higher Glx levels in the MTL. In contrast, there were generally no significant findings in the DLPFC or cerebellum, and significant findings in the medial frontal cortex and frontal white matter were observed only in specific patient subgroups. Preclinical models propose that glutamatergic overactivity in hippocampal areas drive excessive subcortical dopamine release via polysynaptic glutamatergic projections to the striatum. Likewise, abnormalities in striatal glutamate may influence striatal dopaminergic signaling since glutamate in the basal ganglia modulates tonic dopamine release presynaptically via NMDARs. Finally, the thalamus receives efferent input from the striatum, ⁷⁸ and NMDAR antagonism in the thalamus causes cortical neurotoxic injury via corticothalamic loops.

In all regions except the basal ganglia, the HR, FEP, and schizophrenia groups had significant elevations in the Glx or glutamine signal rather than the glutamate signal, which may partially reflect the greater number of studies reporting on Glx rather than glutamate or glutamine separately. Although the glutamate signal accounts for most (80%-90%) of the Glx signal at field strengths of 1.5 T to 3 T,⁶ it is possible that Glx level elevations could be driven by increases in glutamine rather than glutamate levels. Following neurotransmission, glutamate is converted to glutamine in astrocytes for recycling. Elevations in glutamine levels may thus reflect increases in glutamatergic synaptic activity. The previous meta-analysis²⁷ reported reduced medial frontal glutamate but elevated glutamine levels in schizophrenia that were not detected in the present analysis. This discrepancy may reflect improved methods in more recent studies since more studies acquired data at higher field strengths, correct for voxel cerebrospinal fluid, and specify more conservative thresholds for the acceptability of metabolite fitting (eTable 3 in the Supplement).

There was some evidence that the regional degree of glutamatergic elevation may be sensitive to illness stage. The Glx level elevations in the medial frontal cortex were apparent in HR individuals but not in those with schizophrenia. Similarly, medial frontal glutamine levels were elevated in 2 studies^{9,76} of patients with FEP, but no differences were seen in patients with schizophrenia. Conversely, MTL Glx levels were elevated in individuals with schizophrenia but not in the FEP or HR groups. One interpretation of these findings is that the regional pattern of glutamatergic abnormalities progress with the clinical course of the disorder or show differential responses to antipsychotic treatment. Most HR individuals will not develop psychosis and were not receiving antipsychotic medication; exclusion of this group generally resulted in similar effect sizes. This observation suggests that inclusion of HR groups did not dilute the findings in patients and that the same pattern of glutamatergic-level elevation is apparent in individuals at HR for psychosis. Few studies have directly compared different patient groups^{23,24,57-59} or repeatedly assessed glutamatergic metabolites over long periods.⁷⁹ Interpretation was limited because there were insufficient data to analyze all glutamatergic measures for each patient group separately in every region.

The meta-regression did not find support for the hypothesis that glutamatergic metabolite concentrations in patients vary in association with age, antipsychotic treatment, or symp-

tom severity. The latter may be relevant to the interpretation of cross-sectional studies comparing regional glutamatergic measures in association with antipsychotic treatment response⁸ because this finding suggests that such differences may not simply involve group differences in symptom severity. Detailed information on antipsychotic treatment was available in few data sets, and mean chlorpromazine-equivalent doses may not account for medication adherence and do not discriminate between the effects of different antipsychotic medications. Longitudinal MRS studies have not found effects of antipsychotic treatment on medial frontal glutamatergic measures,^{76,79} although medication effects in the striatum have been reported.¹⁷ One previous study²⁰ reported higher medial frontal Glx in patients not receiving vs receiving medications. The meta-analysis found higher medial frontal glutamine (but not glutamate or Glx) levels in patients with FEP, all of whom were not receiving medications.

