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DOI: 10.1007/s00431-016-2780-0

Document Version Peer reviewed version

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Citation for published version (APA):

Drysdale, S. B., Alcazar, M., Wilson, T., Smith, M., Zuckerman, M., Hodemaekers, H. M., Janssen, R., Bont, L., Johnston, S. L., & Greenough, A. (2016). Functional and genetic predisposition to rhinovirus lower respiratory tract infections in prematurely born infants. *European Journal of Pediatrics*, *175*(12), 1943-1949. https://doi.org/10.1007/s00431-016-2780-0

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1	Functional and genetic predisposition to rhinovirus lower respiratory tract				
2	infections in prematurely born infants				
3					
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35 Term born infants are predisposed to human rhinovirus (HRV) lower respiratory 36 tract infections (LRTI) by reduced neonatal lung function and genetic 37 susceptibility. Our aim was to investigate whether prematurely born infants 38 were similarly predisposed to HRV LRTIs or any other viral LRTIs. Infants 39 born less than 36 weeks of gestational age were recruited. Prior to 40 neonatal/maternity unit discharge, lung function (functional residual capacity by 41 helium gas dilution and multiple breath washout, lung clearance index and 42 compliance (Crs) and resistance (Rrs) of the respiratory system) was assessed and 43 DNA samples assessed for eight single nucleotide polymorphisms (SNPs) in 44 seven genes: ADAM33, IL10, MMP16 NFkB1A,SFTPC, VDR and NOS2A. 45 Infants were prospectively followed until one year corrected age. 46 Nasopharyngeal aspirates (NPAs) were sent whenever an infant developed a 47 LRTI and tested for 13 viruses. One hundred and thirty-nine infants were 48 included in the analysis. Infants who developed HRV LRTIs had reduced Crs 49 (1.6 versus 1.2 mL/cmH₂O/kg, p=0.044) at 36 weeks postmenstrual age. A SNP 50 in the gene coding for the vitamin D receptor was associated with the 51 development of HRV LRTIs and any viral LRTIs (p=0.02).

52 *Conclusion* Prematurely born infants may have both a functional and genetic
53 predisposition to HRV LRTIs.

54

55 Key words: human rhinovirus; single nucleotide polymorphisms; compliance
56 and resistance of the respiratory system; functional residual capacity

58 List of abbreviations

60	BPD	Bronchopulmonary dysplasia
61	Crs	Compliance of the respiratory system
62	FRC _{HE}	Functional residual capacity (by helium gas)
63	HRV	Rhinovirus
64	LCI	Lung clearance index
65	LRTI	Lower respiratory tract infection
66	NPA	Nasopharyngeal aspirates
67	PCR	Polymerase chain reaction
68	PMA	Postmenstrual age
69	R _{rs}	Resistance of the respiratory system
70	RSV	Respiratory syncytial virus
71	SNP	Single nucleotide polymorphisms
72	VDR	Vitamin D receptor
73		

74 AUTHORS SUMMARY

75

76 What is known

- Term born infants are predisposed to rhinovirus lower respiratory tract
 (HRV LRTIs) infection by reduced neonatal lung function.
- Term born infants requiring hospitalisation due to HRV bronchiolitis were
 more likely to have single nucleotide polymorphism (SNP) in the IL-10
 gene.
- 82

83 What is new

- Prematurely born infants who developed a HRV LRTI had lower C_{rs} before
 maternity unit discharge.
- A SNP in the gene coding for the vitamin D receptor was associated with
 the development of HRV LRTIs and overall respiratory viral LRTIs in
 prematurely born infants.

92 Human rhinoviruses (HRV) are the most common cause of respiratory tract 93 infection in infants, with almost all infants developing at least one HRV 94 infection in the first year after birth [14, 23]. Both term and prematurely born 95 infants are susceptible to developing LRTIs caused by HRV [3, 11, 13, 21,24]. 96 Some term born infants may be predisposed to wheezy HRV LRTIs by reduced 97 neonatal lung function [22]. The adjusted risk of developing a wheezy HRV 98 LRTI in the first year of life was 1.8 times higher for each standard deviation 99 increase of airway resistance (R_{rs}) measured at two months of age [22]. In 100 addition, some term born infants may be genetically predisposed to HRV 101 infection. Infants developing HRV bronchiolitis requiring hospitalisation at less 102 than six months of age were more likely to have a single nucleotide 103 polymorphism (SNP) in the IL-10 gene compared to unselected blood donors 104 [9]. Other SNPs in genes coding for IL-18, TLR4 and IFN- γ did not confer 105 susceptibility to hospitalisation for HRV infection [9]. The aim of this study 106 was to determine whether prematurely born infants were functionally and genetically predisposed to HRV LRTIs. An additional aim was to determine 107 108 whether prematurely born infants were functionally and genetically predisposed 109 overall to respiratory viral LRTIs.

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- 113

114 MATERIALS AND METHODS

115

Analysis was undertaken of the results of infants entered into a study investigating the risk factors for viral LRTIs in prematurely born infants [7]. Infants were eligible for recruitment into the study if they were born prior to the onset of the RSV season (1st October to 31st March in the UK) in 2008 or 2009 and were born at less than 36 weeks of completed gestation. Ethical approval was obtained from King's College Hospital NHS Foundation Trust Research Ethics Committee.

123

124 Prior to neonatal/maternity unit discharge either blood or buccal swabs were 125 obtained from infants and stored at -20°C until tested. The samples were then 126 sent on dry ice to the National Institute for Public Health and the Environment 127 (RIVM) in Bilthoven, The Netherlands for testing. DNA was isolated from the 128 blood samples or buccal swabs and then stored at -20°C at the RIVM until 129 analysed [6]. Eight single nucleotide polymorphisms (SNPs) were chosen to be 130 tested. The chosen SNPs had previously been associated with HRV infection in 131 term born infants less than six months old [9]. Nuclear factor- κ -B activity has 132 been associated with steroid resistant airway epithelium in HRV infection in 133 vitro and thus the SNP NFkB1A rs2233409 was also included [15]. In addition, 134 we have studied SNPs associated with reduced lung function in previously 135 healthy children at three and five years of age [20], RSV infection in 136 prematurely born infants [19], prematurity [10] or bronchopulmonary dysplasia 137 We also included SNPs associated with RSV infection in (BPD) [8].

prematurely born infants as they may be associated with other viral causes ofbronchiolitis (i.e. HRV) in prematurely born infants.

140

141 The extracted DNA samples were diluted with TE Buffer to 7 ng/ μ L and sent to 142 KBioscience (Herts, UK) for genotyping. Six SNPs (ADAM33 rs2787094, 143 IL10 rs1800872, MMP16 rs2664349, MMP16 rs2664352, NFkB1A rs2233409 144 and SFTPC rs1124) were tested at KBioscience with the KASPar technology 145 and two further SNPs (vitamin D receptor [VDR rs10735810] and nitric oxide 146 synthase 2A [NOS2A rs1060826]) were tested at the RIVM in the Netherlands. 147 Genotyping of VDR rs10735810 was performed by a custom TaqMan SNP 148 genotyping assay (Applied Biosystems, Carlsbad, USA) and genotyping of 149 NOS2A rs1060826 was performed by using TaqMan SNP genotyping assay 150 C 9458082 10. Genotyping of both SNPs tested at the RIVM was carried out 151 on a 7500 Fast Real-Time PCR system (Applied Biosystems) as previously 152 described [6]. The genotype distributions of the eight SNPs were in Hardy-153 Weinberg equilibrium [6].

154

Lung function was assessed at 36 weeks postmenstrual age (PMA) whilst infants were still inpatients on the neonatal or maternity unit. Infants were not sedated or ventilated during lung function testing. Lung volume was assessed by measurement of functional residual capacity (FRC_{He}), using a commercially available helium gas dilution system (EBS 2615, Equilibrated Bio Systems, New York) as previously described [5]. Lung volume was also assessed by the measurement of FRC (FBC_{MBW}) using the commercially available open circuit

162 multiple breath wash-in/out system (Exhalyzer D, Ecomedics, Duernten, 163 Switzerland) and using sulphur hexafluoride as a tracer gas as previously 164 described [6]. The MBW technique also measures ventilation inhomogeneity 165 (VI), measured as lung clearance index (LCI) as previously described [6]. 166 Compliance (C_{rs}) and resistance (R_{rs}) of the respiratory system were measured 167 using the single breath occlusion technique as previously described [7].