One limitation of the present meta-analysis is that, when clinical groups were analyzed separately, the number of studies per group for some regions was small. We conducted the meta-analysis when at least 3 independent data sets were available; however, findings based on a low number of data sets should be considered preliminary and are presented to stimulate further research. Increases in Glx levels in the medial frontal cortex in HR individuals and increases in MTL Glx levels in patients with schizophrenia were reported by relatively large numbers of studies; thus, these investigations represent the most robust of the findings in patient subgroups.

Our HR category included studies of people at increased familial risk for schizophrenia as well as those showing subclinical signs of psychosis since there were too few studies to permit separate meta-analyses of each group. The risk of psychosis differs between these groups, and this heterogeneity may explain why we did not find elevated Glx levels in the basal ganglia and lower glutamate levels in the thalamus that have been reported⁵² in clinical HR individuals. In addition, glutamate may be increased only in HR persons who will later develop psychosis or show poorer outcomes.⁵²

The resonance frequencies of glutamate and glutamine significantly overlap at 1.5 T, whereas glutamate can be largely resolved at 3 T, and field strengths of 4 T or more are needed to measure glutamine accurately.⁶ For glutamine reports, a sufficient number of studies were performed at 4 T only in the thalamus. Given that glutamine may provide a measure of glutamate turnover and our general finding of increased glutamine or Glx rather than glutamate in patients, additional studies at higher field strengths optimized for glutamine resolution should be a priority. Another limitation of this study is that there was much variability in data acquisition and analysis methods (eTable 3 in the Supplement), which will affect data quality. Full reporting of such information in future studies will help to address sources of heterogeneity in subsequent metaanalyses.

Use of proton MRS provides a measure of total concentrations within the voxel studied and thus does not infer the functional significance of the metabolites measured. However, glutamine can act as an indirect measure of neurotransmitter glutamate turnover since 80% of glutamine is used for glutamate neurotransmitter cycling.⁷

Conclusions

This meta-analysis indicates that schizophrenia is associated with glutamatergic-level elevations in several brain regions. These findings further support the idea that pharmacologic compounds that can reduce glutamatergic neurotransmission may have therapeutic potential.

ARTICLE INFORMATION

Submitted for Publication: October 23, 2015; final revision received February 13, 2016; accepted February 15, 2016.

Published Online: June 15, 2016. doi:10.1001/jamapsychiatry.2016.0442.

Author Contributions: Ms Merritt and Dr Egerton contributed equally to the study. Ms Merritt and Dr Egerton had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. *Study concept and design*: Merritt, Egerton, McGuire. *Acauisition, analysis, or interpretation of data*:

Merritt, Egerton, Kempton, Taylor. Drafting of the manuscript: Merritt, Egerton,

McGuire.

Critical revision of the manuscript for important intellectual content: All authors.

Statistical analysis: Merritt, Egerton, Kempton, Taylor.

Obtained funding: McGuire.

Administrative, technical, or material support: McGuire.

Study supervision: Egerton, Taylor, McGuire.

Conflict of Interest Disclosures: Dr Egerton has received consultant fees from Heptares

Therapeutics Ltd. Dr Taylor has received personal fees from Lundbeck, Otsuka, and Sunovion outside the submitted work, and a family member has been an employee of GlaxoSmithKline. Dr McGuire has received consultant fees from Hoffman-LaRoche and Sunovion Pharmaceuticals. No other disclosures were reported.

Funding/Support: This study was supported by a UK Medical Research Council PhD Studentship (Ms Merritt), Medical Research Council grant MR/ LO03988/1 (Dr Egerton), and Medical Research Council fellowship MR/JO08915/1 (Dr Kempton). This study presents independent research funded in part by the National Institute for Health Research (NIHR) Biomedical Research Centre at South London and Maudsley National Health Service (NHS) Foundation Trust and King's College London.

Role of the Funder/Sponsor: The funding organizations had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

Disclaimer: The views expressed are those of the authors and do not necessarily represent those of the NHS, NIHR, or Department of Health.

REFERENCES

1. Olney JW, Farber NB. Glutamate receptor dysfunction and schizophrenia. *Arch Gen Psychiatry*. 1995;52(12):998-1007.

2. Lisman JE, Coyle JT, Green RW, et al. Circuit-based framework for understanding neurotransmitter and risk gene interactions in schizophrenia. *Trends Neurosci*. 2008;31(5):234-242.

3. Javitt DC, Zukin SR. Recent advances in the phencyclidine model of schizophrenia. *Am J Psychiatry*. 1991;148(10):1301-1308.

4. Steiner J, Walter M, Glanz W, et al. Increased prevalence of diverse *N*-methyl-D-aspartate glutamate receptor antibodies in patients with an initial diagnosis of schizophrenia: specific relevance of IgG NR1a antibodies for distinction from *N*-methyl-D-aspartate glutamate receptor encephalitis. *JAMA Psychiatry*. 2013;70(3):271-278.

5. Ripke S, Neale BM, Corvin A, et al; Schizophrenia Working Group of the Psychiatric Genomics Consortium. Biological insights from 108 schizophrenia-associated genetic loci. *Nature*. 2014;511(7510):421-427.

6. Snyder J, Wilman A. Field strength dependence of PRESS timings for simultaneous detection of

glutamate and glutamine from 1.5 to 7T. J Magn Reson. 2010;203(1):66-72.

7. Rothman DLL, De Feyter HMM, de Graaf RAA, Mason GFF, Behar KLL. ¹³C MRS studies of neuroenergetics and neurotransmitter cycling in humans. *NMR Biomed*. 2011;24(8):943-957.

8. Demjaha A, Egerton A, Murray RM, et al. Antipsychotic treatment resistance in schizophrenia associated with elevated glutamate levels but normal dopamine function. *Biol Psychiatry*. 2014;75(5):e11-e13.

9. Théberge J, Bartha R, Drost DJ, et al. Glutamate and glutamine measured with 4.0 T proton MRS in never-treated patients with schizophrenia and healthy volunteers. *Am J Psychiatry*. 2002;159(11): 1944-1946.

10. van Elst LT, Valerius G, Büchert M, et al. Increased prefrontal and hippocampal glutamate concentration in schizophrenia: evidence from a magnetic resonance spectroscopy study. *Biol Psychiatry*. 2005;58(9):724-730.

11. Choe BY, Suh TS, Shinn KS, Lee CW, Lee C, Paik IH. Observation of metabolic changes in chronic schizophrenia after neuroleptic treatment by in vivo hydrogen magnetic resonance spectroscopy. *Invest Radiol.* 1996;31(6):345-352.

12. Bartha R, Williamson PC, Drost DJ, et al. Measurement of glutamate and glutamine in the medial prefrontal cortex of never-treated schizophrenic patients and healthy controls by proton magnetic resonance spectroscopy. *Arch Gen Psychiatry*. 1997;54(10):959-965.

13. Bustillo JR, Chen H, Jones T, et al. Increased glutamine in patients undergoing long-term treatment for schizophrenia: a proton magnetic resonance spectroscopy study at 3 T. *JAMA Psychiatry*. 2014;71(3):265-272.

14. Chang L, Friedman J, Ernst T, Zhong K, Tsopelas ND, Davis K. Brain metabolite abnormalities in the white matter of elderly schizophrenic subjects: implication for glial dysfunction. *Biol Psychiatry*. 2007;62(12):1396-1404.

15. Choe BY, Kim KT, Suh TS, et al. ¹H magnetic resonance spectroscopy characterization of neuronal dysfunction in drug-naive, chronic schizophrenia. *Acad Radiol*. 1994;1(3):211-216.

16. da Silva Alves F, Boot E, Schmitz N, et al. Proton magnetic resonance spectroscopy in 22q11 deletion syndrome. *PLoS One*. 2011;6(6):e21685.