168

169 Following neonatal or maternity unit discharge, infants were followed 170 prospectively until one year corrected age. Whenever an infant developed an 171 LRTI, regardless of whether the child remained at home or required 172 hospitalisation a nasopharyngeal aspirate (NPA) was taken. An infant was 173 diagnosed with a viral LRTI if they had coryzal symptoms together with a 174 respiratory examination demonstrating either a raised respiratory rate for their 175 age, crackles or wheeze or respiratory distress (e.g. tracheal tug or intercostal or 176 subcostal recession). NPAs were tested for 11 viruses (rhinovirus, RSV A and 177 B, human metapneumovirus, influenza A and B, parainfluenza 1-3, enterovirus and parechovirus) using real time reverse transcription polymerase chain 178 179 reaction (PCR) and for adenovirus and bocavirus using real time PCR as 180 previously described [5].

181

182 The neonatal notes were reviewed to document demographic and clinical data 183 and to document the duration of the infants' admission on the neonatal and/or 184 maternity unit. Antenatal, perinatal and postnatal data collected included that on 185 maternal infections, antenatal steroid use, use of surfactant, duration of

186 respiratory support, development of bronchopulmonary dysplasia (BPD),

187 postnatal infant sepsis, breast/formula feeding and use of palivizumab [7].

188

189 Statistical Analysis

190

191 The infants were divided into two groups depending on their HRV LRTI status. 192 The "no LRTI group" consisted of infants who did not develop a viral LRTI 193 throughout the study period and the "HRV LRTI group" consisted of infants 194 who developed at least one HRV LRTI during the study period. The infants in 195 the HRV LRTI group may also have had other viral LRTIs. We also undertook 196 a subsidiary analysis of all infants who had LRTIs with NPAs positive for 197 respiratory viruses and compared their outcomes to infants who had no LRTI. 198 Infants who had LRTIs but no virus was detected from the NPA were excluded 199 from the analysis.

200

201 Data were tested for normality using the Shapiro-Wilk test. Data were analysed 202 using either the independent T-test, the Mann-Whitney U test, the Chi-squared 203 test or the Fisher's exact test as appropriate. A multivariable regression model 204 was used to examine whether lung function at 36 weeks PMA was a predictor of 205 HRV LRTI or respiratory viral LRTIs, independent of other variables which in 206 the univariate analysis were significant at $p \le 0.1$. Statistical analysis was 207 carried out with IBM SPSS Statistics (version 19, New York, USA).

208

A sample size of 28 infants in each group allowed the detection of a difference in the premorbid lung function results equivalent to one standard deviation, with 90% power and two-sided 5% significance. A previous study [2], demonstrated a significant difference in lung function (R_{rs}) equivalent to one standard deviation between the groups.

217

218 **RESULTS**

219

220 During the study period two hundred and fifty one infants met the eligibility 221 criteria for recruitment into the study (Figure 1). One hundred and thirty-nine 222 infants were included in the overall analysis. Their median gestational age (GA) 223 was 34 (range 23-35) weeks and median birth weight 1904 (range 610-3610) g. 224 Four infants received palivizumab of which one was admitted to hospital due to 225 an RSV LRTI. There were significant differences when comparing the 226 demographic data of the HRV group and the no LRTI group. The HRV group 227 were more immature and lighter at birth, more received surfactant, had a longer 228 duration of supplemental oxygen, developed BPD, received palivizumab, 229 developed postnatal sepsis and had a longer duration of neonatal/maternity unit 230 stay (Table 1). Comparison of those infants who developed any respiratory 231 virus LRTI compared to no LRTI is shown in Appendix Table 1. Some infants 232 developed more than one viral LRTI or had more than one virus detected from 233 an NPA during a HRV LRTI (Table 2).

Eight (25%) infants in the HRV LRTI group required hospitalisation (six due to a viral LRTI [two HRV]), one due to a minor head injury and one due to gastroenteritis. Nine (12%) infants in the no LRTI group required hospitalisation (all due to non-respiratory causes).

238

239 The HRV LRTI group were more immature (36 weeks versus 37 weeks PMA, 240 p=0.031) and of lower weight (1908 versus 2113 g, p=0.007) when their lung 241 function was measured. The HRV LRTI group had a smaller FRC_{He} uncorrected for weight (p=0.004), although this was no longer significantly 242 243 different after correcting for weight (p=0.13), a smaller FRC_{MBW} uncorrected for 244 weight (p=0.001) which remained significantly different when corrected for 245 weight (p=0.042), a lower C_{rs} uncorrected for weight (p=0.001) which remained 246 significantly different when corrected for weight (p=0.005) and a higher R_{rs} 247 (p=0.028) (Table 3). Multivariate analysis revealed that after correcting for 248 significant differences in the demographic data the only difference in lung 249 function between the groups that remained significant was in the C_{rs} corrected 250 for weight (Table 3). There were no significant differences in the lung function 251 results of the infants who had any respiratory virus LRTI compared to those who 252 had no LRTI after correcting for differences in their demographics (Appendix 253 Table 2).

254

There were no significant differences at the genotype level in any of the SNPs between the HRV LRTI and no LRTI groups (data not shown). There was a significant difference in the SNP (rs10735810) in the VDR gene at the allele

level. Infants with the G allele were significantly more likely (OR 2.07 (95% CI
[0.98-3.13], p=0.047) to develop HRV LRTIs than those with the A allele
(Table 4). Similarly there was a significant difference in the SNP in the VDR
gene at the allele level between infants who did and did not develop a
respiratory viral LRTI (p=0.02) (Appendix Table 3).

263

264

265 **DISCUSSION**

266

We have demonstrated that prematurely born infants who developed HRV LRTIs had reduced premorbid lung function, that is they had significantly lower Crs than those who did not develop an HRV LRTI. In addition, a SNP in the G allele of the vitamin D receptor gene was associated with an increased risk of developing HRV LRTIs and respiratory viral LRTIs overall.

272

273 Term born infants have been shown to have reduced lung function prior to 274 developing HRV LRTIs [22]. Although, in that study overall there were no 275 significant differences in lung function between the infants who did and did not 276 develop an HRV infection, those infants who wheezed with an HRV infection 277 had significantly reduced lung function (Crs and Rrs) compared with those 278 infants who had HRV infections but did not wheeze [22]. In this study, initial 279 analysis demonstrated several differences in lung function between infants who 280 did and did not develop an HRV LRTI. After adjusting for differences in the 281 demographic data, however, the only significant difference that remained was in

 C_{rs} corrected for weight. A possible explanation is that infants with a low C_{rs} may have less lung distensibility leading to poorer clearance of respiratory secretions. In term born infants, a reduced C_{rs} was associated with an increased susceptibility to hospitalisation with RSV LRTIs as well as post RSV bronchiolitis wheezing [25]. Although there were significant differences in lung function between the all virus LRTI and the no LRTI group, these disappeared after adjusting for confounding factors.

289

290 Vitamin D deficiency has been associated with an increased risk of developing 291 viral LRTIs in infants, in particular RSV LRTIs [1, 18]. In addition, SNPs in the 292 VDR gene have been associated with severe RSV bronchiolitis and other viral 293 LRTIs in infants [12, 17] but no previous study has investigated the role of the 294 VDR in HRV infection. In this study a SNP in the gene coding for VDR was 295 associated with the development of HRV LRTIs in prematurely born infants 296 and, in addition, in infants overall with respiratory viral LRTIs. Vitamin D has 297 an important role in innate immunity [4] it is thus plausible that defects in the 298 VDR will increase an infant's susceptibility to HRV infections. Only one 299 previous study [9] has investigated the genetic susceptibility of infants to HRV 300 infection. In that study [9], term born infants with the A allele of a SNP (at -301 1082) in the gene coding for IL-10 were more likely to be hospitalised for HRV 302 bronchiolitis at less than six months of age than those with the G allele. In this 303 study a different SNP (rs10735810) in the IL-10 gene was not associated with 304 HRV LRTI. The difference in those results suggest that genetic susceptibility to 305 HRV infection is different in term and prematurely born infants. The other SNPs tested in this study have been associated with severe RSV infection, prematurity or BPD in prematurely born infants but not HRV infection and did not appear to influence the development of HRV LRTIs, suggesting they may not have a role in prematurely born infants' susceptibility to HRV LRTI. We also did not find any significant association between the SNPs tested and respiratory viral infections overall. The numbers of infants with each viral infection precluded subanalysis at that level.