17. de la Fuente-Sandoval C, León-Ortiz P, Azcárraga M, et al. Glutamate levels in the associative striatum before and after 4 weeks of antipsychotic treatment in first-episode psychosis: a longitudinal proton magnetic resonance spectroscopy study. *JAMA Psychiatry*. 2013;70(10): 1057-1066.

18. de la Fuente-Sandoval C, León-Ortiz P, Favila R, et al. Higher levels of glutamate in the associative-striatum of subjects with prodromal symptoms of schizophrenia and patients with first-episode psychosis. *Neuropsychopharmacology*. 2011;36(9):1781-1791.

19. Goto N, Yoshimura R, Kakeda S, et al. Six-month treatment with atypical antipsychotic drugs decreased frontal-lobe levels of glutamate plus glutamine in early-stage first-episode schizophrenia. *Neuropsychiatr Dis Treat*. 2012;8: 119-122.

20. Kegeles LS, Mao X, Stanford AD, et al. Elevated prefrontal cortex γ-aminobutyric acid and glutamate-glutamine levels in schizophrenia measured in vivo with proton magnetic resonance spectroscopy. *Arch Gen Psychiatry*. 2012;69(5): 449-459.

21. Kraguljac NV, White DM, Reid MA, Lahti AC. Increased hippocampal glutamate and volumetric deficits in unmedicated patients with schizophrenia. *JAMA Psychiatry*. 2013;70(12):1294-1302.

22. Ota M, Ishikawa M, Sato N, et al. Glutamatergic changes in the cerebral white matter associated with schizophrenic exacerbation. *Acta Psychiatr Scand*. 2012;126(1):72-78.

23. Stanley JA, Williamson PC, Drost DJ, et al. An in vivo proton magnetic resonance spectroscopy study of schizophrenia patients. *Schizophr Bull.* 1996;22(4):597-609.

24. Szulc A, Galinska B, Tarasów E, et al. Glutamatergic system dysfunction in schizophrenia: a proton magnetic resonance spectroscopy (¹H MRS) study. *Pol J Radiol*. 2004;69(1):33-36.

25. Tandon N, Bolo NR, Sanghavi K, et al. Brain metabolite alterations in young adults at familial high risk for schizophrenia using proton magnetic resonance spectroscopy. *Schizophr Res.* 2013;148(1-3):59-66.

26. Tibbo P, Hanstock C, Valiakalayil A, Allen P. 3-T proton MRS investigation of glutamate and glutamine in adolescents at high genetic risk for schizophrenia. *Am J Psychiatry*. 2004;161(6):1116-1118.

27. Marsman A, Mandl RCW, Klomp DWJ, et al. GABA and glutamate in schizophrenia: a 7 T ¹H-MRS study. *Neuroimage Clin*. 2014;6:398-407.

28. Stanley JA, Vemulapalli M, Nutche J, et al. Reduced *N*-acetyl-aspartate levels in schizophrenia patients with a younger onset age: a single-voxel ¹H spectroscopy study. *Schizophr Res.* 2007;93(1-3): 23-32.

29. Purdon SE, Valiakalayil A, Hanstock CC, Seres P, Tibbo P. Elevated 3T proton MRS glutamate levels associated with poor Continuous Performance Test (CPT-OX) scores and genetic risk for schizophrenia. *Schizophr Res.* 2008;99(1-3):218-224.

30. Galińska B, Szulc A, Tarasów E, et al. Duration of untreated psychosis and proton magnetic resonance spectroscopy (¹H-MRS) findings in first-episode schizophrenia. *Med Sci Monit*. 2009;15 (2):CR82-CR88.

31. Block W, Bayer TA, Tepest R, et al. Decreased frontal lobe ratio of N-acetyl aspartate to choline in familial schizophrenia: a proton magnetic resonance spectroscopy study. *Neurosci Lett.* 2000;289(2):147-151.

32. Bartha R, al-Semaan YM, Williamson PC, et al. A short echo proton magnetic resonance spectroscopy study of the left mesial-temporal lobe in first-onset schizophrenic patients. *Biol Psychiatry*. 1999;45(11):1403-1411.

33. Hasan A, Wobrock T, Falkai P, et al. Hippocampal integrity and neurocognition in first-episode schizophrenia: a multidimensional study. *World J Biol Psychiatry*. 2014;15(3):188-199.

34. Hutcheson NL, Reid MA, White DM, et al. Multimodal analysis of the hippocampus in schizophrenia using proton magnetic resonance spectroscopy and functional magnetic resonance imaging. *Schizophr Res.* 2012;140(1-3):136-142.

35. Jessen F, Fingerhut N, Sprinkart AM, et al. *N*-acetylaspartylglutamate (NAAG) and *N*-acetylaspartate (NAA) in patients with schizophrenia. *Schizophr Bull*. 2013;39(1):197-205.

36. Kegeles LS, Shungu DC, Anjilvel S, et al. Hippocampal pathology in schizophrenia: magnetic resonance imaging and spectroscopy studies. *Psychiatry Res*. 2000;98(3):163-175.

37. Keshavan MS, Dick RM, Diwadkar VA, Montrose DM, Prasad KM, Stanley JA. Striatal metabolic alterations in non-psychotic adolescent offspring at risk for schizophrenia: a (¹)H spectroscopy study. *Schizophr Res.* 2009;115(1):88-93.

38. Kraguljac NV, Reid MA, White DM, den Hollander J, Lahti AC. Regional decoupling of *N*-acetyl-aspartate and glutamate in schizophrenia. *Neuropsychopharmacology*. 2012;37(12):2635-2642.

39. Ohrmann P, Kugel H, Bauer J, et al. Learning potential on the WCST in schizophrenia is related to the neuronal integrity of the anterior cingulate cortex as measured by proton magnetic resonance spectroscopy. *Schizophr Res.* 2008;106(2-3):156-163.

40. Ongür D, Jensen JE, Prescot AP, et al. Abnormal glutamatergic neurotransmission and neuronal-glial interactions in acute mania. *Biol Psychiatry*. 2008;64(8):718-726.

41. Ongür D, Prescot AP, McCarthy J, Cohen BM, Renshaw PF. Elevated gamma-aminobutyric acid levels in chronic schizophrenia. *Biol Psychiatry*. 2010;68(7):667-670.

42. Rowland LM, Spieker EA, Francis A, Barker PB, Carpenter WT, Buchanan RW. White matter alterations in deficit schizophrenia. *Neuropsychopharmacology*. 2009;34(6):1514-1522.

43. Szulc A, Galinska B, Tarasow E, et al. Proton magnetic resonance spectroscopy study of brain metabolite changes after antipsychotic treatment. *Pharmacopsychiatry*. 2011;44(4):148-157.

44. Terpstra M, Vaughan TJ, Ugurbil K, Lim KO, Schulz SC, Gruetter R. Validation of glutathione quantitation from STEAM spectra against edited ¹H NMR spectroscopy at 4T: application to schizophrenia. *MAGMA*. 2005;18(5):276-282.

45. Tibbo PG, Bernier D, Hanstock CC, Seres P, Lakusta B, Purdon SE. 3-T proton magnetic spectroscopy in unmedicated first episode psychosis: a focus on creatine. *Magn Reson Med*. 2013;69(3):613-620.

46. Tunc-Skarka N, Weber-Fahr W, Hoerst M, Meyer-Lindenberg A, Zink M, Ende G. MR spectroscopic evaluation of *N*-acetylaspartate's T2 relaxation time and concentration corroborates white matter abnormalities in schizophrenia. *Neuroimage*. 2009;48(3):525-531.