313

314 The current study has strengths and some weaknesses. A large cohort of 315 prematurely born infants from a variety of ethnic backgrounds was prospectively 316 Lung function was assessed before neonatal or maternity unit followed. 317 discharge, that is prior to any of the infants being infected with any viral 318 infection. The wide range of ethnicities in the study may have affected the 319 results, as genotype differences in various ethnic groups may increase the 320 likelihood of associations occurring by chance [16]. No correction was made for 321 multiple testing of the genetics data; it is, therefore, possible the significant 322 differences we demonstrate with respect to VDR could be attributable to chance. 323 Nevertheless, we demonstrate a significant relationship not only with HRV 324 LRTIs but any respiratory viral LRTIs. Although infants born at less than 36 325 weeks GA were eligible for entry into the study most of the infants recruited 326 were born moderately prematurely (median gestational age 34 weeks) and thus 327 the results of this study may not be generalisable to all infants born extremely 328 prematurely.

In conclusion, prematurely born infants may be predisposed to HRV LRTIs by both reduced premorbid lung function and genetic susceptibility. A SNP in the gene coding for VDR was associated with the development overall of respiratory viral LRTIs.

334 **Compliance with ethical standards**

335

336 Funding: SBD was supported by the National Institute for Health Research 337 (NIHR) Biomedical Research Centre at Guy's and St Thomas' NHS Foundation 338 Trust / King's College London. The research nurses (MA, TW) were supported 339 by Abbott Laboratories. SLJ is supported by the Asthma UK Clinical Chair 340 CH11SJ, and ERC FP7 Advanced grant 233015. SLJ and AG are MRC and 341 Asthma UK Centre in Allergic Mechanisms of Asthma Investigators, supported 342 by MRC Centre Grant G1000758. SLJ is an NIHR Senior Investigator. 343 **Conflict of interest:** There is no conflict of interest to declare from all authors. 344 Compliance and ethical standards: All procedures performed in studies 345 involving human participants were in accordance with the ethical standards of 346 the institutional and/or national research committee and with the 1964 Helsinki 347 declaration and its later amendments or comparable ethical standards. 348 Informed consent: Infants whose parents gave informed written consent were 349 recruited. 350 Contributor statement: AG, SLJ and LB designed the study. MS and MZ 351 undertook the virological analyses. SBD undertook the lung function 352 assessments. MA, TW and SBD were responsible for the follow up of the 353 patients. SBD, HMH, RJ and LB undertook the genetic analyses. All authors 354 were involved in the preparation of the manuscript.

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and post-respiratory syncytial virus wheeze in term infants. Eur
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460 Table 1: Demographic data

461 Data are shown as median (range) or n (%)

	No LRTI	HRV LRTI	P value
Ν	74	32	
Gestational age (weeks) Birth weight (g)	34 (25-35) 2070 (895-3610)	33 (23-35) 1558 (610-2546)	0.03 <0.001
Males	49 (53%)	14 (44%)	0.53
Ethnicity:			
Caucasian Black African Black Caribbean Asian Hispanic Mixed ethnicity	23 (31%) 17 (23%) 15 (20%) 3 (4%) 1 (1%) 15 (21%)	8 (25%) 9 (28%) 6 (19%) 2 (6%) 2 (6%) 5 (16%)	$\begin{array}{c} 0.53 \\ 0.57 \\ 0.75 \\ 0.62 \\ 0.16 \\ 0.49 \end{array}$
Antenatal smoking	11 (15%)	6 (19%)	0.78
Antenatal steroids	42 (57%)	24 (75%)	0.09
Maternal sepsis	14 (19%)	4 (13%)	0.58
Surfactant	11 (15%)	13 (41%)	0.006
Duration of ventilation	0 (0-82)	1 (0-103)	0.10
Duration of supplemental	0 (0-118)	1.5 (0-458)	0.041
Bronchopulmonary	4 (5%)	8 (25%)	0.006
Breastfed	62 (84%)	23 (72%)	0.19
Postnatal sepsis	20 (27%)	17 (53%)	0.014
Parental atopy	52 (70%)	20 (63%)	0.50
Number of siblings	0 (0-5)	0 (0-5)	0.64
Palivizumab	0 (0%)	4 (13%)	0.007
Neonatal/maternity unit stay (days)	16 (2-118)	28 (5-276)	0.003

- 463 Table 2: Number of viruses detected by real-time PCR in the HRV LRTI group
- 464 Data shown are the number of times a virus was detected. Some infants had
- 465 more than one viral LRTI.