47. Wood SJ, Yücel M, Wellard RM, et al. Evidence for neuronal dysfunction in the anterior cingulate of patients with schizophrenia: a proton magnetic resonance spectroscopy study at 3 T. *Schizophr Res.* 2007;94(1-3):328-331.

48. Wood SJ, Berger GE, Wellard RM, et al. A ¹H-MRS investigation of the medial temporal lobe in antipsychotic-naïve and early-treated first episode psychosis. *Schizophr Res*. 2008;102(1-3):163-170.

49. Wood SJ, Kennedy D, Phillips LJ, et al. Hippocampal pathology in individuals at ultra-high

risk for psychosis: a multi-modal magnetic resonance study. *Neuroimage*. 2010;52(1):62-68.

50. Yamasue H, Fukui T, Fukuda R, et al. Drug-induced parkinsonism in relation to choline-containing compounds measured by ¹H-MR spectroscopy in putamen of chronically medicated patients with schizophrenia. *Int J Neuropsychopharmacol.* 2003;6(4):353-360.

51. Yoo SY, Yeon S, Choi C-H, et al. Proton magnetic resonance spectroscopy in subjects with high genetic risk of schizophrenia: investigation of anterior cingulate, dorsolateral prefrontal cortex and thalamus. *Schizophr Res.* 2009;111(1-3):86-93.

52. Egerton A, Stone JM, Chaddock CA, et al. Relationship between brain glutamate levels and clinical outcome in individuals at ultra high risk of psychosis. *Neuropsychopharmacology*. 2014;39(12): 2891-2899.

53. Lutkenhoff ES, van Erp TG, Thomas MA, et al. Proton MRS in twin pairs discordant for schizophrenia. *Mol Psychiatry*. 2010;15(3):308-318.

54. Théberge J, Al-Semaan Y, Williamson PC, et al. Glutamate and glutamine in the anterior cingulate and thalamus of medicated patients with chronic schizophrenia and healthy comparison subjects measured with 4.0-T proton MRS. *Am J Psychiatry*. 2003;160(12):2231-2233.

55. Tayoshi S, Sumitani S, Taniguchi K, et al. Metabolite changes and gender differences in schizophrenia using 3-Tesla proton magnetic resonance spectroscopy (¹H-MRS). *Schizophr Res.* 2009;108(1-3):69-77.

56. Bloemen OJN, Gleich T, de Koning MB, et al. Hippocampal glutamate levels and striatal dopamine D(2/3) receptor occupancy in subjects at ultra high risk of psychosis. *Biol Psychiatry*. 2011;70 (1):e1-e2.

57. Natsubori T, Inoue H, Abe O, et al. Reduced frontal glutamate + glutamine and *N*-acetylaspartate levels in patients with chronic schizophrenia but not in those at clinical high risk for psychosis or with first-episode schizophrenia. *Schizophr Bull*. 2014;40(5):1128-1139.

58. Ohrmann P, Siegmund A, Suslow T, et al. Evidence for glutamatergic neuronal dysfunction in

the prefrontal cortex in chronic but not in first-episode patients with schizophrenia: a proton magnetic resonance spectroscopy study. *Schizophr Res.* 2005;73(2-3):153-157.

59. Ohrmann P, Siegmund A, Suslow T, et al. Cognitive impairment and in vivo metabolites in first-episode neuroleptic-naive and chronic medicated schizophrenic patients: a proton magnetic resonance spectroscopy study. *J Psychiatr Res.* 2007;41(8):625-634.

60. Rowland LM, Kontson K, West J, et al. In vivo measurements of glutamate, GABA, and NAAG in schizophrenia. *Schizophr Bull*. 2013;39(5):1096-1104.

61. Stan AD, Ghose S, Zhao C, et al. Magnetic resonance spectroscopy and tissue protein concentrations together suggest lower glutamate signaling in dentate gyrus in schizophrenia. *Mol Psychiatry*. 2015;20(4):433-439.