	Viruses detected
Rhinovirus	40
RSV A	7
RSV B	7
Adenovirus	11
Human metapneumovirus	3
Influenza A	1
Influenza B	3
Parainfluenza 1	3
Parainfluenza 2	0
Parainfluenza 3	4
Enterovirus	14
Parechovirus	3
Bocavirus	4
Dual infections	24
Triple infections	4

470 Table 3: Lung function results

471 Data are shown as median (range).

472

	No LRTI	HRV LRTI	P value*	P value after correcting for confounding factors**
Ν	74	32		luctors
Postmenstrual age (PMA) (weeks)	36 (34-42)	37 (35-43)	0.031	N/A
Weight (g)	2113	1908	0.007	N/A
	(1362-3360)	(1200-2640)	0.004	0.55
FRC _{He} (mL)	55 (30-99)	49 (10-68)	0.004	0.55
FRC _{He} (mL/kg)	25 (17-34)	24 (8-35)	0.13	0.59
FRC _{MBW} (mL)	57 (30-91)	44 (13-64)	0.001	0.16
FRC _{MBW} (mL/kg)	27 (16-35)	23 (10-34)	0.042	0.10
LCI	9.8 (7.0-13.6)	10.3 (7.7-13.8)	0.066	0.60
C _{rs} (mL/cmH ₂ O)	3.2 (1.7-5.8)	2.5 (1.0-5.4)	0.001	0.21
C _{rs} (mL/cmH ₂ O/kg)	1.6 (0.7-2.3)	1.2 (0.4-2.1)	0.005	0.044
R _{rs} (cmH ₂ O/L/s)	69 (48-144)	76 (49-199)	0.028	0.85

⁴⁷³

474 *Univariate analysis comparing the two groups

475 **Multivariate analysis adjusting for confounding factors

476 Table 4: Associations at the allele levels by HRV status

477 Data are shown as n (%).

		Asso	ociation at the	allele leve	el
Gene	Allele	HRV LRTI	No LRTI	Р	OR (95% CI)
Vitamin D receptor	А	13 (22%)	51 (36%)	0.047	0.48 (0.22-1.03)
(VDR)	G	47 (78%)	89 (64%)		2.07 (0.98-3.13)
Nitric oxide synthase	Т	43 (72%)	97 (69%)	0.87	1.12 (0.55-2.31)
type 2A (NOS2A)	С	17 (28%)	43 (31%)		0.89 (0.43-1.82)
A disintegrin and	С	17 (28%)	43 (31%)	0.87	0.89 (0.43-1.82)
metalloprotease 33	G	43 (72%)	97 (69%)		1.12 (0.55-2.31)
(ADAM33)					
NFKB1A	С	50 (86%)	108 (83%)	0.83	1.21 (0.50-2.90)
	Т	8 (14%)	22 (17%)		0.82 (0.34-1.99)
IL10	А	19 (32%)	45 (34%)	0.87	0.90 (0.88-1.81)
	С	41 (68%)	87 (66%)		1.11 (0.55-2.26)
Pulmonary	А	12 (20%)	33 (24%)	0.71	0.81 (0.36-1.80)
surfactant protein C (SFTPC)	G	48 (80%)	107 (76%)		1.23 (0.56-2.78)
Matrix	С	31 (52%)	69 (50%)	0.88	1.07 (0.56-2.05)
metalloproteinase-16 (MMP16) rs2664352	Т	29 (48%)	69 (50%)		0.94 (0.49-1.79)
MMP16 rs2664349	G	39 (65%)	86 (60%)	0.75	1.12 (0.57-2.22)
	А	21 (35%)	52 (40%)		0.89 (0.45-1.76)

478 FIGURE LEGEND

479

480 Figure 1: Flow diagram of eligibility

482 APPENDIX

485 Table 1: Demographic data

487 Data are shown as median (range) or n (%)

	No LRTI	All virus LRTI	P value
Ν	74	65	
Gestational age (weeks)	34 (25-35)	33 (23-35)	0.11
Birth weight (g)	2070 (895-3610)	2000 (1440-3154)	0.001
Males	39 (53%)	37 (57%)	0.73
Ethnicity:			
Caucasian	23 (31%)	14 (22%)	0.25
Black African	17 (23%)	19 (29%)	0.44
Black Caribbean	15 (20%)	16 (25%)	0.55
Asian	3 (4%)	3 (5%)	>0.99
Hispanic	1 (1%)	2 (3%)	0.60
Mixed ethnicity	15 (21%)	11 (14%)	0.67
Antenatal smoking	11 (15%)	11 (17%)	0.82
Antenatal steroids	42 (57%)	52 (80%)	0.004
Maternal sepsis	14 (19%)	16 (25%)	0.54
Surfactant	11 (15%)	20 (31%)	0.04
Duration of ventilation	0 (0-82)	0.5 (0-103)	0.12
(days)			
Duration of supplemental	0 (0-118)	1 (0-458)	0.06
oxygen (days)			
Bronchopulmonary	4 (5%)	11 (17%)	0.052
dysplasia			
Breastfed	62 (84%)	58 (89%)	>0.99
Postnatal sepsis	20 (27%)	23 (35%)	0.27
Parental atopy	52 (70%)	42 (65%)	0.59
Number of siblings	0 (0-5)	1 (0-5)	0.78
Palivizumab	0 (0%)	5 (8%)	0.02
Neonatal/maternity unit	16 (2-118)	25 (3-276)	0.001
stay (days)			

490	Table 2: Lung function results
491	-

Data are shown as median (range).

103					
-75		No LRTI	All virus LRTI	P value*	P value after correcting for confounding factors**
	Ν	74	65		
	Postmenstrual age (PMA) (weeks)	36 (34-42)	36 (34-43)		N/A
	Weight (g)	2113 (1362-3360)	1000 (1440-3154)		N/A
	FRC_{He} (mL)	55 (30-99)	51 (22-99)	0.008	0.98
	FRC _{He} (mL/kg)	25 (17-34)	24 (14-35)	0.27	0.94
	FRC _{MBW} (mL)	57 (30-91)	53 (16-111)	0.02	0.28
	FRC _{MBW} (mL/kg)	27 (16-35)	26 (10-42)	0.21	0.25
	LCI	9.8 (7.0-13.6)	9.8 (6.0-14.1)	0.18	0.56
	C _{rs} (mL/cmH ₂ O)	3.2 (1.7-5.8)	3.1 (1.0-6.7)	0.004	0.96
	C _{rs} (mL/cmH ₂ O/kg)	1.6 (0.7-2.3)	1.3 (0.4-2.4)	0.018	0.55
	R_{rs} (cmH ₂ O/L/s)	69 (48-144)	77 (43-199)	0.03	0.50

*Univariate analysis comparing the two groups **Multivariate analysis adjusting for confounding factors

497 Table 3: Associations at the allele level by HRV status

- 499 Data are shown as n (%).

				Associat	ion at the allele level
Gene	Allele	All virus LRTI	No LRTI	Р	OR (95% CI)
Vitamin D receptor	А	28 (23%)	51 (36%)	0.02	0.52 (0.30-0.90)
(VDR)	G	94 (77%)	89 (64%)		1.92 (1.12-3.32)
Nitric oxide	Т	88 (72%)	97 (69%)	0.68	1.15 (0.67-1.96)
synthase type 2A	С	34 (28%)	43 (31%)		0.87 (0.51-1.49)
(NOS2A)					
A disintegrin and	С	41 (34%)	43 (31%)	>0.99	1.01 (0.59-1.77)
metalloprotease 33	G	81 (56%)	97 (69%)		0.99 (0.56-1.72)
(ADAM33)					
NFĸB1A	С	99 (84%)	108 (83%)	0.87	1.06 (0.54-2.08)
	Т	19 (16%)	22 (17%)		0.94 (0.48-1.84)
IL10	А	42 (34%)	45 (34%)	>0.99	1.02 (0.60-1.71)
	С	80 (66%)	87 (66%)		0.99 (0.59-1.66)
Pulmonary	А	19 (16%)	33 (24%)	0.12	0.60 (0.32-1.11)
surfactant protein	G	103 (84%)	107 (76%)		1.67 (0.89-3.13)
C (SFTPC)					
Matrix	С	61 (50%)	69 (50%)	>0.99	1.0 (0.61-1.62)
metalloproteinase-	Т	61 (50%)	69 (50%)		1.0 (0.61-1.62)
16					
(MMP16)					
rs2664352					
MMP16	G	75 (64%)	86 (60%)	0.84	0.95 (0.57-1.57)
rs2664349	А	43 (36%)	52 (40%)		1.05 (0.63-1.75)