62. Stone JM, Day F, Tsagaraki H, et al; OASIS. Glutamate dysfunction in people with prodromal symptoms of psychosis: relationship to gray matter volume. *Biol Psychiatry*. 2009;66(6):533-539.

63. Thomas MA, Ke Y, Levitt J, et al. Preliminary study of frontal lobe ¹H MR spectroscopy in childhood-onset schizophrenia. *J Magn Reson Imaging*. 1998;8(4):841-846.

64. Merritt K, McGuire P, Egerton A. Relationship between glutamate dysfunction and symptoms and cognitive function in psychosis. *Front Psychiatry*. 2013;4(11):151.

65. Marsman A, van den Heuvel MP, Klomp DWJ, Kahn RS, Luijten PR, Hulshoff Pol HE. Glutamate in schizophrenia: a focused review and meta-analysis of ¹H-MRS studies. *Schizophr Bull*. 2013;39(1):120-129.

66. Moghaddam B, Adams B, Verma A, Daly D. Activation of glutamatergic neurotransmission by ketamine: a novel step in the pathway from NMDA receptor blockade to dopaminergic and cognitive disruptions associated with the prefrontal cortex. *J Neurosci.* 1997;17(8):2921-2927.

67. Hedges L, Olkin I. *Statistical Methods for Metaanalysis*. Orlando, FL: Academic Press; 1985:369.

68. DerSimonian R, Laird N. Meta-analysis in clinical trials. *Control Clin Trials*. 1986;7(3):177-188.

69. Kempton MJ, Salvador Z, Munafò MR, et al. Structural neuroimaging studies in major depressive disorder: meta-analysis and comparison with bipolar disorder. *Arch Gen Psychiatry*. 2011;68 (7):675-690.

70. Kay SR, Fiszbein A, Opler LA. The Positive and Negative Syndrome Scale (PANSS) for schizophrenia. *Schizophr Bull*. 1987;13(2):261-276.

71. Leucht S, Rothe P, Davis JM, Engel RR. Equipercentile linking of the BPRS and the PANSS. *Eur Neuropsychopharmacol.* 2013;23(8):956-959.

72. Sterne JAC, Sutton AJ, Ioannidis JPA, et al. Recommendations for examining and interpreting funnel plot asymmetry in meta-analyses of randomised controlled trials. *BMJ*. 2011;343:d4002.

73. Bustillo JR, Chen H, Gasparovic C, et al. Glutamate as a marker of cognitive function in schizophrenia: a proton spectroscopic imaging study at 4 Tesla. *Biol Psychiatry*. 2011;69(1):19-27.

74. Seese RR, O'Neill J, Hudkins M, et al. Proton magnetic resonance spectroscopy and thought disorder in childhood schizophrenia. *Schizophr Res.* 2011;133(1-3):82-90.

75. Capizzano AA, Toscano JLN, Ho BC. Magnetic resonance spectroscopy of limbic structures displays metabolite differences in young unaffected relatives of schizophrenia probands. *Schizophr Res.* 2011;131(1-3):4-10.

76. Bustillo JR, Rowland LM, Mullins P, et al. ¹H-MRS at 4 tesla in minimally treated early schizophrenia. *Mol Psychiatry*. 2010;15(6):629-636.

77. Shirayama Y, Obata T, Matsuzawa D, et al. Specific metabolites in the medial prefrontal cortex are associated with the neurocognitive deficits in schizophrenia: a preliminary study. *Neuroimage*. 2010;49(3):2783-2790.

78. Lodge DJ, Grace AA. Hippocampal dysregulation of dopamine system function and the pathophysiology of schizophrenia. *Trends Pharmacol Sci.* 2011;32(9):507-513.

79. Théberge J, Williamson KE, Aoyama N, et al. Longitudinal grey-matter and glutamatergic losses in first-episode schizophrenia. *Br J Psychiatry*. 2007;191:325-334.